# ZOOTAXA 

## 4707

# The genus Pseudolycoriella Menzel \& Mohrig, 1998 (Diptera, Sciaridae) in New Zealand 

## ARNE KÖHLER

Senckenberg Deutsches Entomologisches Institut, Eberswalder Straße 90, D-15374 Müncheberg, Germany. E-mail: arne.koehler@senckenberg.de


Magnolia Press
Auckland, New Zealand

ARNE KÖHLER
The genus Pseudolycoriella Menzel \& Mohrig, 1998 (Diptera, Sciaridae) in New Zealand (Zootaxa 4707)

69 pp.; 30 cm .
12 Dec. 2019
ISBN 978-1-77670-827-7 (paperback)
ISBN 978-1-77670-828-4 (Online edition)

FIRST PUBLISHED IN 2019 BY
Magnolia Press
P.O. Box 41-383

Auckland 1346
New Zealand
e-mail: magnolia@mapress.com
https://www.mapress.com/j/zt
© 2019 Magnolia Press

| ISSN 1175-5326 | (Print edition) |
| :--- | :--- |
| ISSN 1175-5334 | (Online edition) |

## Table of Contents

Abstract .....  3
Introduction. ..... 4
Materials and methods .....  5
Key to the males of New Zealand Pseudolycoriella species ..... 6
Species account. ..... 14
Pseudolycoriella cavatica (Skuse, 1888) ..... 14
Pseudolycoriella kaikoura sp. n. ..... 14
Pseudolycoriella sp. I ..... 16
Pseudolycoriella zealandica complex ..... 16
Pseudolycoriella zealandica (Edwards, 1927) ..... 16
Pseudolycoriella aotearoa sp. n. ..... 18
Pseudolycoriella sp. II ..... 20
Pseudolycoriella bispina complex ..... 20
Pseudolycoriella bispina Mohrig, 1999 ..... 21
Pseudolycoriella teo sp. n. ..... 22
Pseudolycoriella puhihi sp. n. ..... 23
Pseudolycoriella orite sp. n. ..... 24
Pseudolycoriella mahanga sp. n ..... 25
Pseudolycoriella porehu sp. n. ..... 26
Pseudolycoriella jejuna complex ..... 28
Pseudolycoriella jejuna (Edwards, 1927) ..... 30
Pseudolycoriella tuakana sp. n. ..... 31
Pseudolycoriella dagae sp. n. ..... 32
Pseudolycoriella raki sp. n. ..... 34
Pseudolycoriella hauta sp. n. ..... 37
Pseudolycoriella jejunella sp. n. ..... 38
Pseudolycoriella tewaipounamu sp. n. ..... 39
Pseudolycoriella whakahara sp. n ..... 41
Pseudolycoriella sudhausi sp. n. ..... 42
Pseudolycoriella whena sp. n. ..... 44
Pseudolycoriella porotaka sp. n. ..... 45
Pseudolycoriella nahenahe sp. n. ..... 47
Pseudolycoriella macrotegmenta complex ..... 47
Pseudolycoriella macrotegmenta Mohrig, 1999 ..... 48
Pseudolycoriella plicitegmenta sp. n. ..... 49
Pseudolycoriella subtilitegmenta sp. n. ..... 51
Pseudolycoriella gonotegmenta sp. n. ..... 53
Pseudolycoriella robustotegmenta sp. n. ..... 54
Pseudolycoriella huttoni sp. n. ..... 55
Pseudolycoriella wernermohrigi sp. n. ..... 56
Pseudolycoriella dentitegmenta sp. n. ..... 58
Pseudolycoriella breviseta Mohrig, 1999 ..... 59
Pseudolycoriella frederickedwardsi Köhler, 2016 ..... 59
Pseudolycoriella tonnoiri Köhler, 2016. ..... 61
Pseudolycoriella jaschhofi sp. n. ..... 61
Pseudolycoriella fiordlandia sp. n. ..... 62
Phylogenetic tree. ..... 63
Discussion. ..... 65
Acknowledgment ..... 66
References ..... 66


#### Abstract

In the course of the present study 28 species of the genus Pseudolycoriella Menzel \& Mohrig, 1998 from New Zealand were described as new to science: Pseudolycoriella aotearoa sp. n., Psl. dagae sp. n., Psl. dentitegmenta sp. n., Psl. fiordlandia sp. n., Psl. gonotegmenta sp. n., Psl. hauta sp. n., Psl. huttoni sp. n., Psl. jaschhofi sp. n., Psl. jejunella sp. n., Psl. kaikoura sp. n., Psl. maddisoni sp. n., Psl. mahanga sp. n., Psl. nahenahe sp. n., Psl. orite sp. n., Psl. plicitegmenta sp. n., Psl. porehu sp. n., Psl. porotaka sp. n., Psl. puhihi sp. n., Psl. raki sp. n., Psl. robustotegmenta sp. n., Psl. subtilitegmenta sp. n., Psl. sudhausi sp. n., Psl. teo sp. n., Psl. tewaipounamu sp. n., Psl. tuakana sp. n., Psl.


wernermohrigi sp. n., Psl. whakahara sp. n., and Psl. whena sp. n. Pseudolycoriella cavatica (Skuse, 1888), a widely distributed species, was recorded from New Zealand for the first time, and recognised as a senior synonym of Spathobdella setigera Hardy, 1960 syn. n. Apart from Psl. kaikoura and Psl. cavatica all New Zealand Pseudolycoriella species group in four different clusters: the Psl. bispina complex, the Psl. jejuna complex, the Psl. macrotegmenta complex, and the Psl. zealandica complex. The monophyly of those four species complexes was confirmed by a genetic analysis based on two mitochondrial genes (COI and 16S) and one nuclear gene (28S). A key to the species is given.

Key words: $16 \mathrm{~S}, 28 \mathrm{~S}$, Australasian region, COI, DNA sequencing, key, morphology, new species, new synonym, systematics, taxonomy

## Introduction

Menzel \& Mohrig (1998) recognised the morphological unity of several sciarid species which were previously scattered within the genera Sciara Meigen, Corynoptera Winnertz, and Lycoriella Frey. Accordingly, they transferred those species to the newly erected genus Pseudolycoriella Menzel \& Mohrig. The two authors stated that the following apomorphic characters demonstrate the monophyly of the new genus: one whip-lash hair on the apical third of the gonostylus, claws with teeth, and a tibial organ consisting of a dense row of bristles without a basal ridge. The monophyly of the genus Pseudolycoriella was later verified by a genetic analysis by Shin et al. (2013). Since 1998 several new species have been described and assigned to this genus. As of May 2019, Pseudolycoriella comprised 132 species (Menzel, personal communication). Several authors have stated that Pseudolycoriella is probably one of the most species-rich genera of the Sciaridae (e.g. Menzel \& Mohrig 2000; Rudzinski et al. 2016). Species of this genus are found in all biogeographic realms, but with different levels of diversity. Only a small proportion of all known species is found in the Palaearctic realm (Menzel \& Mohrig 2000), while the southern hemisphere is much more diverse (e.g. Vilkamaa et al. 2012; Mohrig 2013).

The intrageneric systematics of Pseudolycoriella is still in its infancy. Menzel \& Mohrig (2000) established three species groups—Psl. bruckii group, Psl. horribilis group, and Psl. morenae group-mainly to allow an easier use of identification keys, and thus implemented a first provisional system of Pseudolycoriella. Pseudolycoriella horribilis (Edwards), the only member of the Psl. horribilis species group, is characterized by the presence of a truncated tooth on the gonostylus that is not homologous with the gonostylar tooth of other sciarid taxa (Menzel \& Mohrig 2000). The two species of the Psl. morenae group possess conspicuous lobes with bristles on the inner gonocoxites and lack any teeth on the claws. Menzel \& Mohrig (2000) placed the remaining 17 species known at that time in the Psl. bruckii group, which comprises the majority of all species. The following diagnostic characters were given for this group: toothed claws, absence of lobes on the inner base of the gonocoxites, the absence of a tooth on the gonostylus, and up to five spines on the gonostylus (rarely without any spines). Later Mohrig (2013) amplified the provisional system by adding three further species groups to accomodate newly described Pseudolycoriella species from Papua New Guinea. These groups are the Psl. longicostalis group, the Psl. quadrispinosa group, and the Psl. triacanthula group. The Psl. longicostalis group comprises tiny species with deep sensory pits on the flagellomeres, a gonostylus with two spines (sometimes reduced), a tegmen without any secondary structures, and a conspicuous row of longitudinal bristles on the hind tibia. However, the delimitation between this group and the Psl. bruckii group remained vague. The Psl. quadrispinosa group is characterised by the presence of more than two spines on the gonostylus and a gradual reduction of the whip-lash hair. The species of the Psl. triacanthula group are relatively large and bear three or more robust spines on the gonostylus. The attempt to apply this species group concept to the New Zealand species Psl. frederickedwardsi Köhler, and Psl. tonnoiri Köhler failed, and Köhler \& Mohrig (2016) did not propose a new provisional species group to accomodate them. The first valid intrageneric taxon of Pseudolycoriella was established by Heller (2012), who changed the former genus Ostroverkhovana Komarova into a subgenus of Pseudolycoriella. However, Mohrig \& Kauschke (2019) disagreed with Heller (2012) and included Ostroverkhovana in Pseudolycoriella without distinguishing any subgenera. A possible congenericity of Pseudolycoriella and Eugnoriste Coquillett is still to be investigated (Vilkamaa et al. 2012).

The sciarids of New Zealand have attracted little attention by previous workers. Only two major studies (Tonnoir \& Edwards 1927; Mohrig \& Jaschhof 1999) and a few minor ones (Hutton 1881; Wisely 1959; Davies 1988; Köhler \& Mohrig 2016) dealt with this dipteran family. A total of 54 sciarid species are known from New Zealand, seven of which belong to Pseudolycoriella.

## Materials and methods

Collecting and depository. The majority of the sciarid flies examined were obtained as bycatch from a survey of New Zealand Cecidomyiidae by Catrin and Mathias Jaschhof between 2001 and 2002 (Jaschhof \& Jaschhof 2003a, 2003b, 2004; collecting permit \#9900/143/3/04 issued by the Department of Conservation, New Zealand). Another important part of the material was collected by Peter A. Maddison between 2014 and 2016. Only a small number of examined specimens was caught by other entomologists. Most specimens were collected with Malaise traps, aspirators, or sweep nets. Rather rarely, light traps, pitfall traps, and pan traps were used. Additionally, historic type material was borrowed from the relevant collections. The studied material is deposited in the following scientific collections: the Natural History Museum, London, United Kingdom (BMNH); the Centre for Biodiversity Genomics, Guelph, Canada (CBG); the New Zealand Arthropod Collection, Auckland, New Zealand (NZAC); the private collection of Werner Mohrig, Poseritz, Germany (PWMP); the Senckenberg Deutsches Entomologisches Institut, Müncheberg Germany (SDEI); and the Finnish Museum of Natural History, University of Helsinki, Helsinki, Finland (UZMH).

Morphology. All sciarids were stored in 70\% ethanol after they were collected. Prior to examination, the specimens were mounted in Canada balsam on glass slides following the instructions of Mohrig \& Jaschhof (1999), and Menzel \& Mohrig (2000). Accordingly, on each slide the hypopygium is mounted with the ventral side up, separated from the rest of the body, which lies on its right lateral side. Therefore the lengths of both these separated body parts have to be added, to obtain the total body length. The right wing was also separated and covered by a cover slip. For some specimens belonging to morphologically challenging species tergite IX was dissected from the remaining hypopygium to allow a better view of the tegmen. Morphological examination was conducted with an Olympus BX50 Microscope fitted with an Olympus U-DA drawing tube. The drawings were digitised using the software Adobe Illustrator CS6 according to Coleman (2003, 2009). Scanning electron microscope (SEM) images of details of the male genitalia of a species of the Psl. jejuna complex were obtained. One specimen was dehydrated sequentially with ethanol ( $70-98 \%$ ) and acetone ( $100 \%$ ), then critical-point dried, glued on a micro pin, and sputtered with gold. The images were taken with a SEM JSM-6060LV (JEOL) at an acceleration voltage of $8-12 \mathrm{kV}$.

The general terminology and the application of taxonomically important measurements follow Menzel \& Mohrig (2000) (Fig. 22). The length of the fourth flagellomere, which is necessary for the length-width ratio, represents the whole length i.e. the combined lengths of the flagellomere's body and the flagellomere's neck. The width of the fourth flagellomere was measured at the widest part, mostly at the apex of the flagellomere's body. The notation of the wing veins follows Hippa et al. (2010) (Fig. 9) and the use of the terms anterior portion of gonocoxal apodeme, posterior portion of gonocoxal apodeme, and parameral apodeme derives from Jaschhof \& Jaschhof (2009) (Fig. 11). For specimens of the Psl. bispina and the Psl.jejuna complex the term lateral lobes (e.g. $l l$ in Fig. 22) is used. This term is only used for descriptive purposes and should not be misinterpreted as a statement about homology. Where larger series of type specimens were available, a sample of at least ten specimens was measured using a micrometre. Specimens that could not be reliably determined to species level were not included in the type series, but are included as additional material. The order in which the species are treated is more or less arbitrary, although an attempt has been made to group closely related species together for easier comparison.

Molecular analysis. All specimens that underwent a molecular analysis are labelled with a unique number, consisting of the prefix "SDEI-Dipt-" and a seven-digit number. DNA was extracted with the E.Z.N.A. ${ }^{\text {® }}$ Tissue DNA Kit (Omega Bio-tek Inc.), following the manufacturer's instructions. To obtain a greater amount of DNA from old material, which had usually been stored for longer periods under poor conditions, the first extraction step-the tissue lysis-was prolonged overnight. Additionally, whole specimens were used in the lysis procedure instead of some individual body parts. Attempts were made to obtain sequences of the mitochondrial genes COI and 16S, and the nuclear gene 28 S via polymerase chain reactions (PCR) following Shin et al. (2013). In this phylogenetic study the nuclear gene 18 S was also sequenced. However, the 18 S gene could not be reliably obtained from the mostly older material which was available. In general, the primer pair LCO_1490 and HCO_2198 was used for COI amplifying (Folmer et al. 1994). If those primers did not show sufficient results, the primers LepF1 and LepR1 (Hebert et al. 2004), or LCO_1490 and HCO_2198 in combination with the internal primers sym-C1-J-1718 (Simon et al. 1994) and C1-N1760 (Prous et al. 2011) were used. The length of the COI sequence is 658 base pairs (bp). For the 16S sequence the primers 16SAR-L and 16SBR-H (Palumbi et al. 1991) were used. Due to a high percentage of failures in comparison to the other markers, an alternative primer set was designed. The new primers Psl_16S_for
and Psl_16S_rev bind within the 16 S region and therefore yield a shorter sequence than the primer pair 16SAR-L and $16 \mathrm{SBR}-\mathrm{H}$. The length of the 16 S sequence is 538 bp . A part of the nuclear 28 S sequence was obtained using the four primers 28 S A, 28S-rD7b1, 28S-rD5b, and 28S-Rd48a (Friedrich \& Tautz 1997; Whiting et al. 1997) as done by Shin et al. (2013). Furthermore, the primers 28SF2 and 28SB2 (Boevé et al. 2013) were used to get an additional part of the 28 S gene, which is located upstream (i.e., at the 5 ' end of the coding strand) of the former 28 S locus. Both sequences overlap in the nuclear genome and can therefore be easily pasted together to yield a sequence of $1,857 \mathrm{bp}$. All used primers, their annealing temperature and the the number of PCR cyles are listed in Table 1. PCR products were purified with Exonuclease I and FastAP Thermosensitive Alkaline Phosphatase (Life Technologies, Darmstadt, Germany) and sent to Macrogen Europe (Amsterdam, The Netherlands) for Sanger sequencing. The primers from each PCR were also used for sequencing. The software Geneious 11.0 .5 was used to assamble the resulting ab1-files provided by Macrogen and aligned with the online software MAFFT version 7 (https://mafft. cbrc.jp/alignment/software/; Katoh et al. 2002; Katoh \& Standley 2013; Katoh et al. 2017). The sequences used for the phylogenetic analyses were uploaded to GenBank (https://www.ncbi.nlm.nih.gov/genbank/). The accession numbers are listed in Table 2. Residual DNA extracts are deposited in the SDEI.

The pairwise p-distances (uncorrected) for the genes COI and 28 S were calculated with the software MEGA7 (Kumar et al. 2016) for each species. The maximal intraspecific p-distances between all available sequences of a species and the lowest p-distance to another species ("nearest neighbour") are given in each species account. Due to a future analysis, not all COI sequences used for the calculation of the p-distances are yet available online.

All three genes were used for the phylogenetic analysis. Each gene was aligned separately using default settings and afterwards concatenated using the software FASconCAT-G v1.04 (Kück \& Longo 2014). The phylogenetic analysis was carried out with the software IQ-TREE 1.6.1 (Nguyen et al. 2015) using the implemented tool Modelfinder (Kalyaanamoorthy et al. 2017) for an automatic model selection. The statistical validation of the phylogenetic tree was undertaken using the ultrafast bootstrap approximation [UFBoot] (Minh et al. 2013; Hoang et al. 2018). For the UFBoot estimation 1,000 replicates were deployed. As outgroup, two species of Diadocidiidae (Diptera, Sciaroidea) and one species of Scatopsidae (Diptera, Scatopsoidea) were used (see Table 2). The scatopsid species-Coboldia sp.-was used to root the phylogenetic tree.

## Key to the males of New Zealand Pseudolycoriella species

1. Gonostylus with one or two obvious spines .Psl. bruckii group, 2

- Gonostylus with more than two small spines. ..... Psl. macrotegmenta complex, 26

2. Gonostylus with two spines .....  3
Gonostylus with one spine ..... 11
3. Wing length greater than 2.4 mm ; length of gonocoxites more than $200 \mu \mathrm{~m}$ ..... 4
Wing length less than 2.4 mm ; length of gonocoxites $200 \mu \mathrm{~m}$ or less. .....  5
4. Base of the ejaculatory apodeme horseshoe-shaped . . . . . . . . . . . . . . . . . . . . . . . . . . . . . . . . . . . . Psl. zealandica (Edwards)

- Base of the ejaculatory apodeme elongate and slender Psl. aotearoa sp. n.

5. Fourth flagellomere less than twice as long as wide Psl. cavatica (Skuse)
Fourth flagellomere more than twice as long as wide Psl. bispina complex, 6
6. Fourth flagellomere less than three times as long as wide, with dense setae (Fig. 5) ..... Psl. puhihi sp. n.
Fourth flagellomere at least three times as long as wide, with sparsely scattered setae (e.g. Fig. 4). ..... 7
7. Wings slender, anal area weakly developed ..... 8
Wings of normal shape, anal area well developed . ..... 9
8. Wing length less than 2.2 mm ; c/w ratio greater than 0.65 .Psl. orite sp. n.
Wing length 2.2 mm or more; c/w ratio less than 0.65 ..... Psl. porehu sp. n.
9. Base of the ejaculatory apodeme lyre-shaped; apex of tegmen surrounded by a sclerotized margin (Figs $14 \& 18$ ); body lengthmore than 1.8 mm , gonocoxites of normal length 10
Base of the ejaculatory apodeme u-shaped; apex of tegmen with membranous area (Fig. 15), body length less than 1.8 mm ,gonocoxites short. Psl. teo sp. n.
10. Apex of $R_{5}$ with macrotrichia only on ventral side; hind tibia with ten or more robust bristles in a longitudinal rowPsl. bispina MohrigApex of $\mathrm{R}_{5}$ with macrotrichia on both sides; hind tibia with less than ten robust bristles in a longitudinal row.
Psl. mahanga sp. n.
11. Fourth flagellomere less than twice as long as wide; tegmen apically roundish (Fig. 11). Psl. kaikoura sp. n.

- Fourth flagellomere more than twice as long as wide; tegmen with a pronounced apex (at in Fig. 22) .
Psl. jejuna complex, 12

12. Scape and pedicel bright yellowish, contrasting strongly with the brown flagellomeres ..... 13
Scape and pedicel pale brown, contrasting slightly with flagellomeres. ..... 17
13. Body length less than 2.5 mm ..... 14
Body length greater than 2.5 mm ..... 15
14. Fourth antennal flagellomere at least 2.8 times longer than broad; gonostylus more than 2.8 times longer than broad; winglength less than 2.2 mm (ca. $1.9-2.1 \mathrm{~mm}$ )Psl. whena sp. n.
Fourth antennal flagellomere less than 2.8 times longer than broad; gonostylus less than 2.8 times longer than broad; wing
length 2.2 mm
.Psl. nahenahe sp. n.
15. Ejaculatory apodeme slender (Fig. 39); fourth antennal flagellomere less than 2.6 times as long as broad. Psl. porotaka sp. n.Ejaculatory apodeme broad (Fig. 42); fourth antennal flagellomere at least 2.6 times longer than broad16
16. Gonostylus more than 2.8 times as long as broad; inner base of gonostylus with a conspicuous angle (ba in Fig. 42)
.Psl. dagae sp. n.
Gonostylus less than 2.8 times as long as broad; inner base of gonostylus without or only with a minor angle (Fig. 41)
Psl. raki sp. n.
17. Ratio $R_{1} / R$ less than 0.80 ..... 18
Ratio $\mathrm{R}_{1} / \mathrm{R}$ greater than or equal to 0.80 ..... 22
18. Mesonotum with long and dense setae; 5-6 bristles on the scutellum.Mesonotum with normal setosity; 2-4 bristles on the scutellum19
19. Fourth antennal flagellomere less than 2.8 times longer than broad ..... 20
Fourth antennal flagellomere more than 2.8 times longer than broad Psl. sudhausi sp. n.
20. Small species; wing length less than 2.5 mm ; gonostylus slender .Psl. jejunella sp. n.
Larger species; wing length greater than 2.5 mm .22. Apical cavity on gonostylus prominent and extensively bare, without microtrichia (Figs 20 \& 22).23
Apical cavity on gonostylus absent or inconspicuous, or with microtrichia ..... 24
21. Wing length greater than 3.0 mm Psl. whakahara sp. n.

- Wing length less than 3.0 mm Psl. tewaipounamu sp. n.

24. Ejaculatory apodeme slender (Figs 37 \& 39). Psl. sudhausi sp. n.
Ejaculatory apodeme broad (e.g. Fig. 41) ..... 25
25. Apical spine on the gonostylus short, not protruding from the surrounding setae (Figs 30-31); mid tibia without robust bristlesamong the vestiturePsl. tuakana sp. n.
Apical spine on the gonostylus slightly longer, protruding from the surrounding setae (Figs 25-26); mid tibia with robust brist-les among the vestiturePsl. jejuna (Edwards)
26. Tegmen laterally with small teeth ( $l t$ in Fig. 52) ..... 31
Tegmen laterally without small teeth ..... 27
27. Tegmen basolaterally with robust projections (Fig. 59), apex of gonostylus broadTegmen basolaterally without projections, apex of gonostylus not broader than base28
28. Tegmen large with lateral ledges (Fig. 55); gonostylus strongly curved ..... n.
Tegmen of normal proportions; gonostylus elongated ..... 29
29. Tegmen with a medial, hood-like structure (Fig. 56); gonostylus longer, usually more than 3.5 times as long as wide
Psl. jaschhofi sp. n.Tegmen without a medial structure; gonostylus shorter, usually less than 3.5 times as long as wide30
30. Tegmen with recurved lateral edges; fourth flagellomere more than 2.8 times as long as broad . Psl. frederickedwardsi Köhler
Tegmen without lateral structures; fourth flagellomere equal to or less than 2.8 times as long as broad . . Psl. tonnoiri Köhler
31. Tegmen with teeth laterally and apically (Fig. 60) Psl. dentitegmenta sp. n.Tegmen with lateral teeth only32
32. Tegmen with dorsal structures ( $d s$ in Fig. 52); spines on gonostylus more transversely oriented ..... 33

- Tegmen without dorsal structures; gonostylus bulging apically, spines more longitudinally orientated.Psl. macrotegmenta Mohrig

33. Base of vein $M$ longer than $M$-fork; sclerotized margin of tegmen very broad; tegmen with dorsal structures strongly developed,reaching apex of tegmen.Psl. robustotegmenta sp. n
Base of $M$ vein shorter than M-fork; sclerotized margin of tegmen narrower; dorsal structures mostly not reaching apex oftegmen (except in Psl. wernermohrigi)34
34. Margin of tegmen narrowly sclerotized; gonostylus apically tapered Psl. subtilitegmenta sp. n.
Margin of tegmen broadly sclerotized; gonostylus apically not tapered ..... 35
35. Ejaculatory apodeme long (Fig. 50); sclerotized edge of tegmen apically closed. .Psl. wernermohrigi sp. n.
Ejaculatory apodeme short (e.g. Fig. 51); sclerotized edge of tegmen apically closed or interrupted. ..... 36
36. Base of ejaculatory apodeme $v$-shaped; ratio $R_{1} / R$ mostly greater than 0.65 Psl. gonotegmenta sp. n.

- Base of ejaculatory apodeme $u$-shaped; ratio $\mathrm{R}_{1} / \mathrm{R}$ less than 0.65 ..... 37

37. Gonostylus more than 3.0 times as long as broad; sclerotized edge of tegmen apically interrupted; ratio $\mathrm{R}_{1} / \mathrm{R}$ less than $0.55 \ldots$.Psl. plicitegmenta sp. n.Gonostylus less than 3.0 times as long as broad; sclerotized edge of tegmen apically closed; ratio $\mathrm{R}_{1} / \mathrm{R}$ greater than 0.55
TABLE 1. List of primers used for PCR.

| Locus | Primer | Direction <br> F/R (i) | Primer sequence ( $5^{\prime}$ to $3^{\prime}$ ) | Annealing temperature | PCR cycles | Reference |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| COI | LCO_1490 | F | GGTCAACAAATCATAAAGATATTGG | $49^{\circ} \mathrm{C}$ | 38 | Folmer et al. (1994) |
| COI | HCO_2198 | R | TAAACTTCAGGGTGACCAAAAAATCA | $49^{\circ} \mathrm{C}$ | 38 | Folmer et al. (1994) |
| COI | LepF1 | F | ATTCAACCAATCATAAAGATATTGG | $49^{\circ} \mathrm{C}$ | 38 | Hebert et al. (2004) |
| COI | LepR1 | R | TAAACTTCAGGGTGACCAAAAAATCA | $49^{\circ} \mathrm{C}$ | 38 | Hebert et al. (2004) |
| COI | sym-C1-J-1718 | F (i) | GGAGGATTTGGAAAYTGAYTAGTWCC | $49^{\circ} \mathrm{C}$ | 38 | Simon et al. (1994) |
| COI | C1-N1760 | R (i) | GGTARAAATCARAATCTTATATTAT | $49^{\circ} \mathrm{C}$ | 38 | Prous et al. (2011) |
| 16S | 16SAR-L | F | CGCCTGTTTATCAAAAACAT | $49^{\circ} \mathrm{C}$ | 38 | Palumbi et al. (1991) |
| 16S | 16SBR-H | R | CCGGTCTGAACTCAGATCACGT | $49^{\circ} \mathrm{C}$ | 38 | Palumbi et al. (1991) |
| 16S | Psl_16S_for | F (i) | GTACAAAGGTAGCATAATCRTT | $53^{\circ} \mathrm{C}$ | 38 | present study |
| 16S | Psl_16S_rev | R (i) | AATCCAAMATSGAGGTCGCAA | $53^{\circ} \mathrm{C}$ | 38 | present study |
| 28S | 28 S A | F | GACCCGTCTTGAAACACGGA | $49^{\circ} \mathrm{C}$ | 40 | Whiting et al. (1997) |
| 28S | 28S-rD7b1 | R | GACTTCCCTTACCTACAT | $49^{\circ} \mathrm{C}$ | 40 | Friedrich \& Tautz (1997); <br> Whiting et al. (1997) |
| 28S | 28S-Rd48a | F (i) | ACCTATTCTCAAACTTTAAATGG | $49^{\circ} \mathrm{C}$ | 40 | Whiting et al. (1997) |
| 28S | 28S-rD5b | R (i) | CCACAGCGCCAGTTCTGCTTAC | $49^{\circ} \mathrm{C}$ | 40 | Whiting et al. (1997) |
| 28S | 28SF2 | F | CACGAGCCGATAGCGAACAAG | $54^{\circ} \mathrm{C}$ | 40 | Boevé et al. (2013) |
| 28S | 28 SB 2 | R | CCAAGGCCTCTAATCATTCGC | $54^{\circ} \mathrm{C}$ | $40$ | Boevé et al. (2013 |

TABLE 2. List of sequenced specimens used in this study with GenBank accession numbers or BOLD link.

| Species | Family | Country | Sample number | GenBank Accession Number |  |  | Reference |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  | COI | 16S | 28S |  |
| Diadocidia ferruginosa | Diadocidiidae | Slovakia | SJ1 | KC435634 | KC435562 | KC435598 | Ševčík et al. (2013) |
| Diadocidia spinosula | Diadocidiidae | Slovakia | JSSJ7 | KJ136800 | KJ136737 | KJ136773 | Ševčík et al. (2014) |
| Coboldia sp. | Scatopsidae | China | 905 | JQ613854 | JQ613854 | JQ613758 | Shin et al. (2013) |
| Corynoptera sp. bs9 | Sciaridae | New Zealand | SDEI-Dipt-0000853 | MK906346 | MK906438 | MK906528 |  |
| Ctenosciara etorutao | Sciaridae | New Zealand | SDEI-Dipt-0001305 | MK906386 | MK906481 | MK906571 |  |
| Pseudolycoriella aotearoa | Sciaridae | New Zealand | SDEI-Dipt-0000997 | MK906359 | MK906451 | MK906541 |  |
| Psl. bispina | Sciaridae | New Zealand | SDEI-Dipt-0000946 | MK906352 | MK906444 | MK906534 |  |
| Psl. bispina | Sciaridae | New Zealand | SDEI-Dipt-0000974 | MK906355 | MK906447 | MK906537 |  |
| Psl. bispina | Sciaridae | New Zealand | SDEI-Dipt-0001251 | MK906384 | MK906479 | MK906569 |  |
| Psl. bispina | Sciaridae | New Zealand | SDEI-Dipt-0001518 | MK906399 | MK906497 | MK906587 |  |
| Psl. bispina | Sciaridae | New Zealand | SDEI-Dipt-0001522 | MK906401 | MK906499 | MK906589 |  |
| Psl. cavatica | Sciaridae | New Zealand | SDEI-Dipt-0001085 | MK906405 | MK906455 | MK906545 |  |
| Psl. cavatica | Sciaridae | New Zealand | SDEI-Dipt-0001411 | MK906395 | MK906490 | MK906580 |  |
| Psl. dagae | Sciaridae | New Zealand | SDEI-Dipt-0000628 | MK906330 | MK906421 | MK906511 |  |
| Psl. dagae | Sciaridae | New Zealand | SDEI-Dipt-0000655 | MK906331 | MK906422 | MK906512 |  |
| Psl. frederickedwardsi | Sciaridae | New Zealand | SDEI-Dipt-0000756 | MK906339 | MK906431 | MK906521 |  |
| Psl. frederickedwardsi | Sciaridae | New Zealand | SDEI-Dipt-0000772 | MK906341 | MK906433 | MK906523 |  |
| Psl. frederickedwardsi | Sciaridae | New Zealand | SDEI-Dipt-0001224 | MK906374 | MK906468 | MK906558 |  |
| Psl. frederickedwardsi | Sciaridae | New Zealand | SDEI-Dipt-0001237 | MK906379 | MK906474 | MK906564 |  |
| Psl.frederickedwardsi | Sciaridae | New Zealand | SDEI-Dipt-0001247 | MK906382 | MK906477 | MK906567 |  |
| Psl. gonotegmenta | Sciaridae | New Zealand | SDEI-Dipt-0000977 | MK906356 | MK906448 | MK906538 |  |
| Psl. hauta | Sciaridae | New Zealand | SDEI-Dipt-0000506 | MK906322 | MK906412 | MK906502 |  |
| Psl. horribilis | Sciaridae | Korea | 074 | JQ613788 | JQ613887 | JQ613691 | Shin et al. (2013) |
| Psl. huttoni | Sciaridae | New Zealand | SDEI-Dipt-0000621 | MK906327 | MK906418 | MK906508 |  |
| Psl.jaschhofi | Sciaridae | New Zealand | SDEI-Dipt-0001511 | MK906409 | MK906494 | MK906584 |  |
| Psl.jaschhofi | Sciaridae | New Zealand | SDEI-Dipt-0001513 | MK906410 | MK906495 | MK906585 |  |
| Psl.jaschhofi | Sciaridae | New Zealand | SDEI-Dipt-0001514 | MK906398 | MK906496 | MK906586 |  |

TABLE 2. (Continued)

| Species | Family | Country | Sample number | GenBank Accession Number |  |  | Reference |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  | COI | 16S | 28 S |  |
| Psl. jejuna | Sciaridae | New Zealand | SDEI-Dipt-0000603 | MK906326 | MK906416 | MK906506 |  |
| Psl.jejuna | Sciaridae | New Zealand | SDEI-Dipt-0000617 | MK906403 | MK906417 | MK906507 |  |
| Psl.jejuna | Sciaridae | New Zealand | SDEI-Dipt-0000744 | MK906404 | MK906428 | MK906518 |  |
| Psl. jejuna | Sciaridae | New Zealand | SDEI-Dipt-0000773 | MK906342 | MK906434 | MK906524 |  |
| Psl. jejuna | Sciaridae | New Zealand | SDEI-Dipt-0000829 | MK906344 | MK906436 | MK906526 |  |
| Psl.jejuna | Sciaridae | New Zealand | SDEI-Dipt-0001193 | MK906371 | MK906464 | MK906554 |  |
| Psl.jejuna | Sciaridae | New Zealand | SDEI-Dipt-0001195 | MK906406 | MK906466 | MK906556 |  |
| Psl.jejuna | Sciaridae | New Zealand | SDEI-Dipt-0001339 | MK906390 | MK906485 | MK906575 |  |
| Psl. jejuna | Sciaridae | New Zealand | SDEI-Dipt-0001347 | MK906391 | MK906486 | MK906576 |  |
| Psl. jejunella | Sciaridae | New Zealand | SDEI-Dipt-0000935 | MK906351 | MK906443 | MK906533 |  |
| Psl. kaikoura | Sciaridae | New Zealand | SDEI-Dipt-0000500 | MK906321 | MK906411 | MK906501 |  |
| Psl. macrotegmenta | Sciaridae | New Zealand | SDEI-Dipt-0000757 | MK906340 | MK906432 | MK906522 |  |
| Psl. maddisoni | Sciaridae | New Zealand | SDEI-Dipt-0000627 | MK906329 | MK906420 | MK906510 |  |
| Psl.maddisoni | Sciaridae | New Zealand | SDEI-Dipt-0000664 | MK906332 | MK906423 | MK906513 |  |
| Psl. mahanga | Sciaridae | New Zealand | SDEI-Dipt-0000595 | MK906324 | MK906414 | MK906504 |  |
| Psl. mahanga | Sciaridae | New Zealand | SDEI-Dipt-0001008 | MK906362 | MK906454 | MK906544 |  |
| Psl. orite | Sciaridae | New Zealand | SDEI-Dipt-0000863 | MK906347 | MK906439 | MK906529 |  |
| Psl. plicitegmenta | Sciaridae | New Zealand | SDEI-Dipt-0001153 | MK906366 | MK906459 | MK906549 |  |
| Psl. plicitegmenta | Sciaridae | New Zealand | SDEI-Dipt-0001158 | MK906367 | MK906460 | MK906550 |  |
| Psl. porotaka | Sciaridae | New Zealand | SDEI-Dipt-0001225 | MK906375 | MK906469 | MK906559 |  |
| Psl.porotaka | Sciaridae | New Zealand | SDEI-Dipt-0001250 | MK906383 | MK906478 | MK906568 |  |
| Psl. puhihi | Sciaridae | New Zealand | SDEI-Dipt-0001005 | MK906361 | MK906453 | MK906543 |  |
| Psl. puhihi | Sciaridae | New Zealand | SDEI-Dipt-0001422 | MK906396 | MK906491 | MK906581 |  |
| Psl. raki | Sciaridae | New Zealand | SDEI-Dipt-0000834 | MK906345 | MK906437 | MK906527 |  |
| Psl. robustotegmenta | Sciaridae | New Zealand | SDEI-Dipt-0001358 | MK906392 | MK906487 | MK906577 |  |
| Psl. robustotegmenta | Sciaridae | New Zealand | SDEI-Dipt-0001371 | MK906393 | MK906488 | MK906578 |  |

TABLE 2. (Continued)

| Species | Family | Country | Sample number | GenBank Accession Number |  |  | Reference |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  | COI | 16S | 28 S |  |
| Psl. sp. 1 SS-2012 | Sciaridae | Korea | 586 | JQ613816 | JQ613916 | JQ613720 | Shin et al. (2013) |
| Psl. sp. 2 SS-2012 | Sciaridae | Korea | 640 | JQ613823 | JQ613923 | JQ613727 | Shin et al. (2013) |
| Psl. sp. I | Sciaridae | New Zealand | BIOUG14863-H041 | see BOLD (2019b) | $\mathrm{n} / \mathrm{a}$ | $\mathrm{n} / \mathrm{a}$ | BOLD (2019b) |
| Psl. sp. I | Sciaridae | New Zealand | BIOUG15289-H10 | see BOLD (2019b) | $\mathrm{n} / \mathrm{a}$ | $\mathrm{n} / \mathrm{a}$ | BOLD (2019b) |
| Psl. sp. I | Sciaridae | New Zealand | BIOUG15552-D06 | see BOLD (2019b) | $\mathrm{n} / \mathrm{a}$ | $\mathrm{n} / \mathrm{a}$ | BOLD (2019b) |
| Psl. sp. I | Sciaridae | New Zealand | BIOUG15560-H06 | see BOLD (2019b) | $\mathrm{n} / \mathrm{a}$ | $\mathrm{n} / \mathrm{a}$ | BOLD (2019b) |
| Psl. sp. II | Sciaridae | New Zealand | SDEI-Dipt-0000745 | MK906337 | MK906429 | MK906519 |  |
| Psl. sp. SS-2012 | Sciaridae | China | 084 | JQ613789 | JQ613888 | JQ613692 | Shin et al. (2013) |
| Psl. subtilitegmenta | Sciaridae | New Zealand | SDEI-Dipt-0001151 | MK906364 | MK906457 | MK906547 |  |
| Psl. sudhausi | Sciaridae | New Zealand | SDEI-Dipt-0000885 | MK906348 | MK906440 | MK906530 |  |
| Psl. sudhausi | Sciaridae | New Zealand | SDEI-Dipt-0001167 | MK906368 | MK906461 | MK906551 |  |
| Psl. sudhausi | Sciaridae | New Zealand | SDEI-Dipt-0001170 | MK906369 | MK906462 | MK906552 |  |
| Psl. sudhausi | Sciaridae | New Zealand | SDEI-Dipt-0001175 | MK906370 | MK906463 | MK906553 |  |
| Psl. sudhausi | Sciaridae | New Zealand | SDEI-Dipt-0001459 | MK906397 | MK906493 | MK906583 |  |
| Psl.teo | Sciaridae | New Zealand | SDEI-Dipt-0001528 | MK906402 | MK906500 | MK906590 |  |
| Psl. tewaipounamu | Sciaridae | New Zealand | SDEI-Dipt-0000513 | MK906323 | MK906413 | MK906503 |  |
| Psl. tewaipounamu | Sciaridae | New Zealand | SDEI-Dipt-0000752 | MK906338 | MK906430 | MK906520 |  |
| Psl. tewaipounamu | Sciaridae | New Zealand | SDEI-Dipt-0000907 | MK906350 | MK906442 | MK906532 |  |
| Psl. tewaipounamu | Sciaridae | New Zealand | SDEI-Dipt-0000956 | MK906353 | MK906445 | MK906535 |  |
| Psl. tewaipounamu | Sciaridae | New Zealand | SDEI-Dipt-0000960 | MK906354 | MK906446 | MK906536 |  |
| Psl. tewaipounamu | Sciaridae | New Zealand | SDEI-Dipt-0000994 | MK906358 | MK906450 | MK906540 |  |
| Psl. tonnoiri | Sciaridae | New Zealand | SDEI-Dipt-0000597 | MK906325 | MK906415 | MK906505 |  |
| Psl. tonnoiri | Sciaridae | New Zealand | SDEI-Dipt-0000980 | MK906357 | MK906449 | MK906539 |  |
| Psl. tonnoiri | Sciaridae | New Zealand | SDEI-Dipt-0001140 | MK906363 | MK906456 | MK906546 |  |
| Psl. tonnoiri | Sciaridae | New Zealand | SDEI-Dipt-0001152 | MK906365 | MK906458 | MK906548 |  |
| Psl. tuakana | Sciaridae | New Zealand | SDEI-Dipt-0001194 | MK906372 | MK906465 | MK906555 |  |

TABLE 2. (Continued)

| Species | Family | Country | Sample number | GenBank Accession Number |  |  | Reference |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  | COI | 16S | 28S |  |
| Psl. tuakana | Sciaridae | New Zealand | SDEI-Dipt-0001337 | MK906389 | MK906484 | MK906574 |  |
| Psl. wernermohrigi | Sciaridae | New Zealand | SDEI-Dipt-0000665 | MK906333 | MK906424 | MK906514 |  |
| Psl. wernermohrigi | Sciaridae | New Zealand | SDEI-Dipt-0000670 | MK906334 | MK906425 | MK906515 |  |
| Psl. wernermohrigi | Sciaridae | New Zealand | SDEI-Dipt-0000674 | MK906335 | MK906426 | MK906516 |  |
| Psl. wernermohrigi | Sciaridae | New Zealand | SDEI-Dipt-0001223 | MK906373 | MK906467 | MK906557 |  |
| Psl. wernermohrigi | Sciaridae | New Zealand | SDEI-Dipt-0001227 | MK906376 | MK906470 | MK906560 |  |
| Psl. wernermohrigi | Sciaridae | New Zealand | SDEI-Dipt-0001234 | MK906378 | MK906472 | MK906562 |  |
| Psl. wernermohrigi | Sciaridae | New Zealand | SDEI-Dipt-0001235 | MK906407 | MK906473 | MK906563 |  |
| Psl. wernermohrigi | Sciaridae | New Zealand | SDEI-Dipt-0001238 | MK906380 | MK906475 | MK906565 |  |
| Psl. wernermohrigi | Sciaridae | New Zealand | SDEI-Dipt-0001245 | MK906381 | MK906476 | MK906566 |  |
| Psl. wernermohrigi | Sciaridae | New Zealand | SDEI-Dipt-0001270 | MK906385 | MK906480 | MK906570 |  |
| Psl. wernermohrigi | Sciaridae | New Zealand | SDEI-Dipt-0001334 | MK906387 | MK906482 | MK906572 |  |
| Psl. wernermohrigi | Sciaridae | New Zealand | SDEI-Dipt-0001373 | MK906394 | MK906489 | MK906579 |  |
| Psl. cf. wernermohrigi | Sciaridae | New Zealand | SDEI-Dipt-0000681 | MK906336 | MK906427 | MK906517 |  |
| Psl. cf. wernermohrigi | Sciaridae | New Zealand | SDEI-Dipt-0001232 | MK906377 | MK906471 | MK906561 |  |
| Psl. whakahara | Sciaridae | New Zealand | SDEI-Dipt-0001439 | MK906408 | MK906492 | MK906582 |  |
| Psl. whena | Sciaridae | New Zealand | SDEI-Dipt-0001336 | MK906388 | MK906483 | MK906573 |  |
| Psl.zealandica | Sciaridae | New Zealand | SDEI-Dipt-0000623 | MK906328 | MK906419 | MK906509 |  |
| Psl.zealandica | Sciaridae | New Zealand | SDEI-Dipt-0000886 | MK906349 | MK906441 | MK906531 |  |
| Psl.zealandica | Sciaridae | New Zealand | SDEI-Dipt-0000998 | MK906360 | MK906452 | MK906542 |  |
| Zygoneura contractans | Sciaridae | New Zealand | SDEI-Dipt-0000826 | MK906343 | MK906435 | MK906525 |  |



FIGURES 1-9. Various body parts of Pseudolycoriella spp., in lateral view. 1. Pseudolycoriella dagae sp. n., fourth flagellomere. 2. Pseudolycoriella jejunella sp. n., fourth flagellomere. 3. Pseudolycoriella aotearoa $\mathbf{s p}$. n., fourth flagellomere. 4. Pseudolycoriella bispina Mohrig, fourth flagellomere. 5. Pseudolycoriella puhihi $\mathbf{s p}$. n., fourth flagellomere. 6. Pseudolycoriella jejunella sp. n., palp. 7. Pseudolycoriella robustotegmenta sp. n., head. 8. Pseudolycoriella jejunella sp. n., tibial organ. 9. Pseudolycoriella hauta sp. n., wing. Abbreviations: $\mathrm{aa}=$ anal area; $\mathrm{bM}=$ base of the medial vein; $\mathrm{c}=$ distance between intersection of vein $R_{5}$ and radial vein and the end of costal vein; $\mathrm{CuA}_{1} / \mathrm{CuA}_{2}=$ first and second branch of the anterior branch of the cubital vein; $M_{1} / M_{2}=$ first and second branch of the medial vein ("M-fork"); $R=$ radial vein; $R_{1} / R_{5}=$ first and fifth branch of the radial vein; $r-m=$ crossvein between radial and median veins; $w=$ distance between intersection of vein $R_{5}$ and radial vein and intersection of vein $M_{1}$ and wing margin.

## Species account

## Pseudolycoriella cavatica (Skuse, 1888)

(Fig. 10)

## Synonyms.

Sciara familiaris Skuse, 1888
Sciara festiva Skuse, 1888
Spathobdella setigera Hardy, 1960 syn. n.

Literature. Sciara cavatica Skuse, 1888; S. familiaris Skuse, 1888 \& S. festiva Skuse, 1888: Skuse (1888): 687-690.—Pseudolycoriella cavatica (Skuse, 1888): Broadley et al. (2016): 437, 439-441, Fig. 23 A-D \& Fig. 24 A-B.-Spathobdella setigera Hardy, 1960: Hardy (1960): 234, Fig. 80 a-c.—Bradysia setigera (Hardy, 1960): Steffan (1973): 356.—Leblanc et al. (2009): 1455.—Pseudolycoriella setigera (Hardy, 1960): Menzel \& Heller (2007): 223.—Menzel \& Smith (2009): 41-43, Figs 1.45-47.-Köhler \& Menzel (2013): 69.-Menzel et al. (2013): 292-293, Figs 22-23.-Menzel \& Smith (2017): 21, Fig. 82.-Mohrig et al. (2019): 426-427, Fig. 17 A-E.—Sciaridae indet.: BOLD (2019a).

Material studied. $1 \delta^{\lambda}$ New Zealand: North Island, Auckland, Manurewa, Mill Road, pitfall trap, 03-13.02.2015, leg. P.A. Maddison (SDEI, SDEI-Dipt-0001281). 2 §§ same locality as previous, 14-27.02.2015 (NZAC, SDEI-Dipt-0001283; SDEI, SDEI-Dipt-0001255). 9ふへ North Island, Western Bay of Plenty, Katikati, 140 Wharawhara Road, bush area, pitfall trap, 14.02-05.03.2015, leg. P.A. Maddison (4x NZAC; 5x SDEI, SDEI-Dipt-0001411). $1 \delta^{\lambda}$ same locality as previous, $14.02-05.03 .2015$, Malaise trap, leg. P.A. Maddison (NZAC, SDEI-Dipt-0001085). $2 q$ ㅇ North Island, Western Bay of Plenty, Katikati, 140 Wharawhara Road, kiwifruit block, Malaise trap, 14.0205.03.2015, leg. P.A. Maddison (all SDEI, SDEI-Dipt-0001443 \& SDEI-Dipt-0001458). 3 § $\begin{gathered}\text { § North Island, West- }\end{gathered}$ ern Bay of Plenty, Katikati, Te Mania catchment, Malaise trap, 19.09-27.10.2016, leg. P.A. Maddison (1x NZAC, SDEI-Dipt-0001231; 2x SDEI, SDEI-Dipt-0001229 \& SDEI-Dipt-0001368). $10^{\text {§ N North Island, Western Bay of }}$ Plenty, Katikati, Uretara, mangrove area, Malaise trap, 04.09-22.10.2016, leg. P.A. Maddison (SDEI, SDEI-Dipt0001377). 2 q $q$ North Island, Western Bay of Plenty, Katikati, 17 Francis Drive, Malaise trap, 14.02-05.03.2015, leg. P.A. Maddison (NZAC, SDEI-Dipt-0001298; SDEI, SDEI-Dipt-0001297). 2§ § South Island, Buller, Ahaura, Granville State Forest, altitude 170-250 m, Nothofagus truncata forest, Malaise trap, 01.12.1994, leg. J. Hutcheson (all SDEI). $1 \delta^{\lambda}$ South Island, Christchurch, Halswell quarry, light trap and pan trap, 26.02.2004, leg. R.P. Macfarlane (UZMH).

Description. See Menzel \& Smith (2009) and Broadley et al. (2016).
Genetic distances. All 16 investigated COI sequences are identical and do not show any pairwise distance. Among the New Zealand species, the nearest neighbour is Pseudolycoriella teo, diverging by a minimum of $8.85 \%$. No differences between two conspecific 28S sequences were found. The nearest neighbours are Psl. horribilis and an undescribed species from Korea (Psl. sp. 1 SS-2012). Both p-distances are $0.75 \%$.

Distribution. Australia (New South Wales and Western Australia [BOLD 2019a]); Hawaii, New Caledonia, Seychelles, Tristan da Cunha, and South Africa (BOLD 2019a). New to New Zealand.

Remarks. The COI-barcode of Psl. cavatica is also recorded on the Barcode of Life Data System (BOLD 2019a). There it is listed under the BIN BOLD:ABW3602. All these new records fit well into the already known distribution pattern of this species, which comprises Australia and islands south of the Tropic of Cancer. The frequent occurrence of Psl. cavatica in pitfall traps suggests an epigean ecology as already noted by Menzel \& Smith (2009).

## Pseudolycoriella kaikoura sp. n.

(Fig. 11)
urn:Isid:zoobank.org:act:F587D109-BD83-490F-921F-E515D4356B80

Material studied. Holotype male. New Zealand, South Island, Kaikoura, Waiau, Pillona, altitude 400 m, Kunzea ericoides over water supply seepage, Malaise trap, 26.12.2000-12.01.2001, leg. P.M. Johns (NZAC, SDEI-Dipt0000500 ). Paratype. $1 \AA^{\lambda}$ same locality and same date as holotype (SDEI, SDEI-Dipt-0000501).

Description. Male. Head. Head capsule brown. Eye bridge two to three facets wide. Antennae light brown,
without any contrast between scape, pedicel and flagellum; fourth flagellomere 1.8-1.9 times longer than wide; necks of flagellomeres well differentiated; surface of flagellomeres rough with deep pits, sensilla of two different lengths; setae on the flagellomere as long as flagellomere width. Maxillary palp pale with three palpomeres, first one longest, second and third short and of same length, first palpomere with two to four bristles; patch of sensilla on first palpomere not conspicuous. Prefrons and clypeus bulging. Thorax pale brown. Posterior pronotum bare. Anterior pronotum with three to five setae. Episternum 1 with four to seven setae. Mesonotum with three long lateral bristles and several small ones. Scutellum with four robust bristles and several smaller setae. Katepisternum as long as high, distal part slightly darker than proximal part. Wing. Length 2.0 mm ; width/length ratio 0.35 . Membrane transparent and without macrotrichia, anal area present; all posterior veins distinct; r-m and bM without macrotrichia; $\mathrm{R}_{5}$ with macrotrichia on dorsal side only (the paratype bears two additional macrotrichia at ventral side of $\mathrm{R}_{5}$ apex); $\mathrm{R}_{1}$ approximately $0.7-0.8$ times as long as R ; $\mathrm{c} / \mathrm{w}$ ratio 0.7 ; r-m as long as bM . Haltere brownish; with a short shaft; head of haltere longer than shaft. Legs yellowish brown. Tibial organ surrounded by a circular border and as wide as half width of tibial apex; bristles of tibial organ arranged in an irregular row. Front tibia with one robust bristle among vestiture. Mid tibia with one slightly robust bristle. Hind tibia with a posteriodorsal row of eight robust bristles, comprising apical two-thirds of tibia, additional robust bristles on the ventral and posterior sides. All tibial spurs equal in length. Claws with teeth. Abdomen pale brown, slightly paler than thorax. Hypopygium (Fig. 11). Gonocoxites wider than long, inner side of gonocoxites with short setae, outer side with long hairs; ventral, inner side of gonocoxites v-shaped and basally widely separated; apicolaterally strongly incised. Gonostylus two times longer than wide; with a prominent edge in the middle of the inner side, one spine and one long whip-lash hair subapically, apex with a dense vestiture of robust setae. Tegmen apical roundish, lateral parts sclerotized; parameral apodeme very robust. Area of teeth with approximately ten tiny teeth. Ejaculatory apodeme short and dark; base distinct, vshaped. Posterior portion of gonocoxal apodeme long, brown, widely separated, and medially connected. Anterior portion of gonocoxal apodeme long and dark brown.


10


11
pga

$\frac{10-11}{100 \mu \mathrm{~m}}$

FIGURES 10-11. Pseudolycoriella bruckii group, hypopygia. 10. Pseudolycoriella cavatica (Skuse). 11. Pseudolycoriella kaikoura sp. n. Abbreviations: aga $=$ anterior portion of gonocoxal apodeme; $\mathrm{pa}=$ parameral apodeme; $\mathrm{pga}=$ posterior portion of gonocoxal apodeme.

Body size: 2.0 mm .
Female. Unknown.
Genetic distances. Only one specimen was successfully sequenced. The nearest neighbour is Psl. sp. I, diverging by a minimum of $4.86 \%$. For the 28 S sequence the nearest neighbour is Psl. horribilis, diverging by a minimum of $0.25 \%$.

Etymology. The newly described species is named after the Kaikoura peninsula, where both specimens were caught.

Distribution. New Zealand.

Discussion. The reduction to one gonostylar spine-as in this new species-is an infrequent character state among the species of Pseudolycoriella. Outside of New Zealand only a few species are known to have such a reduction: Pseudolycoriella curviseta Mohrig, Psl. defluviata Rudzinski, Psl. japonensis (Mohrig \& Menzel), Psl. patronata Rudzinski, Psl. pollicis (Pettey) [for this species Mohrig et al. (2013) stated two narrow subequal spines, while Mohrig \& Kauschke (2019) only gave one spine], Psl. pugionata Rudzinski, Psl. microcteniuni (Yang \& Zhang), Psl. morenae (Strobl), Psl. pendleburyi (Edwards), Psl. semialata (Edwards), and Psl. subbruckii (Mohrig \& Hövemeyer). None of these species has a prominent edge in the middle of the inner side of the gonostylus. Therefore, confusion with Psl. kaikoura can be excluded. Among the New Zealand representatives of the genus the species of the Psl.jejuna complex also bear only one spine on the gonostylus. Nevertheless, Psl. kaikoura can easily be distinguished from them by its uniquely shaped gonostylus and its different tegmen structure. Pseudolycoriella kaikoura can be assigned to the Psl. bruckii group.

## Pseudolycoriella sp. I

Literature. Pseudolycoriella indet.: BOLD (2019b).

Barcoded material. 1 q New Zealand: North Island, Waikato, Hamilton, altitude 39 m , Malaise trap, 05.06.2012, leg. I. Hogg (CBG, BIOUG14863-H041). 1 q same locality as previous, Malaise trap, 19.11.2012, leg. I. Hogg (CBG, BIOUG15289-H10). 1 Q same locality as previous, Malaise trap, 11.03.2013, leg. I. Hogg (CBG, BIOUG15552D06). 1 q same locality as previous, Malaise trap, 08.04 .2013 , leg. I. Hogg (CBG, BIOUG15560-H06).

Genetic distances. The maximum p-distance between all four available COI sequences is $0.17 \%$. The nearest neighbour is Psl. kaikoura, diverging by a minimum of $4.86 \%$. No 28 S sequence data are available.

Discussion. The COI barcodes of Psl. sp. I are registered on the Barcode of Life Data System under the BIN: BOLD:ACP1302 (BOLD 2019b). The p-distance of $4.86 \%$ towards the nearest neighbour suggests the existence of a further unknown species. Nevertheless, it was judged premature to prepare a formal description because no males are available and the lack of knowledge about the taxonomy of the females of Pseudolycoriella does not allow solid conclusions to be made on species delimitation.

## Pseudolycoriella zealandica complex

The Pseudolycoriella zealandica complex consists of probably three species-Pseudolycoriella zealandica, Psl. aotearoa, and the undescribed Psl. sp. II. Individuals have a relatively large body, and thus the largest New Zealand Pseudolycoriella specimens can be found in this complex. The males of the described species are characterized by two gonostylar spines, and the trapezoid tegmen with a broad hood-like structure ( $h s$ in Fig. 12). Their phylogenetic relationship is shown in Figure 61.

Discussion. The Psl. zealandica complex can be assigned to the Psl. bruckii group without any contradictions. It is not possible to state which morphological characters might be regarded as autapomorphies of this complex, because the undescribed species is located basally in the clade of the Psl. zealandica complex. Therefore, conclusions about the ground plan of the Psl. zealandica complex are not possible. It can only be stated that this complex comprises the largest species of Pseudolycoriella in New Zealand. Nevertheless, the monophyly is clearly indicated by genetic data.

## Pseudolycoriella zealandica (Edwards, 1927)

(Fig. 12)
Literature. Sciara zealandica Edwards, 1927: Tonnoir \& Edwards (1927): 796, Fig. 180.—Miller (1950): 57.—Davies (1988): 13.-Steffan (1989): 151.—Pseudolycoriella zealandica (Edwards, 1927): Mohrig \& Jaschhof (1999): 37, Fig. 18 a-g.Menzel \& Mohrig (2000): 715-716.—Rudzinski (2000): 183.—Macfarlane et al. (2010): 441

Material studied. Holotype male. New Zealand: North Island, Waikato, Okauia, Matamata, 18.11.1922, leg. A.E.

Brookes (BMNH, BMNH(E)250341). Previously published material: $7 \widehat{\top} \widehat{\jmath}$ North Island, Taupo, Hauhungaroa Range, 5 km southwest of Tihoi, Podocarpus wood with ground ferns, sweep net, 21.12.1992, leg. M. Jaschhof (6x PWMP; 1x SDEI). New records: 22才入 North Island, Ruapehu, Tongariro NP, Mangawhero River Valley 3 km NE Ohakune, 690 m , mixed podocarp/broadleaf forest, Malaise trap, 26.11-28.12.2002, leg. M. \& C. Jaschhof \& U. Kallweit (7x NZAC, SDEI-Dipt-0000622, SDEI-Dipt-0000624, SDEI-Dipt-0000998, SDEI-Dipt-0001002, SDEI-Dipt-0001243 \& SDEI-Dipt-0001246; 15x SDEI, SDEI-Dipt-0000623, SDEI-Dipt-0001001, SDEI-Dipt-0001239, SDEI-Dipt-0001242, SDEI-Dipt-0001244, SDEI-Dipt-0001248, SDEI-Dipt-0001254, SDEI-Dipt-0001538, SDEI-Dipt-0001539, SDEI-Dipt-0001540, SDEI-Dipt-0001541 \& SDEI-Dipt-0001542). 1 q same locality as prevoius, sweep net, 03-04.02.2002, leg. M. Jaschhof (SDEI, SDEI-Dipt-0000783). $9 \delta^{\top}{ }^{\top}, 2 q+$ South Island, Selwyn, Cass, Middle Bush, Nothofagus solandri forest, stream site, Malaise trap, 09.04 .1998 , leg. P.M. Johns (4x NZAC, SDEI-Dipt-0001166, SDEI-Dipt-0001176, SDEI-Dipt-0001186 \& SDEI-Dipt-0001189; 7x SDEI, SDEI-Dipt-0001168, SDEI-Dipt-0001183 [q], SDEI-Dipt-0001185, SDEI-Dipt-0001187, SDEI-Dipt-0001188, SDEI-Dipt-0001190 \& SDEI-Dipt-0001191 [q]). $1 \delta^{\lambda}, 1$ q same locality as previous, 11.04.1998, leg. P.M. Johns (all SDEI, SDEI-Dipt0001163 [ ${ }^{\lambda}$ ] \& SDEI-Dipt-0001162 [ q$]$ ]. $21 \widehat{o}^{\lambda}$ South Island, Southland, Fiordland, Fiordland NP, Hollyford River Valley, Moraine Creek Track, mixed podocarp/Nothofagus forest, Malaise trap, 05-24.01.2002, leg M. \& C. Jaschhof (8x NZAC, SDEI-Dipt-0000883, SDEI-Dipt-0001544, SDEI-Dipt-0001546, SDEI-Dipt-0001548, SDEI-Dipt-0001551, SDEI-Dipt-0001554, SDEI-Dipt-0001557 \& SDEI-Dipt-0001559; 13x SDEI, SDEI-Dipt-0000882, SDEI-Dipt-0000884, SDEI-Dipt-0000886, SDEI-Dipt-0001543, SDEI-Dipt-0001545, SDEI-Dipt-0001547, SDEI-Dipt-0001549, SDEI-Dipt-0001550, SDEI-Dipt-0001552, SDEI-Dipt-0001553, SDEI-Dipt-0001555, SDEI-Dipt0001556 \& SDEI-Dipt-0001558). $1{ }^{\top}$ South Island, Southland, Otago Lakes, Fiordland NP, Hollyford River Valley S Divide Creek, mixed Nothofagus/broadleaf forest, Malaise trap, 06-24.01.2002, leg. M. \& C. Jaschhof (SDEI).

Redescription. Male. Head. Head capsule brown. Eye bridge four facets wide, two to three facets at margin. Scape and pedicel slightly paler than the flagellomeres; fourth flagellomere 2.3-3.2 times longer than wide; necks of flagellomeres well differentiated; surface of flagellomeres rough, sensillae of two different lengths, small ones and longer, curved sensilla; setae on the flagellomere dense and as long as flagellomere width, slightly curved. Maxillary palp long and three-segmented, first palpomere longest, second one shortest; first palpomere with an inconspicuous patch of sensilla and five to eight long bristles, one of these bristles longer and more robust, located on the outer side of the first palpomere. Prefrons and clypeus bulging. Thorax brown, paler than head; some specimens with small brightened parts anteriorly and posteriorly. Posterior pronotum bare. Anterior pronotum with four to eight setae. Episternum 1 with five to 14 setae. Mesonotum with five to nine robust lateral bristles; row of dorsocentral bristles well developed. Scutellum with five to eight robust bristles and several minor setae. Katepisternum slightly longer than high. Wing. Length 2.4-2.9 mm; width/length ratio 0.35-0.39. Membrane smoky and without macrotrichia, anal area present; all posterior veins distinct, stem of $M$ weakest, apical half of $R_{5}$ with macrotrichia on ventral and dorsal side; $\mathrm{r}-\mathrm{m}$ and bM bare, exceptionally $\mathrm{r}-\mathrm{m}$ with one seta; $\mathrm{R}_{1} 0.7-0.9$ times as long as R ; $\mathrm{c} / \mathrm{w}$ ratio $0.73-0.82$; r-m $0.6-1.0$ as long as bM . Haltere short and sooty brown; knob as long as shaft. Legs pale, yellowish brown. Tibial organ surrounded by a circular border and wider than half width of tibial apex; tibial organ consists of a transverse row of bristles. Fore and mid tibia without robust bristles among vestiture. Hind tibia with two to seven robust bristles arranged in a longitudinal, posteriodorsal row. All tibial spurs equal in length. Claws with robust teeth, arranged in decreasing size. Abdomen brown, slightly paler than thorax, with dark setae. Hypopygium (Fig. 12). Gonocoxite wider than long, inner side of gonocoxites with a membranous area scattered with short pale setae, outer side with long setae; ventral, inner side of gonocoxites basally not fused. Gonostylus bean shaped, 2.1-2.8 times longer than wide, apically with a large inner cavity without any microtrichia, two robust spines and one long whip-lash hair present. Tegmen trapezoid with a strongly sclerotized base, parameral apodeme short and robust; medially with a broad hood-like structure ( $h s$ in Fig. 12) laterally flanked by a group of small teeth (not expressed in all specimens, 19 of 30 specimens from the North island and six of 33 from the South Island are without this character). Area of teeth consists of many small teeth. Ejaculatory apodeme dark, short and broad; base of ejaculatory apodeme long, basally widely fused, horseshoe-shaped. Posterior portion of gonocoxal apodeme medium sized and brown. Anterior portion of gonocoxal apodeme long and dark brown.

Body size: 2.1-3.8 mm.
Female. Tonnoir \& Edwards (1927) designated two female paratypes of Psl. zealandica. The metadata of one of those ( $\mathrm{BMNH}(\mathrm{E}) 250343$ ) are available online from the Natural History Museum (2014). However, those types are not included in the present study. Some descriptive information is given by Tonnoir \& Edwards (1927) and by Menzel \& Mohrig (2000).

Genetic distances. The maximum p-distance between all 17 available COI sequences is $2.43 \%$. The nearest neighbour is Psl. teo, diverging by a minimum of $10.63 \%$. All three available conspecific 28 S sequences are identical and show no differences to the sequences of Psl. zealandica and Psl. sp. II.

Remarks. All previous studies containing descriptions of the morphology of Psl.zealandica (Tonnoir \& Edwards 1927; Mohrig \& Jaschhof 1999; Menzel \& Mohrig 2000) did not mention the groups of additional lateral teeth on both sides of the median structure on the tegmen (Fig. 12). In fact, the holotype does not have this unique feature, and even among the material available to Mohrig \& Jaschhof (1999), this feature is rarely present. The quality of the preparation also influences whether the teeth are easy to observe or not. Removal of tergite IX increases the possibility of observing teeth. In addition, specimens from New Zealand's South Island - examined in the present study for the first time-have a higher expression rate of this character and a larger number of teeth, when present. The presence of these lateral teeth is not homogenously distributed across both populations (chi-square test of homogeneity, $\chi^{2} \approx 13.38, \alpha<0.005$ ).

## Pseudolycoriella aotearoa sp. n.

(Figs 3 \& 13)
urn:lsid:zoobank.org:act:02F615B4-A7CB-4F4C-8736-1D568B686260
Literature. Pseudolycoriella zealandica (Edwards, 1927): Mohrig \& Jaschhof (1999): 37 [misidentification].

Material studied. Holotype male. New Zealand: North Island, Ruapehu, Tongariro NP, Mangawhero River Valley 3 km NE Ohakune, altitude 690 m , mixed podocarp/broadleaf forest, Malaise trap, 26.11-28.12.2002, leg. M. \& C. Jaschhof \& U. Kallweit (NZAC, SDEI-Dipt-0000997). Paratypes: $11 \widehat{\delta}^{\widehat{o}}$ same date and same locality as holotype (4x NZAC, SDEI-Dipt-0001000; 7x SDEI, SDEI-Dipt-0000625, SDEI-Dipt-0000758, SDEI-Dipt-0000978 \& SDEI-Dipt-0000999). $2 \AA^{\top}$ đ North Island, Taupo, Hauhungaroa Range, 5 km southwest of Tihoi, Podocarpus wood with ground ferns, sweep net, 21.12.1992, leg. M. Jaschhof (all PWMP [previously misidentified, published as Psl. zealandica in Mohrig \& Jaschhof (1999)]).

Description. Male. Head. Head capsule brown. Eye bridge four facets wide, two to three facets at margin. Scape and pedicel slightly paler than the flagellomeres; fourth flagellomere (Fig. 3) 2.3-2.6 times longer than wide; necks of flagellomeres well differentiated; surface of flagellomeres rough, sensilla of two different lengths, small ones and longer, curved sensilla; setae on the flagellomere dense and as long as flagellomere width, slightly curved. Maxillary palp long and three-segmented, first palpomere longest, second one shortest; first palpomere with an inconspicuous patch of sensilla and four to eight long bristles, one of these bristles longer and more robust, located on the outer side. Prefrons and clypeus bulging. Thorax brown, paler than head; some specimens with small anterior and posterior brightened parts. Posterior pronotum bare. Anterior pronotum with seven to eleven setae. Episternum 1 with four to eleven setae. Mesonotum with six to eleven robust lateral bristles; row of dorsocentral bristles well developed. Scutellum with six to nine robust bristles and several minor setae. Katepisternum slightly longer than high. Wing. Length $2.6-3.3 \mathrm{~mm}$; width/length ratio $0.34-0.38$. Membrane smoky and without macrotrichia, anal area present; all posterior veins distinct, stem of $M$ weakest, apical half of $R_{5}$ with macrotrichia on ventral and dorsal side; bM bare, r-m bare or with up to two setae; $\mathrm{R}_{1} 0.8-1.0$ times as long as R ; $\mathrm{c} / \mathrm{w}$ ratio $0.70-0.81$; r-m $0.7-0.8$ as long as bM. Haltere short and sooty brown; knob as long as shaft, or slightly longer than shaft. Legs pale brown. Tibial organ surrounded by a circular border and wider than half width of tibial apex; tibial organ consists of a transverse row of bristles. Fore and mid tibia without robust bristles among vestiture. Hind tibia with two to seven robust bristles organised in a longitudinal, posteriodorsal row. All tibial spurs equal in length. Claws with robust teeth, arranged in decreasing size. Abdomen brown, slightly paler than thorax, with dark setae. Hypopygium (Fig. 13). Gonocoxites wider than long, inner side of gonocoxites with a membranous area scattered with short pale setae, outer side with long setae; ventral, inner side of gonocoxites basally not fused. Gonostylus bean shaped, 2.1-2.4 times longer than wide, apically with a large inner cavity without any microtrichia, two robust spines and one long whip-lash hair present. Tegmen trapezoid with a strongly sclerotized base, parameral apodeme short and robust; medially with a broad hood-like structure. Area of teeth consists of many small teeth. Ejaculatory apodeme dark, short and narrow; base of ejaculatory apodeme long and narrow. Posterior portion of gonocoxal apodeme medium sized and brown. Anterior portion of gonocoxal apodeme short, comma-shaped, and dark brown.

Body size: 3.2-4.4 mm.
Female. Unknown.


## 13



FIGURES 12-13. Pseudolycoriella zealandica complex, hypopygia. 12. Pseudolycoriella zealandica (Edwards). 13. Pseudolycoriella aotearoa $\mathbf{\text { sp. }}$. Abbreviation: $\mathrm{hs}=$ hood-like structure.

Genetic distances. The maximum p-distance between all six available COI sequences is $0.76 \%$. The nearest neighbour is Psl.zealandica, diverging by a minimum of $12.21 \%$. One successfully obtained 28 S sequence is identical to those of Psl. zealandica and Psl. sp. II.

Etymology. Aotearoa is the Māori word for New Zealand and was chosen as an epithet for the newly described species to underline the close phylogenetic relationship to Psl. zealandica.

Distribution. New Zealand.
Discussion. Pseudolycoriella aoteraoa is the sister species of Psl. zealandica, and therefore morphologically very similar. However, in general Psl. aoteraoa has a larger a body size and a greater wing length. The discrimination of both species, relying on these characters only, is not sufficient, because both ranges show overlaps. The lengths of the gonostyli of both species do not show identical values, but are very close together (170-200 $\mu \mathrm{m} v s$. $210-230 \mu \mathrm{~m}$ ). A more reliable character is the structure of the base of the ejaculatory apodeme. Pseudolycoriella aoteraoa has a base consisting of two long and slender branches while Psl. zealandica has broader and widely connected branches. Further, the tegmen shows differences. The basal width of the tegmen is wider in Psl. aoteraoa and the middle hood-like structure is also broader. Lateral teeth on the tegmen as found in some specimens of Psl. zealandica are absent in Psl. aotearoa.

Due to its large body size Psl. aotearoa also resembles Psl. skusei Mohrig, Kauschke \& Broadley and Psl. tenebrioalata Mohrig. Nevertheless, the structure of the hypopygia differs significantly. Pseudolycoriella skusei has an elongated gonostylus with a sharp apical bend and more spines and more whip-lash hairs than Psl. aotearoa (Mohrig et al. 2016). Pseudolycoriella tenebrioalata shows a similar gonostylar shape and a similar arrangement of spines. However, the tegmen structure and the ejaculatory apodeme differ fundamentally (Mohrig 2013). Pseudolycoriella tenebrioalata has a tegmen that is just slightly tapered towards the apex, and the ejaculatory apodeme is very short. Furthermore Psl. tenebrioalata does not have a border on its tibial organ, and its claws lack any teeth (Mohrig 2013).

## Pseudolycoriella sp. II

Material studied. 1q, New Zealand: South Island, Canterbury, Selwyn, Cass, Middle Bush, 17.11.1998, Malaise trap, leg. P.M. Johns (SDEI, SDEI-Dipt-0000745).

Genetic distances. The nearest neighbour of the COI-sequence of Psl. sp. II is Psl. orite, diverging by a minimum of $10.89 \%$. The one successfully sequenced 28 S sequence is identical to those of Psl. zealandica and Psl. aotearoa.

Discussion. The molecular analysis of the female specimen SDEI-Dipt-0000745 revealed an isolated cluster for all three genetic markers, and consequently a well separated cluster in the consensus tree shown in Figure 61. Therefore, the presence of a new and undescribed species is very likely, but without conspecific male specimens a reasonable description allowing meaningful comparisons with other Pseudolycoriella species is not possible.

## Pseudolycoriella bispina complex

The following six species form the Psl. bispina complex. Common to them is a small body size, and presence of lateral lobes on the tegmen. The phylogeny of this complex is depicted in Figure 61.

Discussion. This complex can be assigned to the Psl. bruckii group. Nevertheless, due to its small body size and the two spines on the gonostylus the species of the Psl. bispina complex resemble the Psl. longicostalis group from Papua New Guinea (Mohrig 2013). Furthermore, some of those species share the very robust bristles arranged in a posteriodorsal row on the hind tibia. However, the two apicolateral lobes on the tegmen of Psl. bispina and the other species of this complex cannot be found in the Psl. longicostalis group. As an additional character Mohrig (2013) emphasised that these species have very robust bristles in the posteriodorsal row on the hind tibia, which could also be found in the Psl. bispina complex, except in Psl. mahanga. Nevertheless, all these species are not assigned to the Psl. longicostalis group. The key characteristics like the conspicuous row of robust bristles on the hind tibia surely result from convergent miniaturisations of two independent stem species with a typical Psl. bruckii group habitus.

Conspicuous apicolateral lobes can be found in other Pseudolycoriella species: for instance, Psl. commoda

Mohrig (Pseudolycoriella triacanthula group; Mohrig 2013), and Psl. microphalli Mohrig and Psl. bitorquia Mohrig (Psl. quadrispinosa group; Mohrig 2013). A confusion of those three species with Psl. bispina and its close relatives is not expected, because all of the Papua New Guinea species have three gonostylar spines and remarkable dorsal teeth on the claws. Among the New Zealand species, Psl. cavatica, Psl. kaikoura, and the species of the Psl. jejuna complex have similar lateral lobes on the tegmen, but these species differ either by the reduction of the number of gonostylar spines or the shape of the gonostylus together with stout flagellomeres. Although the monophyly of the Psl. bispina complex is highly supported by genetic data, no unequivocal apomorphic characters could be identified.

## Pseudolycoriella bispina Mohrig, 1999

(Figs $4 \& 14$ )

Literature. Pseudolycoriella bispina Mohrig, 1999: Mohrig \& Jaschhof (1999): 40-41, Fig. 20 a-g.-Macfarlane et al. (2010): 441.

Material studied. Holotype male. New Zealand: North Island, Wairoa, Urewera National Park, Huiarau Range 30 km southeast of Murupara, altitude $600-1,000 \mathrm{~m}$, Podocarpus-Nothofagus wood, sweep net, 23.12.1992, leg. M. Jaschhof (PWMP). Paratype. $1 \circlearrowleft$ same locality and same date as holotype (PWMP). New records: $1 \jmath^{\wedge}$ North Island, Thames-Coromandel, Coromandel, Kirikiri Saddle, Kaitarakiri Track, secondary mixed podocarp/broadleaf forest, sweep net, 09.02.2002, leg. M. Jaschhof (SDEI). 1 § North Island, Western Bay of Plenty, Katikati, 140 Wharawhara Road, bush area, pitfall trap, 14.02-05.03.2015, leg. P.A. Maddison (SDEI, SDEI-Dipt-0001418). $1 \delta^{\Uparrow}$ North Island, Western Bay of Plenty, Katikati, 449 Lund Road, Malaise trap, 25.07-08.08.2016, leg. P.A. Maddison (SDEI, SDEI-Dipt-0001251). $3 \circlearrowleft^{\top} \delta^{\lambda}$ North Island, Ruapehu, Tongariro NP, Mangawhero River Valley 3 km NE Ohakune, altitude 690 m , mixed podocarp/broadleaf forest, sweep net, 03-04.02.2002, leg. M. Jaschhof ( 2 x NZAC, SDEI-Dipt-0000946 \& SDEI-Dipt-0001526; 1x SDEI, SDEI-Dipt-0001522). 6 ô ${ }^{\lambda}$ North Island, Taupo, Pureora Forest Park, Waipapa Reserve, altitude 600 m , mixed podocarp/broadleaf forest, sweep net, 04-05.02.2002, leg. M. Jaschhof (3x NZAC, SDEI-Dipt-0000974, SDEI-Dipt-0000975 \& SDEI-Dipt-0001516; 3x SDEI, SDEI-Dipt0000844 , SDEI-Dipt-0001517 \& SDEI-Dipt-0001518). 1§ same locality as previous, sweep net, 24-25.11.2002 leg. M. Jaschhof (SDEI).

Redescription. Male. Head. Head capsule brown. Eye bridge three facets wide, two facets at margin. Scape and pedicel slightly paler than the flagellomeres; fourth flagellomere 3.1-3.4 times longer than wide (Fig. 4); necks of flagellomeres well differentiated; surface of flagellomeres rough, sparsely scattered with long setae arising from slight elevations, setae 1.5 times as long as flagellomere width; sensilla of two different lengths present, small ones and longer, curved sensilla. Maxillary palp long and three-segmented, first palpomere usually as long as third, second one shortest; first palpomere with an inconspicuous patch of sensilla and two to five bristles, one of these bristles longer and more robust, located on the outer side. Prefrons and clypeus bulging. Thorax brown, paler than head. Posterior pronotum bare. Anterior pronotum with three to five setae. Episternum 1 with three to eight setae. Mesonotum with four to five robust lateral bristles; row of dorsocentral bristles well developed. Scutellum with four robust bristles and several minor setae. Katepisternum slightly longer than high. Wing. Length $1.7-2.1 \mathrm{~mm}$; width/length ratio $0.35-0.39$. Membrane transparent and without macrotrichia, anal area present; all posterior veins distinct, stem of $M$ weakest, $\mathrm{R}_{5}$ without macrotrichia on ventral side; bM and $\mathrm{r}-\mathrm{m}$ bare; $\mathrm{R}_{1} 0.6-0.8$ times as long as R ; c/w ratio $0.61-0.72$; r-m $0.7-0.9$ as long as bM. Haltere short; knob as long as shaft. Legs pale, trochanter slightly darkened. Tibial organ surrounded by a circular border and wider than half width of tibial apex; tibial organ consists of an irregular transverse row of bristles and some bristles above the row. Fore tibia without robust bristles among the vestiture. Mid tibia mostly without robust bristles, exceptionally four specimens with one bristle among vestiture. Hind tibia with ten to 16 very robust bristles arranged in a longitudinal, posteriodorsal row; on posterior side of hind tibia one to six robust bristles among the vestiture. All tibial spurs equal in length. Claws with robust teeth, arranged in decreasing size. Abdomen brown with dark setae. Hypopygium (Fig. 14). Gonocoxites wider than long, inner side of gonocoxites basally widely separated. Gonostylus short, and curved, 1.9-2.4 times longer than wide, apically with a large inner cavity, dorsal inner margin also apically constricted; two robust spines present, the dorsal one located on a basal lobe and unusually straight; one long whip-lash hair is located on the ventral apex of the gonostylus. Tegmen broader than long, apicolaterally with two strongly sclerotized lobes, apex roundish
and surrounded by a thin sclerotized margin, only exceptionally with a small membranous area; parameral apodeme short and robust. Area of teeth consists of nine to 20 small teeth. Ejaculatory apodeme dark, of medium length and narrow; base of ejaculatory apodeme pale, lyre-shaped e.g. broadly fused basally with two medium long branches. Posterior portion of gonocoxal apodeme of medium size and brown, medially connected by a slender bridge. Anterior portion of gonocoxal apodeme long and broad.

Body size: 1.9-2.5 mm.
Female. Unknown.
Genetic distances. The maximum p-distance between all nine available COI sequences is $3.07 \%$. The nearest neighbour is Psl. mahanga, diverging by a minimum of $7.14 \%$. The maximum p-distance between all five available 28 S sequences is $0.12 \%$. The nearest neighbour is Psl. mahanga, diverging by a minimum of $0.06 \%$.

Distribution. New Zealand.
Remarks. One paratype of Psl. bispina, deposited in the NZAC, was not investigated in the present study and its species affiliation has not been revised. Nevertheless, it has to be mentioned, that it is labelled with another locality than published in Mohrig \& Jaschhof (1999). According to New Zealand Arthropod Collection (2019) the collection data are: one paratype, North Island, Taupo, Hauhungaroa Range, 5 km SW Tihoi, podocarp forest, sweep net, 21.12.1992, leg. Jaschhof (NZAC, NZAC02016076).

## Pseudolycoriella teo sp. n.

(Fig. 15)
urn:Isid:zoobank.org:act:NomenclaturalActs/5CAAD136-9980-4B4D-8536-B0A7034BFD41

Material studied. Holotype male. New Zealand: Taupo, Tongariro NP, Mangawhero River Valley 3 km NE Ohakune, 690 m , mixed podocarp/broadleaf forest; sweep net, 03-04.02.2002, leg. M. Jaschhof (NZAC, SDEI-Dipt-0001528).

Description. Male. Head. Head capsule brown. Eye bridge three facets wide, two facets at margin. Scape, pedicel and flagellomeres concolourous; fourth flagellomere 3.3 times longer than wide; necks of flagellomeres well differentiated; surface of flagellomeres slightly rough without deep pits, sparsely scattered with long setae, which are 1.5 times as long as flagellomere width; small sensilla and longer, curved sensilla present. Maxillary palp threesegmented; palpomeres inappropriately prepared in the holotype; first palpomere with four bristles, one of these bristles longer and more robust. Prefrons and clypeus slightly bulging. Thorax brown, paler than head, central parts brighter. Posterior pronotum bare. Anterior pronotum with two setae. Episternum 1 with five setae. Mesonotum with four robust lateral bristles; row of dorsocentral bristles present. Scutellum with four sockets, which formerly contained robust bristles. Katepisternum longer than high. Wing. Length 1.9 mm ; width/length ratio 0.36 . Membrane transparent and without macrotrichia, anal area well developed; all posterior veins distinct, stem of M weakest, $\mathrm{R}_{5}$ only with macrotrichia on the dorsal side; bM bare and r-m bare; $\mathrm{R}_{1} 0.6$ times as long as R ; $\mathrm{c} / \mathrm{w}$ ratio 0.59 ; r-m 0.9 as long as bM. Haltere short; knob as long as shaft. Legs pale. Tibial organ surrounded by a circular border and as long as half width of tibial apex; tibial organ consists of an irregular transverse row of bristles. Fore and mid tibia without robust bristles among vestiture. Hind tibia with seven very robust bristles arranged in a longitudinal, posteriodorsal row; on posterior side of hind tibia two robust bristles among vestiture. All tibial spurs equal in length. Claws strongly toothed. Abdomen concolourous with thorax. Hypopygium (Fig. 15). Gonocoxites wider than long, inner side of gonocoxites basally not fused. Gonostylus short, bean-shaped and only with a very slight curvation, 2.3 times longer than wide, apex rounded and with a small inner cavity; two robust spines present, not inserting on basal lobes; one whip-lash hair present on the ventral apex of the gonostylus. Tegmen broader than long, apicolaterally with two not very broad, strongly sclerotized lobes, apex membranous; parameral apodeme short and robust. Area of teeth consisting of approximately 15 small teeth. Ejaculatory apodeme brown, medium long and broad; base of ejaculatory apodeme delicate. Posterior portion of gonocoxal apodeme medium sized and brown. Anterior portion of gonocoxal apodeme long and broad.

Body size: 1.7 mm .
Female. Unknown.
Genetic distances. Because there is only a single specimen, no intraspecific distance is calculable. The nearest neighbour is Psl.jaschhofi, diverging by a minimum of $4.92 \%$. For the 28 S sequence the nearest neighbour is $P s l$. mahanga, diverging by a minimum of $0.13 \%$.

Etymology. The Māori adjective teo means small and was therefore deemed to be a suitable name for one of the smallest New Zealand Pseudolycoriella species.

Distribution. New Zealand.
Discussion. Pseudolycoriella teo has the smallest body size of the six species of Psl. bispina complex. Nevertheless, some specimens of Psl. orite have similarly small body sizes. In fact, this species is very similar to Psl. teo. The best distinguishing characters for Psl. teo are the shorter length of the gonocoxites and the very round apex of the gonostylus, which also has a small inconspicuous apical cavity.

## Pseudolycoriella puhihi sp.n.

(Figs 5 \& 16)
urn:lsid:zoobank.org:act:NomenclaturalActs/0C3972BD-BD5F-4A83-A049-03D6FC42CCB7
Material studied. Holotype male. New Zealand: North Island, Taupo, Tongariro NP, Mangawhero River Valley 3 km NE Ohakune, 690 m , mixed podocarp/broadleaf forest, by sweep net, 03-04.02.2002, leg. M. Jaschhof (NZAC, SDEI-Dipt-0001523). Paratypes. $2 \delta^{\top}$ North Island, Bay of Plenty, Katikati, 140 Wharawhara Road, bush area, pitfall traps, 14.02-05.03.2015, leg. P.A. Maddison (NZAC; SDEI, SDEI-Dipt-0001420); $\mathbf{\sigma}^{\top} \sigma^{\lambda}$ North Island, Bay of Plenty, Katikati, 140 Wharawhara Road, bush area, Malaise trap, 14.02-05.03.2015, leg. P.A. Maddison (NZAC, SDEI-Dipt-0001005 [without hypopygium]; SDEI, SDEI-Dipt-0001422). $1 \delta^{\lambda}$ North Island, Taupo, Hauhungaroa Range, 5 km southwest of Tihoi, Podocarpus wood with ground ferns, sweep net, 21.12.1992, leg. M. Jaschhof (PWMP [previously misidentified, published as Psl. bispina in Mohrig \& Jaschhof (1999); incorrect collection locality published in Mohrig \& Jaschhof (1999), slide with the printed label: "Neuseeland, Nordinsel / Hauhungaroa Range, / Tihoi / Podocarpus-Wald / 21.12.1992 Käscher / leg. Jaschhof / coll. Mohrig"]).

Description. Male. Head. Head capsule brown. Eye bridge three facets wide, two facets at margin. Scape and pedicel usually concolourous with flagellomeres, rarely with a slightly paler pedicel; fourth flagellomere 2.3-2.5 times longer than wide (Fig. 5); necks of flagellomeres well differentiated; surface of flagellomeres rough with deep pits, densely scattered with long setae, which are as long as flagellomere width; small sensilla and longer, curved sensilla present. Maxillary palp three-segmented, first palpomere mostly as long as third, second one shortest, first palpomere with three to five bristles, one of these bristles longer and more robust. Prefrons and clypeus slightly bulging. Thorax brown, paler than head, central parts brighter. Posterior pronotum bare. Anterior pronotum with three to seven setae. Episternum 1 with five to seven setae. Mesonotum with four to five robust lateral bristles; row of dorsocentral bristles present. Scutellum with four robust bristles and several minor setae. Katepisternum slightly longer than high. Wing. Length $1.5-1.8 \mathrm{~mm}$; width/length ratio $0.36-0.39$. Membrane transparent and without macrotrichia, anal area well developed; all posterior veins distinct, stem of $M$ weakest, apex of $R_{5}$ with up to five additional setae on the ventral side; bM and r-m bare; $\mathrm{R}_{1} 0.6-0.7$ times as long as R ; $\mathrm{c} / \mathrm{w}$ ratio $0.65-0.71$; r-m $0.9-1.2$ as long as bM . Haltere short; knob as long as shaft. Legs pale. Tibial organ surrounded by a circular border and as long as half width of tibial apex; tibial organ consists of an irregular transverse row of bristles. Fore tibia without robust bristles among vestiture. Mid tibia with one to two bristles among vestiture. Hind tibia with eight to ten robust bristles organised in a longitudinal, posteriodorsal row; on posterior side of hind tibia one to four robust bristles among vestiture. All tibial spurs equal in length. Claws strongly toothed. Abdomen concolourous with thorax. Hypopygium (Fig. 16). Gonocoxites wider than long, inner side of gonocoxites not fused basally. Gonostylus short, bean-shaped, and slightly curved, 1.9-2.2 times longer than wide, inner median edge strongly concave, apex blunt and with an inner cavity; two robust spines present, not inserted on basal lobes; one whip-lash hair present on the ventral apex of the gonostylus; on inner median site with conspicuous tendon insertion ( $t i$ in Fig. 16). Tegmen broader than long, apicolaterally with two not very broad, strongly sclerotized lobes, apex with a membranous edge; parameral apodeme short and robust. Area of teeth consists of approximately 15 small teeth. Ejaculatory apodeme brown, of medium length and narrow; base of ejaculatory apodeme delicate with thin branches. Posterior portion of gonocoxal apodeme of medium size and brown. Anterior portion of gonocoxal apodeme long and narrow.

Body size: 1.8-2.2 mm.
Female. Unknown.
Genetic distances. The maximum p-distance between all four available COI sequences is $4.44 \%$. The nearest neighbour is Psl. orite, diverging by a minimum of $7.54 \%$. Both available 28 S sequences are identical and show no differences to a sequence of Psl. orite.

Etymology. Pūhihi is the Māori word for antenna. The epithet refers to the distinctive flagellomeres compared to the other species closely related to Psl. bispina.

Distribution. New Zealand.
Discussion. Among the species of the Psl. bispina complex, Psl. puhihi is the only one where the length to width ratio of the fourth flagellomere is less than three. This ratio results from the greater width of the flagellomeres (Fig. 5). In addition, the flagellomeres are much more densely setose than in the other five species of this species complex. Furthermore, the length of the wing is shorter than in most other species around Psl. bispina ( $\leq 1.8 \mathrm{~mm}$ ). The only exception that was examined is one specimen of Psl. bispina (SDEI-Dipt-0001418), which also had a short wing length ( $<1.8 \mathrm{~mm}$ ).

## Pseudolycoriella orite sp. n.

(Fig. 17)
urn:lsid:zoobank.org:act:FB195DA5-5D08-4CE0-990C-111633F9F7E1

Material studied. Holotype male. New Zealand: North Island, Ruapehu, Tongariro NP, Mangawhero River Valley 3 km NE Ohakune, altitude 690 m , mixed podocarp/broadleaf forest, sweep net, 03-04.02.2002, leg. M. Jaschhof (NZAC, SDEI-Dipt-0000863). Paratypes: $8 \delta^{\Uparrow}$ § same locality and same date as holotype (3x NZAC, SDEI-Dipt-0001519, SDEI-Dipt-0001527 \& SDEI-Dipt-0001529; 5x SDEI, SDEI-Dipt-0001520, SDEI-Dipt-0001521, SDEI-Dipt-0001524, SDEI-Dipt-0001525 \& SDEI-Dipt-0001533). $1 \circlearrowleft^{\top}$ North Island, Ruapehu, Tongariro NP, Mangawhero River Valley 7 km NE Ohakune, mixed Nothofagus/podocarp forest, sweep net, 03.02.2002, leg. M. Jaschhof (SDEI). $3 \delta^{\lambda} \delta^{\lambda}$ North Island, Masterton, Tararua Forest Park, Blue Range Hut Track 10 km SW Mt. Bruce, mixed Nothofagus/podocarp/broadleaf forest, sweep net, 02.02.2002, leg. M. Jaschhof (2x NZAC; 1x SDEI).

Description. Male. Head. Head capsule brown. Eye bridge three facets wide, two facets at margin. Scape and pedicel concolourous with flagellomeres; fourth flagellomere 3.0-3.6 times longer than wide; necks of flagellomeres well differentiated; surface of flagellomeres rough with deep pits, sparsely scattered with long setae, which are 1.5 times as long as flagellomere width; small sensilla and longer, curved sensilla present. Maxillary palp three-segmented, first palpomere mostly as long as third, second one shortest, first palpomere with two to four bristles, one of these bristles longer and more robust. Prefrons and clypeus slightly bulging. Thorax brown, paler than head, central parts brighter. Posterior pronotum bare. Anterior pronotum with two to four setae. Episternum 1 with three to six setae. Mesonotum with four robust lateral bristles; row of dorsocentral bristles present. Scutellum with four robust bristles and several minor setae. Katepisternum longer than high. Wing. Length $1.8-2.2 \mathrm{~mm}$; width/length ratio $0.34-0.36$. Membrane transparent and without macrotrichia, anal area weakly developed; all posterior veins distinct, stem of M weakest, $\mathrm{R}_{5}$ only with setae on the dorsal side; bM and r-m bare; $R_{1} 0.5-0.7$ times as long as R ; $\mathrm{c} / \mathrm{w}$ ratio $0.66-0.70$; r-m 0.5-1.0 as long as bM. Haltere short; shaft slightly longer than knob. Legs pale. Tibial organ surrounded by a circular border and as long as half of the width of the tibial apex; tibial organ consists of an irregular transverse row of bristles. Fore tibia without robust bristles among vestiture. Mid tibia mostly with one bristle among vestiture. Hind tibia with eight to nine very robust bristles arranged in a longitudinal, posteriodorsal row; on posterior side of hind tibia one to three robust bristles among vestiture. All tibial spurs equal in length. Claws strongly toothed. Abdomen concolourous with thorax. Hypopygium (Fig. 17). Gonocoxites wider than long, inner side of gonocoxites not fused basally. Gonostylus short, bean-shaped, and slightly curved, 2.0-2.3 times longer than wide, on inner median side with a clearly developed tendon insertion, apex blunt and with a large apical cavity; two robust spines present, dorsal spine on a small basal lobe; one whip-lash hair present on the ventral apex of the gonostylus. Tegmen broader than long, apicolaterally with two long, strongly sclerotized lobes, apex with a small membranous edge; parameral apodeme robust, short to medium long. Area of teeth with six to twelve small teeth. Ejaculatory apodeme brown, medium long and broad; base of ejaculatory apodeme delicate with two thin branches. Posterior portion of gonocoxal apodeme medium sized and brown. Anterior portion of gonocoxal apodeme long and broad.

Body size: 1.8-2.1 mm.
Female. Unknown.
Genetic distances. The maximum p-distance between all six available COI sequences is $0.39 \%$. The nearest neighbour is Psl.jaschhofi, diverging by a minimum of $6.35 \%$. The p-distance between both available 28 S sequences is $0.12 \%$. One of those sequences is identical to one sequence of Psl. puhihi.

Etymology. In Māori the word ōrite means being equal or being similar. The epithet indicates the morphologi-
cal similarity of the newly described species to its sibling species．
Distribution．New Zealand．
Discussion．It is not easy to distinguish this new species from its relatives within the Psl．bispina complex． However，Psl．bispina and Psl．mahanga show diagnostic differences in the apical edge of the tegmen（fully scle－ rotized $v s$ ．membranous）and the base of the ejaculatory apodeme（lyre－shaped $v s$ ．narrow and bifurcated along the whole length）．Pseudolycoriella puhihi can be distinguished by the shorter and wider fourth flagellomere and the more clearly developed tendon insertion of the gonostylus adductor．Pseudolycoriella teo has shorter gonocoxites and gonostyli，and a lower c／w ratio（ $0.59 \mathrm{vs} . \geq 0.66$ ）．The new species and Psl．porehu are the most similar species of this complex．Of all members，both species have the narrowest wings，and therefore the lowest ratio of width to length of wings and the least developed anal area．In order to distinguish between these species，the body size must be taken into account．On average，Psl．porehu has a larger body size，but there is some overlap with Psl．orite．How－ ever，the wing length is always less than 2.2 mm in Psl．orite，and greater than or equal to 2.2 mm in Psl．porehu．The $\mathrm{c} / \mathrm{w}$ ratio also differs：it is less than 0.65 in Psl．porehu and greater than 0.65 in Psl．orite．Further minor distinguish－ ing characters are found in the robust bristles among the vestiture of the tibiae：on the mid tibia Psl．orite bears one （or no）robust bristle while Psl．porehu has three to five bristles；on the hind tibia the longitudinal，posteriodorsal row contains three to six bristles in Psl．porehu and eight to nine bristles in Psl．orite．Unfortunately，for Psl．porehu no genetic data are available，which could be used to verify this morphological species delimitation．

## Pseudolycoriella mahanga sp．n．

（Fig．18）
urn：lsid：zoobank．org：act：14AD5BCD－06F0－4CBC－BB21－8E519C072047
Material studied．Holotype male．New Zealand：South Island，Southland，Otago Lakes，Fiordland NP，Eglinton River Valley，opposite Gladehouse Track，Nothofagus forest，Malaise trap，01－21．12．2002，leg．M．\＆C．Jaschhof \＆U．Kallweit（NZAC，SDEI－Dipt－0001008）．Paratypes： 15 む ${ }^{\top}$ South Island，Westland， 5 km SW Arawhata， mixed Nothofagus／podocarp forest，aspirator，20．10．2001，leg．M．Jaschhof（7x NZAC；8x SDEI）． $1 \circlearrowleft^{\lambda}$ South Island， Queenstown Lakes，Otago Lakes，Mt Aspiring NP，Makarora River Valley near Cameron Creek Track，mixed Not－ hofagus／broadleaf forest，sweep net，18－19．12．2002，leg．M．Jaschhof（SDEI，SDEI－Dipt－0000595）． $2{ }^{\top}{ }^{\Uparrow}$ South Island，Southland，Fiordland，Fiordland NP， 3 km E Milford Sound，mixed podocarp／broadleaf forest，Malaise trap，01－21．12．2002，leg．M．\＆C．Jaschhof \＆U．Kallweit（NZAC；SDEI）． $1 \delta^{\pi}$ same locality as previous，sweep net，21．12．2002，leg．M．Jaschhof（NZAC）． $1 \delta^{\text {§ }}$ South Island，Southland，Fiordland，Fiordland NP， 4 km E Milford Sound，mixed broadleaf／Nothofagus forest，sweep net，07．01．2002，leg．M．Jaschhof（NZAC；SDEI－Dipt－0000904）． 10 South Island，Southland，Otago Lakes，Fiordland NP，Eglinton River Valley，opposite Earl Mountain Tracks， Nothofagus forest，Malaise trap，05－24．01．2002，leg．M．\＆C．Jaschhof（SDEI，SDEI－Dipt－0000849）．Addition－ al material： 5 ふす South Island，Kaikoura，Blue Duck Reserve，altitude $300-400 \mathrm{~m}$ ，mixed podocarp／Nothofagus solandri forest，sweep net，29．11．2001，leg．M．Jaschhof；U．Kallweit \＆A．Stark（SDEI）．

Description．Male．Head．Head capsule brown．Eye bridge three facets wide，two facets at margin．Scape and pedicel concolourous with flagellomeres；fourth flagellomere 3．0－3．5 times longer than wide；necks of flagellom－ eres well differentiated；surface of flagellomeres rough，sparsely scattered with long setae arising from slight eleva－ tions，setae 1.5 times as long as flagellomere width；sensilla of two different lengths present，small ones and longer， curved sensilla．Maxillary palp long and three－segmented，first palpomere slightly longer than third，second one shortest；first palpomere with an inconspicuous patch of sensilla and two to five bristles，one of these bristles longer and more robust，located on the outer side．Prefrons and clypeus bulging．Thorax brown，central parts brightened． Posterior pronotum bare．Anterior pronotum with three to seven setae．Episternum 1 with five to ten setae．Mesono－ tum with four to six robust lateral bristles；row of dorsocentral bristles well developed．Scutellum with four robust bristles（exceptionally five）and several minor setae．Katepisternum as long as high，or slightly longer than high． Wing．Length 2．0－2．4 mm；width／length ratio 0．35－0．42．Membrane transparent and without macrotrichia，anal area present；all posterior veins distinct，stem of $M$ weakest，apical quarter to apical third of $R_{5}$ with macrotrichia on both sides；bM and r－m bare； $\mathrm{R}_{1} 0.6-1.0$ times as long as R ； $\mathrm{c} / \mathrm{w}$ ratio $0.66-0.76$（specimens from Kaikoura show a c／w ratio from $0.53-0.60$ ）；r－m $0.7-1.1$ as long as bM（specimens from Kaikoura $0.9-1.5$ ）．Haltere short；knob as long as shaft．Legs pale brown，trochanter slightly darkened．Tibial organ surrounded by a circular border and wider than half width of tibial apex；tibial organ consists of an irregular transverse row of bristles and some bristles above
the row. Fore tibia without robust bristles among vestiture. Mid tibia without or exceptionally with one robust bristle among vestiture. Hind tibia with three to nine robust bristles organised in a longitudinal, posteriodorsal row; on posterior side of hind tibia one to three robust bristles among vestiture. All tibial spurs equal in length. Claws with robust teeth. Abdomen brown with dark setae. Hypopygium (Fig. 18). Gonocoxites wider than long, basal inner side of gonocoxites widely separated. Gonostylus short, curved, and hunchbacked, 1.7-2.3 times longer than wide, apically with a large inner cavity; two robust spines present, the dorsal one located on a basal lobe; one long whiplash hair is located on the ventral apex of the gonostylus. Tegmen wider than long, apicolaterally with two strongly sclerotized lobes, apex roundish and surrounded by a strongly sclerotized margin, only exceptionally with a small membranous area; parameral apodeme short and robust. Area of teeth consists of 15 to 30 small teeth. Ejaculatory apodeme dark, broad, and short to medium long; base of ejaculatory apodeme pale, lyre-shaped, branches of base of ejaculatory apodeme of different width. Posterior portion of gonocoxal apodeme medium sized and brown. Anterior portion of gonocoxal apodeme long and broad.

Body size: 1.9-2.7 mm.
Female. Unknown.
Genetic distances. The maximum p-distance between the two available COI sequences is $1.14 \%$. The nearest neighbour is Psl. bispina, diverging by a minimum of $7.14 \%$. Both available 28 S sequences are identical. The nearest neighbour is Psl. bispina, diverging by a minimum of $0.06 \%$.

Etymology. The Māori word māhanga-meaning twin or twins-was selected as the epithet, to illustrate that the new described species is one of the sibling species around Psl. bispina.

Distribution. New Zealand.
Discussion. Of all species of this complex, only Psl. mahanga and Psl. bispina have in common a lyre-shaped base of the ejaculatory apodeme and a strongly sclerotized apex of the tegmen. Pseudolycoriella mahanga can be distinguished from Psl. bispina by the setosity of vein $\mathrm{R}_{5}$. In Psl. mahanga macrotrichia are located on both the ventral and dorsal sides, whereas in Psl. bispina they occur only on the dorsal side. On the hind tibia Psl. mahanga has a less developed longitudinal row of robust bristles. Their number does not exceed nine and they are also smaller than in Psl. bispina.

Five males originating from the Blue Duck Reserve in the Kaikoura District differ significantly in $\mathrm{c} / \mathrm{w}$ ratio from remaining specimens ( $0.53-0.60 \mathrm{vs} .0 .66-0.76$ ). Also, the $\mathrm{r}-\mathrm{m} / \mathrm{bM}$ ratio is slightly higher in some specimens from Kaikoura ( $0.7-1.1 \mathrm{vs} .0 .9-1.5$ ). Genetic data on this population are not available and no further morphological differences have been found, so it was decided not to describe a further new species. Because of this uncertainty, these specimens were not assigned to the type series of Psl. mahanga.

## Pseudolycoriella porehu sp. n.

(Fig. 19)
urn:Isid:zoobank.org:act:NomenclaturalActs/0B09BE48-A529-4737-B03E-CF2ED15CFAF9
Material studied. Holotype male. New Zealand: South Island, Clutha, Catlins, Purakaunui Scenic Reserve, mixed podocarp/broadleaf/Nothofagus forest, sweep net, 03.01.2002, leg. M. Jaschhof (NZAC, SDEI-Dipt-0001394). Paratypes: $4 \delta^{\lambda}$ § same locality and same date as holotype (1x NZAC; 3x SDEI, SDEI-Dipt-0001485).

Description. Male. Head. Head capsule brown. Eye bridge three facets wide, two facets at margin. Scape and pedicel concolourous or slightly paler than flagellomeres; fourth flagellomere 3.3-3.9 times longer than wide; necks of flagellomeres well differentiated; surface of flagellomeres rough, sparsely scattered with long setae arising from slight elevations, setae 1.5 times as long as flagellomere width; sensilla of two different lengths present, small ones and longer, curved sensilla. Maxillary palp long and three-segmented, first palpomere as long as third, second one shortest; first palpomere with an inconspicuous patch of sensilla and two to three bristles, one of these longer and more robust, located on the outer side. Prefrons and clypeus bulging. Thorax pale brown to brown, central parts brighter. Posterior pronotum bare. Anterior pronotum with three to five setae. Episternum 1 with four to seven setae. Mesonotum with five to six robust lateral bristles; row of dorsocentral bristles well developed. Scutellum with four robust bristles and several minor setae. Katepisternum as long as high, or slightly longer than high. Wing. Length $2.2-2.3 \mathrm{~mm}$; width/length ratio $0.33-0.36$. Membrane transparent and without macrotrichia, anal area weakly developed; all posterior veins distinct, stem of $M$ weakest, apical fifth to apical third of $R_{5}$ with macrotrichia on ventral and dorsal side; bM and $\mathrm{r}-\mathrm{m}$ bare; $\mathrm{R}_{1} 0.6-0.7$ times as long as R ; $\mathrm{c} / \mathrm{w}$ ratio $0.52-0.61$; r-m $0.7-0.9$ as long as bM . Haltere short; knob as long as shaft. Legs pale brown, trochanter slightly darkened. Tibial organ surrounded by a

$100 \mu \mathrm{~m}$
FIGURES 14-19. Pseudolycoriella bispina complex, hypopygia. 14. Pseudolycoriella bispina Mohrig. 15. Pseudolycoriella teo sp. n. 16. Pseudolycoriella puhihi sp. n. 17. Pseudolycoriella orite sp. n. 18. Pseudolycoriella mahanga sp. n. 19. Pseudolycoriella porehu $\mathbf{~ s p . ~ n . ~ A b b r e v i a t i o n : ~} \mathrm{ti}=$ insertion of the tendon of the gonostylus adductor.
circular border and wider than half width of tibial apex; tibial organ consists of a patch of several robust bristles arranged in a transverse band. Fore tibia without robust bristles among vestiture. Mid tibia with three to five robust bristles among vestiture, located anterioventrally and posterioventrally. Hind tibia with four to seven robust bristles organised in a longitudinal, posteriodorsal row; on posterior side of hind tibia five to six robust bristles among vestiture. All tibial spurs equal in length. Claws with robust teeth. Abdomen brown with dark setae. Hypopygium (Fig. 19). Gonocoxites wider than long, basal inner side of gonocoxites widely separated. Gonostylus short, broad, and slightly curved; 2.0-2.3 times longer than wide, apex blunt with an inner cavity; two robust spines present, the dorsal one located on a basal lobe; one long whip-lash hair is located on the ventral apex of the gonostylus. Tegmen broader than long, apicolaterally with two strongly sclerotized lobes, apex roundish with a small membranous edge; parameral apodeme robust and short to medium long. Area of teeth with ten to 15 small teeth. Ejaculatory apodeme dark, broad, and of medium length; base of ejaculatory apodeme pale, branches broad and basally connected. Posterior portion of gonocoxal apodeme medium sized and brown. Anterior portion of gonocoxal apodeme long and broad.

Body size: 1.9-2.5 mm.
Female. Unknown.
Genetic distances. No genetic information available.
Etymology. Porehu is a Māori modifier and means mysterious. It was chosen as the epithet because the taxonomic rank of these specimens remained unclear to the author for a long time.

Distribution. New Zealand.
Discussion. See discussion paragraph for Psl. orite.

## Pseudolycoriella jejuna complex

The Psl.jejuna complex is a group of thirteen closely related and cryptic species. These species share a complex and synapomorph structure of their hypopygia. In Figures 20-22 a hypopygium of Psl. tewaipounamu is depicted. The gonostylus ( $g s$ ) is elongated and curved with an inwardly bent apex. The apex possesses one spine ( $s p$ ) and a whiplash hair ( $w h$ ) and-in most species-on the inward part a cavity ( $a c$ ). The tegmen is highly derivative compared with the tegmen of typical sciarids. Its general shape is onion-shaped: more or less roundish with an apical contraction (at). Adjacent to the apical contraction the tegmen has two lateral lobes $(l l)$ on the ventral side, which can easily be observed in light microscopy. In the middle of the ventral side of the tegmen a membranous area ( ma ) bordered by two median folds ( mf ) can easily be observed by using scanning electron microscopy. In light microscopy these delicate structures are hardly visible. At this membranous area a bulge of the base of the ejaculatory apodeme (be) can be observed. An area of teeth is absent. In light microscopy, the parameral apodemes ( $p a$ ) do not fuse medially. Instead they are stretched to the apex of the tegmen and form the apical contraction (at). On the dorsal side of the tegmen the parameral apodemes protrude and form a dorsal carina $(d c)$. This carina is bent around the apex and ends subapically on the ventral side of the tegmen. The bulges of the parameral apodemes are accompanied by a dorsal fold (df), which can also be observed in light microscopy. The phylogeny of this complex is shown in Figure 61.

Discussion. The species of the Psl.jejuna complex can be assigned to the Psl. bruckii group. The monophyly of this complex is strongly supported by genetics and can also be demonstrated by some autapomorphic morphological characters such as the extraordinary tegmen, the reduction of the number of gonostylar spines, and the secondary absence of an area of teeth. Due to this unique character set, all species of the Psl.jejuna complex can easily be recognized. Only Psl. fuscorubroides (Mohrig \& Blasco-Zumeta) from Spain has a similar tegmen (Mohrig \& BlascoZumeta 1996). It also possesses a dorsal carina that derives from the parameral apodemes. However, there are differences: the parameral apodemes are fused and not separated by a gap, as in the Psl.jejuna complex, and the carina in Psl.fuscorubroides arises vertically in the middle base of the tegmen. Furthermore, Psl.fuscorubroides possesses a large area of conspicous teeth (Mohrig \& Blasco-Zumeta 1996). Several other structures of the hypopygium are also disimilar. For instance, the gonostylus of Psl.fuscorubroides is armed with two short spines and has a conspicuous lobe on the dorsal margin and therefore differs significantly from the gonostyli of the Psl.jejuna complex. It is not clear whether the dorsal carina of the tegmen is a convergent or a homologous character. Maybe Psl. fuscorubroides is closely related to the Psl. jejuna complex, but it certainly does not belong to the crown group of the Psl. jejuna complex, since the absence of an area of teeth can be regarded as a synapomorpy of the New Zealand Psl.


FIGURES 20-24. Pseudolycoriella jejuna complex, Pseudolycoriella tewaipounamu sp. n., details of hypopygium. 20. Hypopygium, ventral. 21. Hypopygium, dorsal, tergite IX removed. 22. Hypopygium, ventral. 23. Tegmen, ventral. 24. Tegmen, dorsal. Abbreviations: ac = apical cavity; at = apex of tegmen; be = base of ejaculatory apodeme; be* = base of ejaculatory apodeme covered by the membranous area; dc = dorsal carina; $\mathrm{df}=$ dorsal fold; $\mathrm{ea}=$ ejaculatory apodeme; $\mathrm{gc}=$ gonocoxite; $\mathrm{gs}=$ gonostylus; $\mathrm{ll}=$ lateral lobe; $\mathrm{ma}=$ membranous area; $\mathrm{mf}=$ median fold; $\mathrm{pa}=$ parameral apodeme; $\mathrm{pa}^{*}=$ parameral apodeme covered by integument; $\mathrm{sp}=$ gonostylar spine; $\mathrm{wh}=$ whip-lash hair.
jejuna complex members. In other words, the hypothesis of a monophyly of the New Zealand representatives of the Psl.jejuna complex cannot be rejected.

The occurrence of two atavisms among the specimens of the Psl. jejuna complex confirms the above character polarisation regarding the single gonostylar tooth and the absence of the area of teeth. One paratype of Psl. dagae has two spines on the left gonostylus while the right one only bears one. This atavistic state is unique among all 233 investigated specimens belonging to the Psl. jejuna complex and indicates that the species of the Psl.jejuna complex derivate from an ancestor that bears two spines at the gonostylus, like the extant species of the Psl. bispina complex or the Psl. zealandica complex. The second detected atavism occured in two paratypes of Psl. whena, where an area of the teeth is present, although to a varying degree-one paratype shows only five tiny teeth, the other twelve.

## Pseudolycoriella jejuna (Edwards, 1927)

(Figs 25-29)

Literature. Sciara jejuna Edwards, 1927: Tonnoir \& Edwards (1927): 796, Fig. 181.—Miller (1950): 57.—Steffan (1989): 150.—Pseudolycoriella jejuna (Edwards, 1927): Mohrig \& Jaschhof (1999): 37-39, Fig. 19 a-g.-Menzel \& Mohrig (2000): 715.-Rudzinski (2000): 183.-Macfarlane et al. (2010): 441.

Material studied. Holotype male. New Zealand: North Island, Manawatu-Wanganui, Ohakune, Oct.-Nov. 1923, leg. T.R. Harris (BMNH, BMNH(E)250339). Previously published material: $1 \delta$ North Island, Far North, Waipoua Forest, 45 km northwest of Dargaville, old Kauri-Podocarpus wood, sweep net, 31.12.1992, leg. M. Jaschhof (PWMP). $2 \circlearrowleft^{\top} \delta^{\top}$ North Island, Taupo, Hauhungaroa Range, 5 km southwest of Tihoi, Podocarpus wood with ground ferns, sweep net, 21.12.1992, leg. M. Jaschhof (PWMP; SDEI [Slide with the handwritten label: "Neuseeland: Nordins. / Pureora Forest / Kescher, Podocarpus / 21.12.92 / leg. Jaschhof". The locality information is obviously incorrect, because the sample locality and the date do not correspond with locality information in Mohrig \& Jaschhof (1999).]). 2 §§ North Island, Wairoa, Urewera National Park, Huiarau Range 30 km southeast of Murupara, altitude $600-1,000 \mathrm{~m}$, Podocarpus-Nothofagus wood, sweep net, 23.12.1992, leg. M. Jaschhof (all PWMP). $1 \delta^{\top}$ North Island, Stratford, Mount Egmont National Park, East Egmont, altitude 650 m, Podocarpus wood with rotten wood and a dense layer of herbs, sweep net, 09.01.1993, leg. M. Jaschhof (PWMP). New records: $3 \delta^{\top} \delta^{\lambda}$ North Island, Taupo, Pureora Forest, Select Loop Road, 500 m , mixed mature podocarp forest, Malaise trap, 15.07-23.08.2001, leg. M. \& C. Jaschhof (SDEI, SDEI-Dipt-0000579, SDEI-Dipt-0000583 \& SDEI-Dipt-0000588). $1 \overbrace{}^{\lambda}$ North Island, Taupo, Pureora Forest Park, Waipapa Reserve, altitude 600 m , mixed podocarp/broadleaf forest, sweep net, 24-25.11.2002, leg. M. Jaschhof (SDEI). $2 \AA^{\AA} \delta^{\lambda}$ North Island, Ruapehu, Tongariro NP, Mangawhero River Valley 3 km NE Ohakune, altitude 690 m , mixed podocarp/broadleaf forest, sweep net, 03-04.02.2002, leg. M. Jaschhof (NZAC, SDEI-Dipt-0000773; SDEI, SDEI-Dipt-0000829). 8 đð same locality as previous, Malaise trap, 26.11-28.12.2002, leg. M. \& C. Jaschhof \& U. Kallweit (3x NZAC SDEI-Dipt-0000744 \& SDEI-Dipt-0001195 \& SDEI-Dipt-0001347; 5x SDEI, SDEI-Dipt-0000603, SDEI-Dipt-0000617, SDEI-Dipt-0001193 \& SDEI-Dipt-0001339). 2 § ${ }^{\Uparrow}$ same locality as previous, sweep net, 26.11.2002, leg. M. Jaschhof (NZAC; SDEI).

Redescription. Male. Head. Head capsule brown. Eye bridge three facets wide, two facets at margin. Scape and pedicel mostly slightly paler than the flagellomeres; fourth flagellomere 2.7-3.3 times longer than wide; necks of flagellomeres well differentiated; surface of flagellomeres rough with deep pits, sensilla of two different lengths, small ones and longer, curved sensilla; setae on the flagellomere as long as flagellomere width, slightly curved. Maxillary palp long and three-segmented, first palpomere longest, second one shortest; first palpomere with an inconspicuous patch of sensilla and three to five long bristles (exceptionally two), one bristle longer and more robust, located on the outer side. Prefrons and clypeus bulging. Thorax brown, paler than head; some specimens with lateral brightened areas. Posterior pronotum bare. Anterior pronotum with two to six setae. Episternum 1 with four to eight setae. Mesonotum with four to six robust lateral bristles; row of dorsocentral bristles well developed; anteriorly on the mesonotum two to seven (median four) small arcostichal setae (Figs 28-29). Scutellum with four robust bristles (the inner ones longer than the outer two) and several minor setae. Katepisternum as long as high. Wing. Length $2.3-3.1 \mathrm{~mm}$; width/length ratio $0.35-0.40$. Membrane transparent, slightly shaded and without macrotrichia, anal area present; all posterior veins distinct, except faint stem of M , apical one-fifth to half of $\mathrm{R}_{5}$ with macrotrichia on ventral and dorsal side; bM and r-m bare, exceptionally three specimens bearing one setae on $\mathrm{r}-\mathrm{m}$ (in two cases only
on one wing), $\mathrm{R}_{1} 0.8-1.2$ times as long as R ; $\mathrm{c} / \mathrm{w}$ ratio $0.75-0.82$; r-m $0.8-1.4$ as long as bM (the holotype has an extraordinarily short $\mathrm{r}-\mathrm{m}$ on the right wing, resulting in a $\mathrm{r}-\mathrm{m} / \mathrm{bM}$ ratio of 2.1 ). Haltere long and sooty brown; shaft longer than knob. Legs pale brown, paler than thorax, mid and hind coxae slightly darker than front coxae. Tibial organ surrounded by a circular border and as broad as half of the width of the tibial apex; tibial organ consists of an irregular row of bristles. Front tibia without robust bristles among the vestiture (exceptionally one specimen with one bristle among the vestiture). Mid tibia with one to three robust bristles among the vestiture (one specimen without robust bristles among vestiture). Posteriodorsal row of bristles inconspicuous, consisting of three to six bristles. All tibial spurs equal in length. Claws with robust teeth. Abdomen brown, slightly paler than thorax, with long, dark setae. Hypopygium (Fig. 25). Gonocoxites wider than long, inner side of gonocoxites with medium sized setae, outer side with long setae; ventral, inner side of gonocoxites not fused basally. Gonostylus elongate and curved, 2.4-3.1 times longer than wide, apex differentiated, apex width varies between slender and broad (compare Figs 25 \& 26), inner side of gonostylus concave and scattered with microtrichia; apex with one long spine and one whiplash hair. Tegmen of the typical ground plan of the Psl. jejuna complex, mostly onion-shaped (in some specimens more elongated), apical contraction broad; dorsal folds present; parameral apodeme basally strongly sclerotized, medially connected, posterior branches of the parameral apodemes nearly parallel. Area of teeth absent. Ejaculatory apodeme dark, broad, and long; base of ejaculatory apodeme very delicate, long, and slender (one specimen shows an exceptionally broad base [Fig. 27]). Posterior portion of gonocoxal apodeme medium sized and brown, medially fused. Anterior portion of gonocoxal apodeme long and dark brown.

Body size: 2.5-3.3 mm.
Female. A female paratype (BMNH(E)250340, BMNH) was designated by Tonnoir \& Edwards (1927) and described by Menzel \& Mohrig (2000). This type is not included in the present study.

Genetic distances. The maximum p-distance between all eight available COI sequences is $2.75 \%$. The nearest neighbour is Psl. teo, diverging by a minimum of $7.55 \%$. All ten available 28 S sequences are identical. The nearest neighbour is Psl. dagae, diverging by a minimum of $0.75 \%$.

## Distribution. New Zealand.

Remarks. Although Menzel \& Mohrig (2000) gave an accurate redescription of Psl.jejuna, some details have to be amended. These authors stated that Psl. jejuna possesses an area of teeth twice as high as broad, with long single-pointed teeth (Menzel \& Mohrig, 2000). This could not be verified because an area of teeth could not be observed in the holotype or in any other investigated specimen. Just two paratypes of another species of this complex—Psl. whena—show an atavistic area of teeth (see discussion of Psl. jejuna complex). Moreover, Menzel \& Mohrig (2000) gave a lower value for the body length of Psl. jejuna than measured in the present study ( 2.2 mm vs. 2.5-3.3 mm). Maybe this is a result of measuring the shrunken holotype without taking the length of the hypopygium into account. The holotype of Psl. jejuna has an extraordinary ratio of the length of the wing vein r-m and bM . On the right wing the $\mathrm{r}-\mathrm{m} / \mathrm{bM}$ ratio is 2.1 and on the left wing 1.8 . The $\mathrm{r}-\mathrm{m} / \mathrm{bM}$ ratio of the newly examined specimens ranges from 0.8 to 1.4.

## Pseudolycoriella tuakana sp. n.

(Figs 30-34)
urn:lsid:zoobank.org:act:NomenclaturalActs/9392599F-A28F-4DCC-B7AF-C558BCBCFCEB

Material studied. Holotype male. New Zealand: North Island, Manawatu-Wanganui, Tongariro NP, Mangawhero River Valley 3 km NE Ohakune, altitude 690 m , mixed podocarp/broadleaf forest, Malaise trap, 26.11-28.12.2002, leg. M. \& C. Jaschhof \& U. Kallweit (NZAC, SDEI-Dipt-0001194). Paratype. $1{ }^{\wedge}$ same locality and same date as holotype (SDEI, SDEI-Dipt-0001337).

Description. Male. Head. Head capsule brown. Eye bridge three facets wide, two facets at margin. Scape and pedicel slightly paler than the flagellomeres; fourth flagellomere 2.8-2.9 times longer than wide; necks of flagellomeres well differentiated; surface of flagellomeres rough with deep pits, sensilla of two different lengths, small ones and longer, curved sensilla; setae on the flagellomere as long as flagellomere width, slightly curved. Maxillary palp long and three-segmented, first palpomere longest, second one shortest; first palpomere with an inconspicuous patch of sensilla and two to four long bristles, one of these bristles longer and more robust, located on the outer side. Prefrons and clypeus bulging. Thorax brown, concolourous with head; laterally extensively brighter. Posterior pronotum bare. Anterior pronotum with five setae. Episternum 1 with three to four setae. Mesonotum with four to five robust lateral
bristles; row of dorsocentral bristles well developed; anteriorly on the mesonotum a patch of nine closely arranged arcostichal setae (Figs 33-34). Scutellum with three to four robust bristles and several minor setae. Katepisternum as long as high. Wing. Length $2.7-2.9 \mathrm{~mm}$; width/length ratio $0.39-0.40$. Membrane transparent, slightly shaded and without macrotrichia, anal area present; all posterior veins distinct, except faint stem of $M$, apical third of $R_{5}$ with macrotrichia on ventral and dorsal side; bM and r-m bare (the paratype bears on the right wing setae on r-m), $\mathrm{R}_{1} 1.0-1.1$ times as long as R; c/w ratio $0.77-0.80$; r-m 1.1-1.3 as long as bM. Haltere long and sooty brown; shaft longer than knob. Legs pale brown, mid and hind coxae slightly darker than fore coxae. Tibial organ surrounded by a circular border and slightly narrower than half width of tibial apex; tibial organ consists of an irregular row of bristles. Front tibia without robust bristles among the vestiture (the holotype bears on the left tibia one bristle among vestiture). Mid tibia without bristles among vestiture. Posteriodorsal row of bristles very inconspicuous, consisting of two to three bristles. All tibial spurs equal in length. Claws with robust teeth. Abdomen brown, slightly paler than thorax, with long, dark setae. Hypopygium (Fig. 30). Gonocoxites wider than long, inner side of gonocoxites with medium sized setae, outer side with long setae; ventral, inner side of gonocoxites basally not fused. Gonostylus elongate and curved, 2.9-3.0 times longer than wide, apex slightly differentiated, inner side of gonostylus concave and with scattered microtrichia; apex with one medium-sized spine that is only slightly longer than the surrounding setae (Figs $30 \& 31$ ); one whip-lash hair present. Tegmen of the typical ground plan of the Psl.jejuna complex, onion-shaped but elongated, apical contraction broad; dorsal folds well developed; parameral apodemes basally strongly sclerotized, median connected, posterior branches of the parameral apodemes nearly parallel and close. Area of teeth absent. Ejaculatory apodeme dark, broad, and short; base of ejaculatory apodeme very delicate, long, and broad. Posterior portion of gonocoxal apodeme medium sized and brown, medially joined. Anterior portion of gonocoxal apodeme of medium length and dark brown.

Body size: $3.0-3.2 \mathrm{~mm}$.
Female. Unknown.
Genetic distances. Both available COI sequences are identical and do not show any p-distance. The nearest neighbour is Psl. dagae, diverging by a minimum of $5.93 \%$. Both available 28 S sequences are identical. The nearest neighbour is Psl. dagae, diverging by a minimum of $0.11 \%$.

Etymology. The Māori word tuakana means elder brother or elder sister. Thus, the epithet refers to the close relationship of the new species to Psl.jejuna.

Distribution. New Zealand.
Discussion. Superficially Psl. tuakana is identical to Psl. jejuna and they would be regarded as conspecific, if the genetic analysis did not show a considerable difference between the species (Fig. 61). Given that both type specimens of Psl. tuakana originated from the same collection locality as Psl.jejuna the existence of one single species with two distinct genetic variants is highly unlikely. A subsequent closer examination revealed some slight morphological differences. Nevertheless, these have to be interpreted carefully in the light of the small number of specimens of Psl. tuakana, which does not enable a robust statistical analysis. Pseudolycoriella tuakana does not have any bristles among the vestiture on the mid tibia; while 21 out of 22 specimens of Psl. jejuna bear one to two (exceptionally three) bristles on the mid tibia. The spine on the gonostylus is shorter in Psl. tuakana (Fig. 35a) and therefore does not protrude from the surrounding setae as in Psl.jejuna. Another difference is the longer ejaculatory apodeme in Psl.jejuna (Fig. 35c). Nevertheless, there are exceptions: one specimen of Psl.jejuna (SDEI-Dipt-0000588) has an aberrant short ejaculatory apodeme (Fig. 27). The following weaker differences have more auxiliary character. The number of arcostichal bristles on the mesonotum differs between the species. Pseudolycoriella jejuna has two to seven arcostichal bristles while Psl. tuakana has nine (Figs 28, 29, 33, $34 \& 36$ ). The length to width ratio of the gonostylus of both species might show different median values, but a huge overlap has to be considered (Fig. 35b). Additionally, the tegmen of most Psl.jeju$n a$ specimens is more rounded while the tegmen of Psl. tuakana is more tapered.

## Pseudolycoriella dagae sp. n.

(Figs $1 \& 42$ )
urn:lsid:zoobank.org:act:8C8A28A2-ADE2-4D2F-9D16-01A2A4F788F2
Literature. Pseudolycoriella jejuna (Edwards, 1927): Mohrig \& Jaschhof (1999): 37 [misidentification].

Material studied. Holotype male. New Zealand: North Island, Western Bay of Plenty, Katikati, 449 Lund Road, Malaise trap, 25.07-08.08.2016, leg. P.A. Maddison (NZAC, SDEI-Dipt-0000679). Paratypes. $3 \circlearrowleft^{\lambda} \sigma^{\lambda}$ same locality
and same date as holotype (1x NZAC, SDEI-Dipt-0001249; 2x SDEI, SDEI-Dipt-0000628 \& SDEI-Dipt-0000655). $1 \circlearrowleft^{7}$ North Island, Western Bay of Plenty, Katikati, Te Mania catchment, Malaise trap, 19.09-27.10.2016, leg. P.A. Maddison (NZAC, SDEI-Dipt-0001367). $1 \delta^{\lambda}$ North Island, Wairoa, Urewera National Park, Huiarau Range 30 km southeast of Murupara, altitude $600-1,000 \mathrm{~m}$, Podocarpus-Nothofagus wood, sweep net, 23.12 .1992 , leg. M. Jaschhof (SDEI [previously misidentified, published as Psl. jejuna in Mohrig \& Jaschhof 1999]). 1ठ South Island, Kaikoura, Blue Duck Reserve, altitude 300-400 m, mixed podocarp-Nothofagus solandri forest, Malaise trap, 12.05-09.06.2001, leg M. \& C. Jaschhof (SDEI).

Description. Male. Head. Head capsule brown. Eye bridge three to four facets wide. Scape and pedicel yellow, strongly contrasting with the dark brown flagellomeres; fourth flagellomere (Fig. 1) 2.8-3.3 times longer than wide; necks of flagellomeres well differentiated and pale; surface of flagellomeres rough with deep pits, sensilla of two different lengths, small ones and longer, curved sensilla; setae on the flagellomere longer than flagellomere width, slightly curved. Maxillary palp long and three-segmented, first and third palpomeres of equal length, second one shortest, oval; first palpomere with long sensilla and two to five long bristles, one bristle longer and more robust, located on the outer side. Prefrons and clypeus bulging. Thorax as brown as head, lateral and frontal parts brightened. Posterior pronotum bare. Anterior pronotum with two to three setae. Episternum 1 with four to ten setae. Mesonotum with five to seven robust lateral bristles. Scutellum with three to four robust bristles and several minor setae. Katepisternum as long as high. Wing. Length $2.5-3.1 \mathrm{~mm}$; width/length ratio $0.38-0.40$. Membrane transparent, slightly shaded and without macrotrichia, anal area present; all posterior veins distinct, except faint stem of $M$, apical $40-67 \%$ of $R_{5}$ with macrotrichia on ventral and dorsal side (exceptionally: one paratype with macrotrichia only on the dorsal side); bM bare, the holotype and two paratypes bear one seta on r-m, the remaining three paratypes with a bare r-m; $\mathrm{R}_{1} 0.7-1.0$ times as long as R ; $\mathrm{c} / \mathrm{w}$ ratio $0.70-0.74$; r-m 1.0-1.4 as long as bM. Haltere long and sooty brown; head of haltere longer than shaft. Legs pale brown, paler than thorax, mid and hind coxae slightly darker than front coxae. Tibial organ surrounded by a circular border and slightly wider than half width of tibial apex; tibial organ consists of an irregular row of bristles. Front tibia without robust bristles among vestiture. Mid tibia with one to three robust bristles among vestiture. Posteriodorsal row of bristles inconspicuous. All tibial spurs equal in length. Claws with robust teeth. Abdomen brown, slightly paler than thorax, with long, dark setae. Hypopygium (Fig. 42). Gonocoxites wider than long, inner side of gonocoxites with medium sized setae, outer side with long setae; ventral, inner side of gonocoxites basally separated. Gonostylus elongated and curved, 2.8-3.2 times longer than wide, apex not well differentiated, inner side strongly concave, apical cavity with some microtrichia, remaining inner side with several microtrichia, one spine (exceptionally one paratype [SDEI-Dipt-0000655] with two gonostylar spines on the right gonostylus) and one whip-lash hair present, inner base of gonostylus with a conspicuous angle ( $b a$ in Fig. 42). Tegmen of the typical ground plan of the Psl.jejuna complex, onion-shaped, apical contraction broad; dorsal folds present; parameral apodemes basally strongly sclerotized, median connected. Area of teeth absent. Ejaculatory apodeme dark, broad, and long; base of ejaculatory apodeme pale, long, in some specimens y-shaped. Posterior portion of gonocoxal apodeme medium sized and brown, medial joined. Anterior portion of gonocoxal apodeme long and dark brown.

Body size: 2.5-3.2 mm.
Female. Unknown.
Genetic distances. All five available COI sequences are identical and do not show any pairwise distance. The nearest neighbour is Psl. tuakana, diverging by a minimum of $5.93 \%$. The p-distance between both available 28 S sequences is $0.06 \%$. The nearest neighbour is Psl. tuakana, diverging by a minimum of $0.11 \%$.

Etymology. The epithet dagae is an anagram of the German abbreviation of the German Society of general and applied Entomology (DGaaE).

## Distribution. New Zealand.

Discussion. Pseudolycoriella dagae is one of three species of this complex which have a yellowish scape and pedicel strongly contrasting with a dark flagellum. Another of those-Psl. porotaka-has a much shorter length/ width ratio of the fourth flagellomere (2.3-2.4 vs. 2.8-3.3 in Psl. dagae) and a much narrower ejaculatory apodeme. The remaining species-Psl. raki-can be distinguished by a more slender gonostylus shape, which manifests itself in a larger length to width ratio of the gonostylus (2.8-3.2 vs. 2.4-2.6 in Psl. dagae). In practice, however, discrimination between these two species can be as challenging as in Psl.jejuna and Psl. tuakana. In particular, preparations in which the gonostylus has not been mounted in a perfect ventral view can lead to misidentifications. A helpful additional character is a bulging basal angle at the inner base of gonostylus, which is well developed in Psl. dagae (Fig. 42) and absent or less conspicuous in Psl. raki. With respect to the wing, there are two other slight differences:

Psl. dagae has macrotricha on both the dorsal and ventral side for more than the apical half of vein $\mathrm{R}_{5}$, while in Psl. raki this double-sided setosity is limited to a maximum of the apical two-fifths of $\mathrm{R}_{5}$. The $\mathrm{c} / \mathrm{w}$ ratio of Psl. dagae is equal to or less than 0.74 , while in Psl. raki it is equal to or greater than 0.74 . Another slight difference, which should be treated with caution: the antepronotum of Psl. dagae bears $2-3$ setae while that of Psl. raki bears $3-8$ setae. One paratype of Psl. dagae has two gonostylar spines on the right gonostylus, which is regarded as an atavism (see Psl. jejuna complex discussion).

## Pseudolycoriella raki sp. n.

(Fig. 41)
urn:lsid:zoobank.org:act:NomenclaturalActs/01500E3C-735A-400F-968D-3B3E1F4A14C9

Literature. Pseudolycoriella jejuna (Edwards, 1927): Mohrig \& Jaschhof (1999): 37 [misidentification].—Köhler \& Mohrig (2016): 108 [misidentification].

Material studied. Holotype male. New Zealand: North Island, Far North, Waipoua Forest, alongside Highway 12, altitude 300 m , mixed mature Kauri/podocarp forest, Malaise trap, 24.07-17.08.2001, leg. M. \& C. Jaschhof (NZAC, SDEI-Dipt-0000834). Paratypes. 3 đ̋ same locality and same date as holotype (1x NZAC; 2x SDEI). 1 đ North Island, Far North, Puketi Forest, Waipapa River Valley, mixed secondary wood of Kauri and Podocarpus with tree fern, sweep net, 01-02.01.1993, leg. M Jaschhof (PWMP [previously misidentified, published as Psl.jejuna in Mohrig \& Jaschhof 1999]). $1 \delta^{\lambda}$ North Island, Thames-Coromandel, Coromandel Range, 5 km east of Coromandel, dense Podocarpus secondary wood with tree ferns, sweep net, 28.12.1992, leg. M. Jaschhof (PWMP [previously misidentified, published as Psl. jejuna in Mohrig \& Jaschhof (1999)]). $1 \circlearrowleft^{\lambda}$ North Island, Whakatane-Distrikt, White Pine Bush 3 km southwest of Whakatane, Podocarpus wood, sweep net, 26.12.1992, leg. M. Jaschhof (PWMP [previously misidentified, published as Psl.jejuna in Köhler \& Mohrig 2016]).

Description. Male. Head. Head capsule brown. Eye bridge three to four facets wide. Scape and pedicel bright yellow, strongly contrasting with the brown flagellum; fourth flagellomere 2.6-3.4 times longer than wide; necks of flagellomeres well differentiated, paler than body of flagellomeres, flagellomere necks at antenna apex show distal dark rings; surface of flagellomeres rough with deep pits, sensilla of two different lengths, small ones and longer, curved sensilla; setae on the flagellomere longer than the flagellomere width, slightly curved, arising from small elevations; apical flagellomeres very rough with prominent elevations. Maxillary palp very long and three-segmented, first or third palpomere longest, second one shortest; first palpomere with long sensilla and four to five long bristles, one bristle longer and more robust, located on the outer side. Prefrons and clypeus bulging. Thorax pale brown, slightly paler than head, with some indistinct lateral brightening. Posterior pronotum bare. Anterior pronotum with three to eight setae. Episternum 1 with two to eight setae. Mesonotum with four to five robust lateral bristles. Scutellum with three to four robust bristles and several minor setae. Katepisternum as long as high. Wing. Length $2.2-2.9 \mathrm{~mm}$; width/length ratio $0.38-0.41$. Membrane transparent and without macrotrichia, anal area present; all posterior veins distinct, except faint stem of $M$, apical fifth to two-fifths of $R_{5}$ with macrotrichia on ventral and dorsal side; bM and r-m bare; $\mathrm{R}_{1} 0.8-1.0$ times as long as R ; $\mathrm{c} / \mathrm{w}$ ratio $0.74-0.79$; r-m $0.8-1.2$ as long as bM. Haltere brownish; head of haltere slightly longer than shaft. Legs yellowish pale brown, paler than thorax. Tibial organ surrounded by a circular border and wider than half of the width of the tibial apex; tibial organ consists of an irregular row of bristles. Fore tibia with one robust bristle among the vestiture, or without. Mid tibia with one to three robust bristles among the vestiture. Posteriodorsal row of seven to eleven bristles occupies half to two-thirds of the apex of the tibia. All tibial spurs equal in length. Claws with robust teeth. Abdomen concolourous with thorax, with long, dark setae. Hypopygium (Fig. 41). Gonocoxites wider than long, inner side of gonocoxites with small setae, outer side with long setae; ventral, inner side of gonocoxites basally separated. Gonostylus curved and slender, 2.4-2.6 times longer than wide, apex not clearly separated, apical end broad, blunt, on ventral side with a huge apical cavity scattered with few microtrichia, one spine and one whip-lash hair present, inner base of gonostylus without or only with a minor angle. Tegmen of the typical ground plan of the Psl. jejuna complex, onion-shaped but more elongated than in Psl. jejuna; dorsal folds present; parameral apodeme basally strongly sclerotized, distal branches forming a triangle. Area of teeth absent. Ejaculatory apodeme dark, broad and long; base of ejaculatory apodeme delicate, long, u-shaped. Posterior portion of gonocoxal apodeme medium sized and brown, joined medially. Anterior portion of gonocoxal apodeme long and dark brown, widely separated.


FIGURES 25-36. Pseudolycoriella jejuna complex, Pseudolycoriella jejuna (Edwards) and Pseudolycoriella tuakana sp. n. 25. Psl. jejuna (SDEI-Dipt-0000583), hypopygium. 26. Psl. jejuna (SDEI-Dipt-0000603), gonostylus. 27. Psl. jejuna (SDEI-Dipt-0000583), tegmen with aberrant ejaculatory apodeme. 28. Psl. jejuna (SDEI-Dipt-0000603), mesonotum. 29. Psl. jejuna holotype, mesonotum. 30. Psl. tuakana holotype, hypopygium. 31. Psl. tuakana paratype, gonostylus. 32. Psl. tuakana paratype, tegmen. 33. Psl. tuakana holotype, mesonotum (lateral view). 34. Psl. tuakana paratype, mesonotum (lateral view). 35a. Comparison of the length of gonostylar spine of both species; values for holotypes are indicated by black asterisks. 35b. Comparison of the length-width ratio of gonostylus of both species; values for holotypes are indicated by black asterisks. 35c. Comparison of the length of ejaculatory apodeme of both species; values for holotypes are indicated by black asterisks. 36. Histogram of arcostichal bristles of both species. Abbreviations: $\mathrm{AC}=$ arcostichal bristles; $\mathrm{DC}=$ dorsocentral bristles.


39


41


40


FIGURES 37-42. Pseudolycoriella jejuna complex, hypopygia. 37. Pseudolycoriella sudhausi sp. n. 38. Pseudolycoriella whena sp. n. 39. Pseudolycoriella porotaka sp. n. 40. Pseudolycoriella nahenahe sp. n. 41. Pseudolycoriella raki sp. n. 42. Pseudolycoriella dagae sp. n. Abbreviation: ba = basal angle.

Body size: 2.3-2.9 mm.
Female. Unknown.
Genetic distances. Only one specimen was successfully sequenced. For the COI sequence, the nearest neighbour is Psl. tuakana, diverging by a minimum of $8.42 \%$. For the 28 S sequence the nearest neighbour is also Psl. tuakana, diverging by a minimum of $0.44 \%$.

Etymology. Raki is the Māori word for northern.
Distribution. New Zealand.
Discussion. See discussion paragraph for Psl. dagae.

## Pseudolycoriella hauta sp. n.

(Figs 9 \& 44)
urn:lsid:zoobank.org:act:0B34C8C6-9E06-40BA-8731-229742E97203
Material studied. Holotype male. New Zealand: South Island, Clutha, Catlins, Purakaunui Scenic Reserve, mixed podocarp/broadleaf/Nothofagus forest, Malaise trap, 27.01-05.03.2002, leg. M. \& C. Jaschhof (NZAC, SDEI-Dipt0001134). Paratypes. 3 ふ̃ふ same locality and same date as holotype (1x NZAC, SDEI-Dipt-0001137; 2x SDEI, SDEI-Dipt-0000506 \& SDEI-Dipt-0001142).

Description. Male. Head. Head capsule dark brown, laterally with several longer setae among the normal setosity. Eye bridge three to four facets wide. Scape and pedicel slightly paler brown than the flagellum; fourth flagellomere 2.5-2.8 times longer than wide; necks of flagellomeres well differentiated; surface of flagellomeres rough with deep pits, sensilla of two different lengths, small ones and longer, curved sensilla; setae on the flagellomere longer than flagellomere width, curved. Maxillary palp long and three-segmented, first palpomere is the longest, second one shortest; first palpomere with long sensilla and two to six long bristles, one bristle longer and more robust, located on the outer side. Prefrons and clypeus bulging. Thorax brown, mesonotum slightly darker. Posterior pronotum bare. Anterior pronotum with five to seven long and middle-sized setae. Episternum 1 with seven to nine long and medium sized setae. Mesonotum with six to seven robust lateral bristles. Scutellum with five to six robust bristles and several minor setae. Long dorsocentral bristles forming a complete row on the mesonotum. Katepisternum as long as high. Wing (Fig. 9). Length $2.6-2.8 \mathrm{~mm}$; width/length ratio $0.33-0.39$. Membrane transparent and without macrotrichia, anal area present; all posterior veins distinct, apical $33-50 \%$ of $\mathrm{R}_{5}$ with macrotrichia on ventral and dorsal side; bM bare, r-m with 1-3 macrotrichia (exceptionally r-m on one wing of one paratype bare); $\mathrm{R}_{1} 0.6-0.7$ times as long as R ; c/w ratio $0.70-0.75$; r-m $0.7-0.9$ as long as $b M$. Haltere brownish; head of haltere slightly longer than shaft. Legs brown, coxae as brown as thorax. Tibial organ surrounded by a circular border and as wide as half width of tibial apex; tibial organ consists of more than ten bristles arranged in an irregular row. Front tibia without robust bristle among vestiture. Mid tibia with one or two slightly robust bristles among vestiture. Posteriodorsal row of five to eight bristles mainly on middle of hind tibia. All tibial spurs equal in length. Claws with robust teeth. Abdomen brown, paler than thorax, with long, dark setae. Hypopygium (Fig. 44). Gonocoxites wider than long, inner side of gonocoxites with short setae, outer side with long setae; ventral, inner side of gonocoxites basally separated. Gonostylus slender, 2.7-3.3 times longer than wide, apical third curved inward, not distinctly delimited; inner side concave; apical cavity long and remaining inner side without microtrichia, ventral margin distinct; one spine present, located at the apex; long whip-lash hair present. Tegmen of the typical ground plan of the Psl.jejuna complex, with an acuminate basic shape; dorsal folds present; parameral apodemes basally strongly sclerotized, medially connected. Area of teeth absent. Ejaculatory apodeme short and dark, base of ejaculatory apodeme pale brown. Posterior portion of gonocoxal apodeme medium sized and brown, joined medially. Anterior portion of gonocoxal apodeme of medium-length and dark brown.

Body size: $2.7-3.2 \mathrm{~mm}$.
Female. Unknown.
Genetic distances. All four available COI sequences are identical and do not show any pairwise distance. The nearest neighbour is Psl. tewaipounamu, diverging by a minimum of $8.68 \%$. The nearest neighbour of the only available 28S sequence is Psl. tewaipounamu, diverging by a minimum of $0.17 \%$.

Etymology. Hauta is the Māori word for south. The name for this new species reflects the southern distribution of this species.

Distribution. New Zealand.

Discussion. Among the species of the Psl. jejuna complex examined during this study, Psl. hauta has the most developed setosity on the mesonotum, the anterior pronotum, the episternum 1, and the scutellum. Furthermore, the shape of the gonostylus is unique and allows a clear delimitation from the other species of the complex. The spine on the gonostylus lies at the inner angle of the gonostylus apex and is more apically located than in all the other species. Another distinguishing feature is the prominent ventral margin. Due to the more concave inner side of the gonostylus the ventral margin is more obvious than in the related species.

## Pseudolycoriella jejunella sp. n.

(Figs 2, 6, 8 \& 46)
urn:lsid:zoobank.org:act:06EB5442-74EE-4676-A0C1-2CC6F04C7EF9
Material studied. Holotype male. New Zealand: South Island, Southland, Fiordland, Fiordland NP, Hollyford River Valley, Moraine Creek Track, mixed podocarp/Nothofagus forest, Malaise trap, 05-24.01.2002, M. \& C. Jaschhof (NZAC, SDEI-Dipt-0000880). Paratypes. $1 \AA^{\lambda}$ same locality and same date as holotype (SDEI, SDEI-Dipt0000935). $1 才$ South Island, Southland, Otago Lakes, Fiordland NP, Hollyford River Valley S Divide Creek, mixed Nothofagus/broadleaf forest, Malaise trap, 06-24.01.2002, M. \& C. Jaschhof (SDEI).

Description. Male. Head. Head capsule brown, concolourous with thorax. Eye bridge one to three facets wide. Scape and pedicel not paler brown than the flagellum; fourth flagellomere (Fig. 2) 1.9-2.3 times longer than wide; necks of flagellomeres well differentiated; surface of flagellomeres rough with deep pits, sensilla of two different lengths, small ones and longer, curved sensilla; setae on the flagellomere as long as flagellomere width, slightly curved. Maxillary palp (Fig. 6) short and three-segmented, first palpomere is the longest, third one shortest, first palpomere with long sensilla and three to four long bristles, one bristle longer and more robust, located on the outer side. Prefrons and clypeus bulging. Thorax brown, mesonotum slightly darker. Posterior pronotum bare. Anterior pronotum with four setae. Episternum 1 with five setae. Mesonotum with five robust lateral bristles. Scutellum with two robust bristles and several minor setae. Katepisternum as long as high. Wing. Length $2.0-2.1 \mathrm{~mm}$; width/length ratio $0.38-0.41$. Membrane transparent and without macrotrichia, anal area present; all posterior veins distinct, stem of M weak; apical $33-40 \%$ of $\mathrm{R}_{5}$ with macrotrichia on ventral and dorsal side; r-m and bM without macrotrichia; $R_{1}$ approximately $0.6-0.7$ times as long as R ; c/w ratio 0.7 ; r-m 1.2-1.8 as long as bM. Haltere brownish; head of haltere as long as shaft. Legs brownish, coxae as brown as thorax. Tibial organ surrounded by a circular border and as wide as half width of tibial apex; tibial organ (Fig. 8) consists of approximately eight bristles arranged in an irregular row. Front tibia without robust bristles among the vestiture. Mid tibia with one or two slightly robust bristles. Posteriodorsal row of bristles on hind tibia present but inconspicuous. All tibial spurs equal in length. Claws with five robust teeth. Abdomen brown, concolourous with thorax; with long, dark setae. Hypopygium (Fig. 46). Gonocoxites wider than long, inner side of gonocoxites with short setae, outer side with long setae; ventral, inner side of gonocoxites basally separated. Gonostylus slender, 3.0-3.4 times longer than wide; apical fourth kinked inwards, apical cavity present, but inconspicuous, scattered with some microtrichia; one spine present, located subapically, medium long whip-lash hair present. Tegmen of the typical ground plan of the Psl. jejuna complex, with an acuminate basic shape; the dorsal folds strikingly developed; parameral apodemes basally strongly sclerotized. Area of teeth absent. Ejaculatory apodeme short and dark, base of ejaculatory apodeme not visible. Posterior portion of gonocoxal apodeme medium sized and brown, joined medially. Anterior portion of gonocoxal apodeme short and dark brown.

Body size: 1.9-2.5 mm.
Female. Unknown.
Genetic distances. Both available COI sequences are identical and do not show any distance. The nearest neighbour is Psl. maddisoni, diverging by a minimum of $10.79 \%$. The nearest neighbour of the single available 28 S sequence is Psl. whakahara, diverging by a minimum of $0.24 \%$.

Etymology. The epithet jejunella is the diminutive of jejuna, and refers to the smallness of the new species compared with Psl. jejuna.

Distribution. New Zealand.
Discussion. By its small body and wing length Psl. jejunella can be easily separated from most other species of this complex. Only specimens of Psl. whena and small specimens of Psl. sudhausi are similarly small. In addition, the structure of the gonostylus is unique in this complex. With a gonostylar index of 3.0-3.4, the gonostylus of Psl.
jejunella is more slender than in most other species. Only some individuals of Psl. dagae and Psl. hauta lie in this range. Furthermore, the gonostylus appears more tubular and does not show the typical cavity on the inner basal side, and the spine on the gonostylus is more subapically located than in other species.

## Pseudolycoriella maddisoni sp. n.

(Fig. 47)
urn:lsid:zoobank.org:act:FB0BF871-1F03-4675-AF10-AFC8DCB8C269

Material studied. Holotype male. New Zealand: North Island, Western Bay of Plenty, Katikati, 449 Lund Road, Malaise trap, 25.07-08.08.2016, leg. P.A. Maddison (NZAC, SDEI-Dipt-0000627). Paratypes. 2 § $\widehat{\text { N North Island, }}$ Western Bay of Plenty, Katikati, Te Mania catchment, Malaise trap, 19.09-27.10.2016, leg. P.A. Maddison (SDEI, SDEI-Dipt-0000664 \& SDEI-Dipt-0001369).

Description. Male. Head. Head capsule brown, slightly darker than thorax. Eye bridge two to three facets wide. Scape and pedicel pale brown, slightly paler than the flagellum; fourth flagellomere 2.7 times longer than wide (flagella are missing in both paratypes); necks of flagellomeres well differentiated, bright; surface of flagellomeres rough with deep pits, sensilla of two different lengths, small ones and longer, curved sensilla; setae on the flagellomere as long as flagellomere width, slightly curved. Maxillary palp long and three-segmented, first palpomere is the longest, second is shortest, first palpomere with long sensilla and four to six long bristles, one bristle longer and more robust, located on the outer side. Prefrons and clypeus bulging. Thorax dark brown. Posterior pronotum bare. Anterior pronotum with four to six setae. Episternum 1 with four to seven setae. Mesonotum with four to five robust lateral bristles. Scutellum with two to three robust bristles and several minor setae. Katepisternum as long as high. Wing. Length $2.5-2.7 \mathrm{~mm}$; width/length ratio $0.38-0.42$. Membrane transparent, slightly shaded and without macrotrichia, anal area present; all posterior veins distinct, stem of $M$ weak; apical $40-50 \%$ of $R_{5}$ with macrotrichia on both ventral and dorsal sides; r-m and bM without macrotrichia; $\mathrm{R}_{1} 0.7$ times as long as R ; $\mathrm{c} / \mathrm{w}$ ratio $0.74-0.80$; r-m 0.7-0.8 as long as bM. Haltere brownish; head of haltere as long as shaft. Legs brownish, slightly paler than thorax. Tibial organ surrounded by a circular border and as wide as half width of tibial apex; tibial organ arranged in an irregular row. Front tibia without or with one robust bristle among vestiture. Mid tibia with one or two slightly robust bristles. Posteriodorsal row of bristles extending two-thirds of apical tibia length. All tibial spurs equal in length. Claws with robust teeth. Abdomen pale brown, paler than thorax; with long, dark setae. Hypopygium (Fig. 47). Gonocoxites wider than long, inner side of gonocoxites with several medium sized setae, outer side with long setae; ventral, inner side of gonocoxites basally separated. Gonostylus swollen, 2.4-2.5 times longer than wide; apex well differentiated, bent inwards, apical cavity conspicuous without microtrichia; one spine on the gonostylus, located subapically, and one long whip-lash hair. Tegmen of the typical ground plan of the Psl.jejuna complex, onion-shaped and roundish; the dorsal folds strikingly developed; parameral apodemes basally strongly sclerotized, apically broad. Area of teeth absent. Ejaculatory apodeme short and dark; base of ejaculatory apodeme pale yellow, v -shaped. Posterior portion of gonocoxal apodeme of medium size and brown, medially joined. Anterior portion of gonocoxal apodeme medium long and dark brown.

Body size: 2.6-2.9 mm.
Female. Unknown.
Genetic distances. All three available COI sequences are identical and do not show any pairwise distance. The nearest neighbour is Psl. tewaipounamu, diverging by a minimum of $5.84 \%$. Both available 28S sequences are identical. The nearest neighbour is Psl. tewaipounamu, diverging by a minimum of $0.17 \%$.

Etymology. This new species is named after Peter A. Maddison to acknowledge his valuable collection work. Distribution. New Zealand.
Discussion. Among all species of the Psl.jejuna complex Psl. maddisoni and Psl. whakahara have the broadest gonostyli. However due to the hunchbacked shape of the gonostylus Psl. maddisoni is easy to identify.

## Pseudolycoriella tewaipounamu sp. n .

(Figs 20-24, 43)
urn:lsid:zoobank.org:act:NomenclaturalActs/CC9D30E2-C277-42B6-9BB0-257662D26B46

Material studied．Holotype male．New Zealand：South Island，Southland，Otago Lakes，Fiordland NP，Eglinton Riv－ er Valley，Deer Flat，Nothofagus forest，Malaise trap，01－20．12．2002，leg．M．\＆C．Jaschhof \＆U．Kallweit（NZAC， SDEI－Dipt－0000956）．Paratypes：6ふへへ ，same date and same locality as holotype（2x NZAC，SDEI－Dipt－0000952； 4 x SDEI，SDEI－Dipt－0000964）． 4 ठ $^{\AA}$ same locality as holotype，Malaise trap，04－24．01．2002，leg．M．\＆C．Jas－ chhof（ 2 x NZAC；2x SDEI）． $4 \widehat{\jmath}$ South Island，Buller， 5 km W Maruia Springs，mixed Nothofagus forest，Malaise trap，26．11－25．12．2001，leg．M．\＆C．Jaschhof（2x NZAC，SDEI－Dipt－0000988；2x SDEI，SDEI－Dipt－0000942 \＆ SDEI－Dipt－0000973）． $35{ }^{\top}{ }^{\lambda}$ ，South Island，Buller，Ahaura，Granville State Forest，altitude 170－250 m，Nothofagus truncata forest，Malaise trap，01．12．1994，leg．D．Hutcheson（17x NZAC，18x SDEI）． $7{ }^{\top}{ }^{\top}$ South Island，Buller， Maruia Forest，Shenandoah Saddle，altitude 500 m ，Nothofagus forest，Malaise trap，09．10－03．11．2001，leg．M．\＆ C．Jaschhof（3x NZAC，SDEI－Dipt－0000788，SDEI－Dipt－0000960 \＆SDEI－Dipt－0001179；4x SDEI，SDEI－Dipt－ 0000752，SDEI－Dipt－0000787，SDEI－Dipt－0001180 \＆SDEI－Dipt－0001181）．3 ふた same locality as previous，Ma－ laise trap，03－25．11．2001，leg．M．\＆C．Jaschhof（1x NZAC，SDEI－Dipt－0001045；2x SDEI，SDEI－Dipt－0001038 \＆SDEI－Dipt－0001044）． $1 \overbrace{}^{\lambda}$ same locality as previous，sweep net， 25.12 .2001 ，leg．M．Jaschhof（SDEI）． $2 \delta^{\lambda}{ }^{\lambda}$ South Island，Buller，Paparoa NP， 5 km E Punakaiki，Inland Pack Track，mixed podocarp／Nothofagus forest，Malaise trap， 30．09－05．11．2001，leg．M．\＆C．Jaschhof（NZAC；SDEI）．4 § South Island，Buller，Rahu Scenic Reserve， 12 km NW Springs Junction，altitude 550 m ，mixed Nothofagus／podocarp forest，Malaise trap，27．11－25．12．2001，leg．M． \＆C．Jaschhof（2x NZAC，SDEI－Dipt－0001481 \＆SDEI－Dipt－0001490；2x SDEI，SDEI－Dipt－0001492 \＆SDEI－ Dipt－0001493）． $1 \delta$ South Island，Westland，Waiatoto，Kahikatea swamp forest，Malaise trap，16．10－20．11．2001， M．\＆C．Jaschhof（SDEI）． $1 才$ South Island，Westland，Westland NP，SE Gillespies Beach，mixed podocarp forest， Malaise trap，14．10－21．11．2001，leg．M．\＆C．Jaschhof（SDEI，SDEI－Dipt－0001028）．3§§ South Island，Queens－ town Lakes，Otago Lakes，Mt Aspiring NP，Makarora River Valley near Cameron Creek Track，mixed Nothofagus／ broadleaf forest，sweep net，18－19．12．2002，leg．M．Jaschhof（1x NZAC，SDEI－Dipt－0000513；2x SDEI，SDEI－ Dipt－0001506）． $2{ }^{\top} \delta^{\lambda}$ South Island，Southland，Fiordland，Fiordland NP，Lake Gunn，Nothofagus forest，sweep net， 08.01 .2002 ，leg．M．Jaschhof（NZAC；SDEI）． $1 \circlearrowleft^{\lambda}$ same locality as previous，Malaise trap，01－21．12．2002，leg．M．\＆ C．Jaschhof \＆U．Kallweit（SDEI）． $2{ }^{\top}$ South Island，Southland，Fiordland，Fiordland NP， 3 km E Milford Sound， mixed podocarp／broadleaf forest，Malaise trap，01－21．12．2002，leg．M．\＆C．Jaschhof \＆U．Kallweit（NZAC； SDEI）． $1 \delta^{\lambda}$ same locality as previous，sweep net，21．12．2002，leg．M．Jaschhof（SDEI）． $8 \delta^{\Uparrow} \delta^{\pi}$ South Island，South－ land，Fiordland，Fiordland NP，Hollyford River Valley，Moraine Creek Track，mixed podocarp／Nothofagus forest， Malaise trap，05－24．01．2002，leg．M．\＆C．Jaschhof（4x NZAC，SDEI－Dipt－0000910，SDEI－Dipt－0000927，SDEI－ Dipt－0000928 \＆SDEI－Dipt－0000934；4x SDEI，SDEI－Dipt－0000879，SDEI－Dipt－0000881，SDEI－Dipt－0000907 \＆ SDEI－Dipt－0000924）． $7{ }^{\top} \circlearrowleft^{\Uparrow}$ South Island，Southland，Otago Lakes，Fiordland NP，Hollyford River Valley S Divide Creek，mixed Nothofagus／broadleaf forest，Malaise trap，06－24．01．2002，leg．M．\＆C．Jaschhof（3x NZAC；4x SDEI，SDEI－Dipt－0000994）． $1 \sigma^{\text {® }}$ South Island，Southland，Otago Lakes，Fiordland NP，Eglinton River Valley，op－ posite Earl Mountain Tracks，Nothofagus forest，Malaise trap，05－24．01．2002，leg．M．\＆C．Jaschhof（SDEI，SDEI－ Dipt－0000871）． $3 \circlearrowleft^{\top} \circlearrowleft^{\lambda}$ same locality as previous，Malaise trap，01－21．12．2002，M．\＆C．Jaschhof \＆U．Kallweit （1x NZAC；2x SDEI）． $7 \delta^{\top}$ South Island，Southland，Otago Lakes，Fiordland NP，Eglinton River Valley， 2 km N entrance Earl Mountain Tracks，Nothofagus forest，Malaise trap，01－21．12．2002，leg．M．\＆C．Jaschhof \＆U．Kall－ weit（3x NZAC，SDEI－Dipt－0001169 \＆SDEI－Dipt－0001173；4x SDEI，SDEI－Dipt－0001165，SDEI－Dipt－0001172 \＆SDEI－Dipt－0001174）． 2 § South Island，Southland，Otago Lakes，Fiordland NP，Eglinton River Valley， 2 km N entrance Gladehouse Track，Nothofagus forest，Malaise trap 01－20．12．2002，leg．M．\＆C．Jaschhof \＆U．Kallweit （NZAC，SDEI－Dipt－0001206；SDEI，SDEI－Dipt－0001143）．7 $\overbrace{}^{\lambda}$ South Island，Southland，Otago Lakes，Fiordland NP，Eglinton River Valley，opposite Gladehouse Track，Nothofagus forest，Malaise trap，01－21．12．2002，leg．M． \＆C．Jaschhof \＆U．Kallweit（4x NZAC，SDEI－Dipt－0001006，SDEI－Dipt－0001014 \＆SDEI－Dipt－0001016；3x SDEI，SDEI－Dipt－0001012，SDEI－Dipt－0001017 \＆SDEI－Dipt－0001025）． 5 § ${ }^{\wedge}$ same locality as previous，sweep net，03－04．12．2002，leg．M．Jaschhof（ 2 x NZAC；3x SDEI）． $1{ }^{\text {§ }}$ South Island，Clutha，Catlins，Purakaunui Scenic Reserve，mixed podocarp／broadleaf／Nothofagus forest，sweep net，03．01．2002，leg．M．Jaschhof（SDEI，SDEI－Dipt－ 0001484）． $3{ }^{\top}{ }^{\top}$ same locality as previous，Malaise trap，27．01－05．03．2002，leg．M．\＆C．Jaschhof（ 1 x NZAC，SDEI－ Dipt－0001139；2x SDEI，SDEI－Dipt－0001132 \＆SDEI－Dipt－0001156）．4才 ${ }^{\top}$ South Island，Clutha，Catlins Coastal Rain Forest Park，Catlins River Valley，silver beech forest，Malaise trap，27．01－05．03．2002，leg．M．\＆C．Jaschhof （2x NZAC，SDEI－Dipt－0000614 \＆SDEI－Dipt－0000801；2x SDEI，SDEI－Dipt－0000804 \＆SDEI－Dipt－0000814）．
Additional material：1 ${ }^{\lambda}$ ，SEM－object，same date and same locality as holotype（SDEI，SDEI－Dipt－0000968）．
Description．Male．Head．Head capsule brown．Eye bridge two to three facets wide．Scape and pedicel mostly
slightly paler brown than the flagellum, or as brown as flagellum; fourth flagellomere $2.5-3.3$ times longer than wide; necks of flagellomeres well differentiated; surface of flagellomeres rough with deep pits, sensilla of two different lengths, longer sensilla robust and curved; setae on the flagellomere slightly shorter than flagellomere width, slightly curved. Maxillary palp long and pale containing three palpomeres, first one longest, third shortest, first palpomere with four to five long bristles; sensilla on first palpomere long. Prefrons and clypeus bulging. Thorax. lateral side yellowish brown, mesonotum and metanotum brownish. Posterior pronotum bare. Anterior pronotum with two to four setae. Episternum 1 with four to seven setae. Mesonotum with five to seven robust lateral bristles. Scutellum with three to four robust bristles and several smaller setae. Katepisternum as long as high, distal part slightly darker than proximal part. Wing. Length $2.3-2.7 \mathrm{~mm}$; width/length ratio $0.35-0.41$. Membrane transparent and without macrotrichia, anal area present; all posterior veins distinct, stem of M weak; apical half to apical third of $\mathrm{R}_{5}$ with macrotrichia on both sides; exceptionally in one specimen (SDEI-Dipt-0001156) three macrotrichia on $\mathrm{M}_{1}$ on left wing and one macrotricha on the right $\mathrm{M}_{1} ; \mathrm{r}-\mathrm{m}$ and bM without macrotrichia (exceptionally one macrotrichium on r-m on one wing of one specimen [SDEI-Dipt-0001490]); $\mathrm{R}_{1}$ approximately $0.9-1.3$ times as long as R ; $\mathrm{c} / \mathrm{w}$ ratio $0.6-0.8$; r-m 1.1-1.3 as long as bM. Haltere brownish; head of haltere as long as shaft. Legs pale brown, much brighter than thorax. Tibial organ surrounded by a circular border and as wide as half of the width of the tibial apex; tibial organ consists of approximately ten bristles arranged in an irregular row. Front tibia without or with one robust bristle among the vestiture. Mid tibia with one or two slightly robust bristles. Hind tibia with a posteriodorsal row of robust bristles, comprising $50-66 \%$ of apical part of tibia, several longer additional bristles on the ventral side. All tibial spurs equal in length. Claws with five robust teeth. Abdomen pale brown, slightly paler than thorax; with long, dark setae. Hypopygium (Figs 20-24 \& 43). Gonocoxites wider than long, inner side of gonocoxites with short setae, outer side with long setae; ventral, inner side of gonocoxites widely separated basally. Gonostylus 2.2-2.7 times longer than wide; basal part broad, dorsal margin slightly stretched; apical part large, curved inwards with an apical cavity without microtrichia (Fig. 20); subapically one spine and one long whip-lash hair. Tegmen (Fig. 23) with typical structure of the Psl. jejuna complex, parameral apodemes strongly sclerotized up to the apex of the tegmen. Area of teeth absent. Ejaculatory apodeme short and dark, with a distinct base; base pale bown and slender. Posterior portion of gonocoxal apodeme medium sized, brown, and medially connected. Anterior portion of gonocoxal apodeme short and dark brown.

Body size: 2.7-3.5 mm.
Female. Unknown.
Genetic distances. The maximum p-distance between all 48 available COI sequences is $2.28 \%$. The nearest neighbour is Psl. whakahara, diverging by a minimum of $3.51 \%$. All six available 28 S sequences are identical and do not show differences to Psl. whakahara.

Etymology. Te Waipounamu is the Māori name for the South Island of New Zealand. The epithet of the new species refers to the restricted distribution of this species.

Distribution. New Zealand.
Discussion. Pseudolycoriella tewaipounamu is one of the larger species of the Psl. jejuna complex. Some specimens reach a body size similar to that of Psl. whakahara, but never have a similarly large wing length. Furthermore, Psl. tewaipounamu shows a well-developed apical cavity on the gonostylus, without any microtrichia. The tegmen of this species has strongly sclerotized parameral apodemes, which are darkly coloured up to the apex of the tegmen. In nearly all other species only the proximal branches of these parameral apodemes are strongly sclerotized. Only Psl. whakahara has dark coloured distal branches like Psl. tewaipounamu, but they are always darker in Psl. tewaipounamu.

## Pseudolycoriella whakahara sp. n.

(Fig. 45)
urn:lsid:zoobank.org:act:NomenclaturalActs/2227602E-6F96-471F-BED4-CAEB57B55893

Material studied. Holotype male. New Zealand: South Island, Tasman, Kahurangi NP, Takaka River Valley (Cobb Dam Road), altitude 450-800 m, mixed podocarp/broadleaf forest, Malaise trap, 31.08-07.10.2001, leg. M. \& C. Jaschhof (NZAC, SDEI-Dipt-0001439). Paratypes. 2 | đ |
| :---: |
| same locality and same date as holotype (all SDEI, SDEI- | Dipt-0001386 and a second specimen in poor condition).

Description. Male. Head. Head capsule brown. Eye bridge two to three facets wide. Scape and pedicel pale
brown, slightly paler than the flagellum. Fourth flagellomere 3.5 times longer than wide; necks of flagellomeres well differentiated; surface of flagellomeres rough, with deep pits, sensilla of two different lengths, setae on the flagellomere robust, longer than flagellomere width, slightly curved. Maxillary palp long and pale, consisting of three palpomeres, first one longest, second shortest, first palpomere with four bristles, one of them more robust; sensilla on first palpomere present. Prefrons and clypeus bulging. Thorax pale brown. Posterior pronotum bare. Anterior pronotum with three to four setae. Episternum 1 with five to nine setae. Mesonotum with five to six robust lateral bristles. Scutellum with four to six robust bristles and several smaller setae. Katepisternum as long as high. Wing. Length 3.1-3.2 mm; width/length ratio $0.32-0.39$. Membrane slightly shaded and without macrotrichia, anal area present; all posterior veins distinct, stem of M weak; apical $50 \%$ to $60 \%$ of $\mathrm{R}_{5}$ with macrotrichia on both sides; $\mathrm{R}_{1}$ $0.9-1.0$ times as long as R ; c/w ratio $0.75-0.77$; r-m $0.6-0.7$ as long as bM . Haltere brownish; head of haltere shorter than shaft. Legs pale brown, brighter than thorax. Tibial organ surrounded by a circular border and as wide as half width of tibial apex; bristles of tibial organ arranged in an irregular row. Front tibia without robust bristle among the vestiture; mid tibia with two robust bristles among the vestiture; hind tibia with an inconspicuous posteriodorsal row of longer setae, two to three robust bristles in the middle of this row. All tibial spurs equal in length. Claws with robust teeth. Abdomen slightly paler than thorax; with long, dark setae. Hypopygium (Fig. 45). Gonocoxites wider than long, inner side of gonocoxites with medium sized setae, outer side with long setae; ventral, inner side of gonocoxites widely separated basally. Gonostylus $2.2-2.5$ times longer than wide; broad, slightly tumid in the middle, apical part not strikingly set off; apical cavity conspicuous, whole inner side without microtrichia; subapically one spine and one long whip-lash hair. Tegmen of the typical ground plan of the Psl. jejuna complex; the dorsal folds strikingly developed; parameral apodemes strongly sclerotized basally, moderately sclerotized apically. Area of teeth absent. Ejaculatory apodeme short and strongly sclerotized, base of ejaculatory apodeme delicate, broad, longer than remaining ejaculatory apodeme. Posterior portion of gonocoxal apodeme of medium size and brown, joined medially. Anterior portion of gonocoxal apodeme of medium length and dark brown.

Body size: $3.4-3.8 \mathrm{~mm}$.
Female. Unknown.
Genetic distances. Only one specimen was successfully sequenced. The nearest neighbour is Psl. tewaipouna$m u$, diverging by a minimum of $3.51 \%$. For the 28 S sequence the nearest neighbours are Psl. sudhausi and Psl. tewaipounaти.

Etymology. Whakahara is the Māori modifier for large or giant. It was chosen as the epithet to reflect the large size of the new species.

Distribution. New Zealand.
Discussion. With a body size of more than 3.4 mm , Psl. whakahara is the largest species of the Psl.jejuna complex and thus nearly unmistakable. There is a small overlap with larger specimens of Psl. tewaipounamu. Nevertheless, such specimens can be separated by comparing wing lengths. The wing of Psl. tewaipounamu never reaches in the length of the wing of Psl. whakahara ( $2.3-2.7 \mathrm{~mm} v s .3 .1-3.2 \mathrm{~mm}$, respectively). Some specimens of Psl.dagae also has a wing length which reaches the lower span of Psl. whakahara, but the former species can be separated by the more slender and concave gonostylus.

## Pseudolycoriella sudhausi sp. n.

(Fig. 37
urn:lsid:zoobank.org:act:NomenclaturalActs/427BBF3C-2DDD-44CA-B7BF-76C0B15212BA)

Material studied. Holotype male. New Zealand: Stewart Island, Southland, Kaipipi Bay 3 km W Halfmoon Bay, mixed broadleaf/podocarp forest, sweep net, 07.03.2002, leg. M. Jaschhof (NZAC, SDEI-Dipt-0001090). Paratypes. $1 \delta^{\lambda}$ same locality and same date as holotype (SDEI, SDEI-Dipt-0001093). $2 \delta^{\lambda} \delta^{\lambda}$ South Island, Westland, Waiatoto; Kahikatea swamp forest, Malaise trap, 16.10-20.11.2001, leg. M. \& C. Jaschhof (all SDEI). 2 § $^{\top}$ © South Island, Westland, Westland NP, Lake Matheson, mixed podocarp forest, exhauster, 22.10.2001, leg. M. Jaschhof (NZAC; SDEI). $2 \widehat{§}^{\lambda}$ South Island, Westland, Westland NP, SE Gillespies Beach, mixed podocarp forest, exhauster, 19.10.2001, leg. M. Jaschhof (NZAC; SDEI). 3 §§ South Island, Southland, Fiordland, Fiordland NP, Hollyford River Valley, Moraine Creek Track, mixed podocarp/Nothofagus forest, Malaise trap, 05-24.01.2002, leg. M. \& C. Jaschhof (2x NZAC, SDEI-Dipt-0000885 \& SDEI-Dipt-0000922; 1x SDEI, SDEI-Dipt-0000929). 6ðð South Island, Southland, Otago Lakes, Fiordland NP, Eglinton River Valley, 2 km N entrance Earl Mountain Tracks,

Nothofagus forest, Malaise trap, 01-21.12.2002, leg. M. \& C. Jaschhof \& U. Kallweit (3x NZAC, SDEI-Dipt0001167 \& SDEI-Dipt-0001175; 3x SDEI, SDEI-Dipt-0001170 \& SDEI-Dipt-0001459). $6{ }^{\AA}$ § South Island, Southland, Otago Lakes, Fiordland NP, Eglinton River Valley, 2 km N entrance Gladehouse Track, Nothofagus forest, Malaise trap, 01-20.12.2002, leg. M. \& C. Jaschhof \& U. Kallweit (2x NZAC, SDEI-Dipt-0001147 \& SDEI-Dipt0001148; 4x SDEI, SDEI-Dipt-0001146, SDEI-Dipt-0001149 \& SDEI-Dipt-0001177). $2{ }^{\Uparrow}{ }^{\Uparrow}$ South Island, Southland, Otago Lakes, Fiordland NP, Eglinton River Valley, opposite Gladehouse Track, Nothofagus forest, sweep net, 03-04.12.2002, leg. M. Jaschhof (NZAC; SDEI). 4ठ $\begin{gathered}\text { § same locality as previous, Malaise trap, 01-21.12.2002, leg. }\end{gathered}$ M. \& C. Jaschhof \& U. Kallweit (3x NZAC, SDEI-Dipt-0001009, SDEI-Dipt-0001013 \& SDEI-Dipt-0001015; 1x SDEI, SDEI-Dipt-0001011). $1 \circlearrowleft^{\lambda}$ South Island, Southland, Otago Lakes, Fiordland NP, Eglinton River Valley, Deer Flat, Nothofagus forest, Malaise trap, 04-24.01.2002, leg. M. \& J. Jaschhof (SDEI). 10 same locality as previous, Malaise trap, 01-20.12.2002, leg. M. \& C. Jaschhof \& U. Kallweit (NZAC). $1 \overbrace{}^{\lambda}$ Stewart Island, Southland, Rakiura Track between Kaipipi Bay and Sawdust Bay, mixed podocarp/broadleaf forest, sweep net, 07.03.2002, leg. M. Jaschhof (NZAC). $6 \delta^{\lambda}$ Stewart Island, Southland, Rakiura Track between Sawdust Bay and North Arm Hut, mixed podocarp/broadleaf forest, sweep net, 08.03.2002, leg. M. Jaschhof (2x NZAC; 4x SDEI, SDEI-Dipt-0001071 \& SDEI-Dipt-0001116). 5 ${ }^{\text {§ }}$ Stewart Island, Southland, Freshwater Landing Hut, mixed podocarp/manuka/broadleaf forest, sweep net, 12.03.2002, leg. M. Jaschhof (3x NZAC; 2x SDEI). Additional material. $1 \delta^{\lambda}$ South Island, Southland, Otago Lakes, Fiordland NP, Eglinton River Valley, opposite Earl Mountain Tracks, Nothofagus forest, Malaise trap, 01.12-21.12.2002, leg. M. \& C. Jaschhof \& U. Kallweit (SDEI; specimen in poor condition).

Description. Male. Head. Head capsule brown. Eye bridge two to three facets wide. Scape and pedicel slightly paler brown than the flagellum; fourth flagellomere 2.9-3.5 times longer than wide; necks of flagellomeres well differentiated, length of necks increase towards the antennal apex; surface of flagellomeres rough with deep pits, sensilla of two different lengths, small ones and longer, curved sensilla; setae on the flagellomere longer than the flagellomere width, slightly curved. Maxillary palp long and three-segmented, first palpomere is the longest one, second one shortest; first palpomere with long sensilla and two to four long bristles, one bristle longer and more robust, located on the outer side of the first palpomere. Prefrons and clypeus bulging. Thorax pale brown, mesonotum slightly darker. Posterior pronotum bare. Anterior pronotum with four to five setae. Episternum 1 with three to seven setae. Mesonotum with four to five robust lateral bristles. Scutellum with three to four robust bristles and several minor setae. Dorsocentral bristles forming a complete row on the mesonotum, anterior ones smaller. Katepisternum as long as high. Wing. Length $2.2-2.8 \mathrm{~mm}$; width/length ratio $0.37-0.41$. Membrane transparent and without macrotrichia, anal area present; all posterior veins distinct, except faint stem of M, apical 33-50\% of $\mathrm{R}_{5}$ with macrotrichia on ventral and dorsal side; bM and r-m bare (except one paratype [SDEI-Dipt-0000922] with one macrotricha on $\mathrm{r}-\mathrm{m}$ on one wing); $\mathrm{R}_{1} 0.6-0.9$ times as long as R ; $\mathrm{c} / \mathrm{w}$ ratio $0.76-0.81$; $\mathrm{r}-\mathrm{m} 0.9-1.3$ as long as bM. Haltere brownish; shaft of haltere longer than head. Legs pale brown, slightly paler than thorax, occasionally mid and hind coxae concolourous with thorax. Tibial organ surrounded by a circular border and slightly wider than half of the width of the tibial apex; tibial organ consists of an irregular row of bristles. Front tibia without robust bristles among the vestiture (occasionally one bristle). Mid tibia with one slightly robust bristle among vestiture, or bristle missing. Posteriodorsal row of bristles inconspicuous, one to four weak bristles. All tibial spurs equal in length. Claws with robust teeth. Abdomen concolourous with thorax, with long, dark setae. Hypopygium (Fig. 37). Gonocoxites wider than long, inner side of gonocoxites with medium sized setae, outer side with long setae; ventral, inner side of gonocoxites basally separated. Gonostylus elongate, 2.6-3.1 times longer than wide, apex globular and slightly curved inwards, inner side slightly tumid, apical cavity small without microtrichia, remaining inner side scattered with microtrichia, one spine and one whip-lash hair present. Tegmen of the typical ground plan of the Psl. jejuna complex, onion-shaped but elongated; dorsal folds present; parameral apodemes basally strongly sclerotized, medially connected, distal branches close and nearly parallel. Area of teeth absent. Ejaculatory apodeme pale, narrow and long, proximal end slightly thickened and slightly darkened; base of ejaculatory apodeme delicate, narrow and long. Posterior portion of gonocoxal apodeme medium sized and brown, joined medially. Anterior portion of gonocoxal apodeme long and dark brown, nearly reaching the proximal end of the hypopygium.

Body size: 2.3-2.8 mm.
Female. Unknown.
Genetic distances. The maximum p-distance between all 19 available COI sequences is $1.37 \%$. The nearest neighbour is Psl. whena, diverging by a minimum of $5.02 \%$. All five available 28 S sequences are identical and do not show differences to sequences of Psl. porotaka and Psl. whena. One of those sequences is also identical to the
sequence of Psl. whakahara. The reason for this is that both sequences are fragmentary and only share a middle part of about 480 bp in the alignment.

Etymology. The new species is named after the phylogenetic biologist Prof. Dr Walter Sudhaus.
Distribution. New Zealand.
Discussion. Pseudolycoriella sudhausi and the species Psl. nahenahe, Psl. whena, and Psl. porotaka have in common a much narrower ejaculatory apodeme than the remaining species of the Psl. jejuna complex. Of these species, Psl. sudhausi and Psl. whena have a width/length ratio of the fourth flagellomere greater than 2.8. Pseudolycoriella sudhausi can be distinguished from Psl. whena by the wing length ( $2.2-2.8 \mathrm{~mm} v s .1 .9-2.1 \mathrm{~mm}$, respectively). The body size also differs, but shows a small range of overlap at 2.3 mm . Pseudolycoriella whena has a comparatively smaller tegmen, a shorter gonostylus, and a shorter anterior portion of gonocoxal apodeme than Psl. sudhausi.

## Pseudolycoriella whena sp. n.

(Fig. 38)
urn:Isid:zoobank.org:act:73E8E6FC-CF31-4DD9-9751-C8653FAFE299
Material studied. Holotype male. New Zealand: North Island, Western Bay of Plenty, Katikati, Te Mania catchment, Malaise trap, 19.09-27.10.2016, leg. P.A. Maddison (NZAC, SDEI-Dipt-0001336). Paratypes. $6{ }^{\wedge} ð^{\wedge}$ South Island, Marlborough, Marlborough Sounds, Okiwi Bay, Moncrieff Scenic Reserve, mixed Nothofagus/podocarp forest, sweep net, 24.12.2001, leg. M. Jaschhof (3x NZAC, SDEI-Dipt-0001118; 3x SDEI, SDEI-Dipt-0001100 \& SDEI-Dipt-0001127).

Description. Male. Head. Head capsule brown. Eye bridge two to three, exceptionally four, facets wide, two at margin. Scape and pedicel mostly paler, contrasting with the brown flagellum. Fourth flagellomere 2.8-3.2 times longer than wide; necks of flagellomeres well differentiated; surface of flagellomeres rough with deep pits, sensilla of two different lengths, setae on the flagellomere robust, longer than flagellomere width, slightly curved. Maxillary palp of medium length and pale, consisting of three palpomeres, first one longest, second shortest, first palpomere with two to three bristles, one of them longer and more robust, located on the outer side; sensilla on first palpomere long. Prefrons and clypeus bulging. Thorax pale brown, mesonotum slightly darker. Posterior pronotum bare. Anterior pronotum with two to four setae. Episternum 1 with four to seven setae. Mesonotum with two to four robust lateral bristles. Scutellum with four robust bristles and several smaller setae. Dorsocentral bristles forming a complete row on the mesonotum, anterior ones smaller. Katepisternum as long as high. Wings of the holotype damaged. Length 1.9-2.1 mm; width/length ratio $0.37-0.41$. Membrane transparent and without macrotrichia, anal area present; all posterior veins distinct, except faint stem of $M$, apical $33-40 \%$ of $R_{5}$ with macrotrichia on ventral and dorsal side (except two paratypes which each bear only one macrotrichium on the ventral side of $\mathrm{R}_{5}$ ); bM and $\mathrm{r}-\mathrm{m}$ without macrotrichia; $\mathrm{R}_{1} 0.6-0.7$ times as long as R; c/w ratio $0.72-0.78 ; \mathrm{r}-\mathrm{m} 1.0-1.5$ as long as bM . Haltere brownish; head of haltere shorter than shaft. Legs pale brown, slightly brighter than thorax. Tibial organ surrounded by a circular border and as wide as or even smaller than half the width of the tibial apex; tibial organ consists of an irregular row of bristles. Fore tibia without robust bristles among the vestiture (two paratypes with only one bristle on one side). Mid tibia with none to two slightly robust bristles among the vestiture. Posteriodorsal row of bristles inconspicuous, four to nine weak bristles. All tibial spurs equal in length. Claws with robust teeth. Abdomen pale brown, as thorax; with long, dark setae. Hypopygium (Fig. 38). Gonocoxites wider than long, inner side of gonocoxites with short setae, outer side with long setae; ventral, inner side of gonocoxites widely separated basally. Gonostylus 2.8-3.1 times longer than wide; elongate and curved, apical part slightly curved inwards, inner side slightly tumid, apical cavity small with scattered macrotrichia; subapically one spine and one long whip-lash hair present. Tegmen of the typical ground plan of the Psl. jejuna complex, with an acuminate basic shape; the dorsal folds strikingly developed; parameral apodemes basally strongly sclerotized, medially fused. Area of teeth absent, exceptionally two paratypes (SDEI-Dipt-0001100 \& SDEI-Dipt-0001127) with small aggregations of five and twelve tiny teeth, respectively. Ejaculatory apodeme long and of medium thickness, base of ejaculatory apodeme very delicate, short, and u-shaped. Posterior portion of gonocoxal apodeme medium sized and brown, joined medially. Anterior portion of gonocoxal apodeme long and dark brown.

Body size: 1.8-2.3 mm.
Female. Unknown.
Genetic distances. DNA was successfully isolated from only one specimen. The nearest neighbour is Psl. sudhausi, diverging by a minimum of $5.02 \%$. The available 28 S sequence is identical to those of Psl. porotaka and Psl. sudhausi.

Etymology. The Māori word for dwarf—whena-was chosen as epithet, to highlight the minute body size of the new species.

Distribution. New Zealand.
Discussion. With a body size of up to 2.3 mm , Psl. whena together with Psl. jejunella, Psl. nahenahe, and Psl. sudhausi belongs to the smaller species of this species complex. Pseudolycoriella whena can be distinguished from Psl. jejunella and Psl. nahenahe by the longer fourth flagellomere ( $>2.8 v s .<2.5$ ). Pseudolycoriella jejunella also differs significantly in the shape of the gonostylus. For the distinction of Psl. whena and Psl. sudhausi see discussion paragraph for Psl. sudhausi.

In two paratypes of Psl. whena atavistic remains of the area of the teeth are present (see Psl. jejuna complex discussion).

## Pseudolycoriella porotaka sp. n.

(Fig. 39)
urn:lsid:zoobank.org:act:NomenclaturalActs/EC053068-0F35-494E-9F41-4A28247CA143
Material studied. Holotype male. New Zealand: North Island, Bay of Plenty, Katikati, Te Mania catchment, Malaise trap, 19.09-27.10.2016, leg. P.A. Maddison (NZAC, SDEI-Dipt-0001225). Paratype. $1 \delta^{\lambda}$ North Island, Bay of Plenty, Katikati, 449 Lund Road, Malaise trap, 25.07-08.08.2016, leg. P.A. Maddison (SDEI, SDEI-Dipt-0001250).

Description. Male. Head. Head capsule brown. Eye bridge three facets wide. Scape and pedicel much paler than the brown flagellomeres; fourth flagellomere 2.3-2.4 times longer than wide; necks of flagellomeres well differentiated, paler than body of flagellomeres; surface of flagellomeres rough with deep pits, sensilla of two different lengths, small ones and longer, curved sensilla; densely arranged setae on the flagellomere shorter than flagellomere width, slightly curved, arising from small elevations. Maxillary palp very long and three-segmented, first palpomere longest, second one shortest; first palpomere with an inconspicuous patch of sensilla and four long bristles, one of which is longer and more robust, located on the outer side of the first palpomere. Prefrons and clypeus bulging. Thorax brown, laterally brighter. Posterior pronotum bare. Anterior pronotum with three to five setae. Episternum 1 with seven to nine setae. Mesonotum with five robust lateral bristles. Scutellum with four robust bristles and several minor setae. Katepisternum slightly longer than high. Wing. Length $2.7-2.8 \mathrm{~mm}$; width/length ratio 0.36-0.37. Membrane transparent and without macrotrichia, anal area present; all posterior veins distinct, except faint stem of M , apical half to three-fifths of $\mathrm{R}_{5}$ with macrotrichia on both ventral and dorsal sides; bM and r-m bare; $\mathrm{R}_{1} 0.8-0.9$ times as long as R ; $\mathrm{c} / \mathrm{w}$ ratio $0.77-0.78$; $\mathrm{r}-\mathrm{m}$ as long as bM . Haltere brownish; head of haltere slightly longer than shaft. Legs yellowish pale brown, paler than thorax; coxae pale brown. Tibial organ surrounded by a circular border and as wide as half of the width of the tibial apex; tibial organ consists of an irregular row of bristles. Front tibia without robust bristles among the vestiture. Mid tibia without (holotype) or with two (paratype) robust bristles among the vestiture. Posteriodorsal row of bristles inconspicuous, extending along two-thirds of tibia apex. All tibial spurs equal in length. Claws with robust teeth. Abdomen concolourous with long, dark setae. Hypopygium (Fig. 39). Gonocoxites wider than long, inner side of gonocoxites with small setae, outer side with long setae; ventral, inner side of gonocoxites not fused basally. Gonostylus elongate, slightly tumid in the middle, with a dorsal edge, 2.7-2.9 times longer than wide; apex moderately separated, slightly bent, apical end broad, blunt, on ventral side with a apical cavity without microtrichia, one spine and one whip-lash hair present. Tegmen of the typical ground plan of the Psl.jejuna complex, of very rounded shape; dorsal folds present; parameral apodemes basally strongly sclerotized, medially fused, distal branches very parallel and close. Area of teeth absent. Ejaculatory apodeme pale brown, slender, and long; base very delicate, long, and u-shaped. Posterior portion of gonocoxal apodeme medium sized and brown, joined medially. Anterior portion of gonocoxal apodeme short and dark brown.

Body size: 2.9-3.1 mm.
Female. Unknown.
Genetic distances. Both available COI sequences show a p-distance of $1.16 \%$. The nearest neighbour is Psl. whena, diverging by a minimum of $9.30 \%$. Both available 28 S sequences are identical and do not show differences to the sequences of Psl. sudhausi and Psl. whena.

Etymology. Porotaka is a Māori word which means round or circular. The epithet refers to the roundish shape of the tegmen.

Distribution. New Zealand.


FIGURES 43-47. Pseudolycoriella jejuna complex, hypopygia. 43. Pseudolycoriella tewaipounamu sp. n. 44. Pseudolycoriella hauta sp. n. 45. Pseudolycoriella whakahara sp. n. 46. Pseudolycoriella jejunella sp. n. 47. Pseudolycoriella maddisoni sp. n.

Discussion. Pseudolycoriella porotaka can be distinguished from all other species of the Psl.jejuna complex by the combination of a very rounded tegmen and a low width/length ratio of the fourth flagellomere. In addition, this species has flagellomeres with shorter and denser setae compared to the other species. Like Psl. nahenahe, Psl. sudhausi, and Psl. whena, Psl. porotaka has a narrow ejaculatory apodeme.

## Pseudolycoriella nahenahe sp. n.

(Fig. 40)
urn:lsid:zoobank.org:act:B0518863-0580-4308-B7E2-61F10B824DB8
Material studied. Holotype male. New Zealand: Coromandel, Wharekawa Redwood Picnic Area near Opoutere, redwood plantation with pines and tree fern, sweep net, 09.02.2002, leg. M. Jaschhof (NZAC).

Description. Male. Head. Head capsule brown. Eye bridge four facets wide, two facets at margin. Scape and pedicel pale, contrasting with the brown flagellum; fourth flagellomere 2.5 times longer than wide; necks of flagellomeres well differentiated; surface of flagellomeres rough with deep pits, sensilla of two different lengths, small ones and longer, curved sensilla; setae as long as a flagellomere's width, slightly curved. Maxillary palp long and three-segmented, first palpomere is the longest, the second shortest; first palpomere with long sensilla and three long bristles. Prefrons and clypeus bulging. Thorax brown, with frontal and lateral pale brown patches. Posterior pronotum bare. Anterior pronotum with three setae. Episternum 1 with six setae. Mesonotum with five robust lateral bristles. Scutellum with four robust bristles and several minor setae. Longer and shorter dorsocentral bristles forming a complete row on the mesonotum. Katepisternum as long as high. Wing. Length 2.2 mm ; width/length ratio 0.37. Membrane transparent and without macrotrichia, anal area present; all posterior veins distinct, except faint stem of M , apical two-thirds of $\mathrm{R}_{5}$ with macrotrichia on ventral and dorsal side; bM and r-m bare; $\mathrm{R}_{1} 0.7$ times as long as R; c/w ratio 0.71 ; r-m 1.2 as long as bM. Haltere brownish; shaft of haltere slightly longer than knob. Legs pale brown, slightly paler than thorax, mid and hind coxae concolourous with thorax. Tibial organ surrounded by a circular border and slightly wider than half of the width of the tibial apex; tibial organ consists of an irregular row of bristles. Front tibia without robust bristles among the vestiture. Mid tibia with one robust bristle among vestiture. Posteriodorsal row of bristles inconspicuous, consisting of three to five bristles. All tibial spurs equal in length. Claws with robust teeth. Abdomen concolourous with thorax, with long, dark setae. Hypopygium. (Fig. 40) Gonocoxites wider than long, inner side of gonocoxites with medium sized setae, outer side with long setae; ventral, inner side of gonocoxites basally separated. Gonostylus elongated, 2.6 times longer than wide, apex globular and slightly curved inwards, inner side tumid, apical cavity small; one spine and one whip-lash hair present. Tegmen of the typical ground plan of the Psl. jejuna complex, onion-shaped but elongated; parameral apodeme basally strongly sclerotized, medially connected. Area of teeth absent. Ejaculatory apodeme pale, narrow and short; base of ejaculatory apodeme not present. Posterior portion of gonocoxal apodeme long and brown, medially joined by a slender bridge. Anterior portion of gonocoxal apodeme long and dark brown.

Body size: 2.2 mm .
Female. Unknown.
Genetic distances. No genetic data available.
Etymology. The Māori adjective nahenahe-meaning being separate, alone etc.-was chosen as epithet, indicating that only one specimen of this newly described species was available.

Distribution. New Zealand.
Discussion. Due to the slender ejaculatory apodeme and the tumid gonostylus Psl. nahenahe is similar to Psl. sudhausi, Psl. porotaka, and Psl. whena. Among these species Psl. nahenahe can be distinguished by a combination of a small body size ( 2.2 mm ) and a low length-width index of the fourth flagellomere. Both Pseudolycoriella sudhausi and Psl. whena have a fourth flagellomere with a length-width index greater than 2.8 , while Psl. porotaka has a similar low index of the fourth flagellomere, but has a body size of more than 2.8 mm .

## Pseudolycoriella macrotegmenta complex

Thirteen of the species examined during this study have a very similar hypopygia structure. The number of gonostylar spines, ranging from four to ten, is greater than in most other Pseudolycoriella species. The length of those spines is also reduced. Consequently, the spines are very difficult to distinguish from the surrounding setae at the
apex of the gonostylus. The apical whip-lash hair is also mostly reduced in length, and in several species two or even three hairs are present. The tegmen possesses a variety of unique characters. Some species bear teeth on the lateral margin of the tegmen ( $l t$ in Fig. 52). In the most derived state these teeth cover the whole apicolateral part of the tegmen (Fig. 60). On the dorsal side of the tegmen of some species there are creases arising from the outer base and converging towards the apex. These dorsal structures ( $d s$ in Fig. 52) can vary in length and number. The phylogeny of the Psl. macrotegmenta complex is shown in Figure 61.

Discussion. An assignment of this assemblage of species to one of the existing species groups of Pseudolycoriella was not possible, as already mentioned by Köhler \& Mohrig (2016) for Psl. frederickedwardsi and Psl. tonnoiri. With the bulging prefrons and clypeus, and the ground plan of the tibial organ this species complex is undoubtedly part of the genus Pseudolycoriella. The increase in the number of gonostylar spines and the reduction of the length and the multiplication of the whip-lash hair can be considered to be morphological evidence for the monophyly of the Psl. macrotegmenta complex. The result of the molecular analysis also strongly supports the monophyly (Fig. 61). With the numerous gonostylar spines and the reduction of the length of the whip-lash hair the Psl. macrotegmenta complex resembles the Psl. quadrispinosa group from Papua New Guinea (Mohrig 2013). However, in the Psl. quadrispinosa group the gonostylar spines are easily distinguishable from the surrounding setae and the whiplash hair is lost, except in Psl. bitorquia Mohrig where a small whip-lash hair persists. In the Psl. macrotegmenta complex the spines are not as easy to distinguish from the surrounding setae and the whip-lash hair is never completely reduced, only shortened. Further, several species of the Psl. macrotegmenta complex possess two or three whip-lash hairs. Thus, the gonostylar spines and the whip-lash hair(s) do not necessarily share the same taxonomic value and it is likely that their appearance is a result of convergence. Another similar species is Psl. curvimedia Mohrig \& Rulik-a species recorded from the Dominican Republic. It also has a similar pattern of spines on the gonostylus. However, there are other characters that differ significantly from those in the Psl. macrotegmenta complex and therefore do not support integration into the Psl. macrotegmenta complex. The flagellomeres of Psl. curvimedia have a rather smooth surface (Mohrig et al. 2004), while those of the species of Psl. macrotegmenta complex are rough with deep pits. Also, the wing venation of Psl. curvimedia is very aberrant in comparison to representatives of the Psl. macrotegmenta complex. Pseudolycoriella curvimedia shows a very short $\mathrm{R}_{1}$ that only comprises a fourth of R, a very unusual M-fork, and a nearly rectangularly bent $\mathrm{CuA}_{2}$. Furthermore, the gonocoxites of Psl. curvimedia are fused ventrally, while they are widely separated in the Psl. macrotegmenta complex. Another similar species is the Australian Psl. rubroalata Mohrig, Kauschke \& Broadley, which also bears eight to ten short spines (Mohrig et al. 2018), but shows none of the characters on the tegmen that occur in the New Zealand Psl. macrotegmenta complex. However, without an accurate examination, preferably incorporating genetic analyses, a well-founded conclusion about the relationship of the Psl. quadrispinosa group, Psl. curvimedia, and Psl. rubroalata to the species of the Psl. macrotegmenta complex is not possible. Until such an examination, it remains unclear whether these species belong to the Psl. macrotegmenta complex or whether the similar patterns of gonostylar spines are a result of convergence. Nevertheless, it can be stated that surely these species do not belong to the New Zealand crown group of the Psl. macrotegmenta complex. The combination of an increased number of reduced (i.e. thinner) spines and flagellomeres with a pitted surface can be held to be a synapomorphy for the New Zealand members of this complex. Apart from the morphological considerations noted above, the very small genetic distances between the New Zealand representatives of this complex indicate a more recent radiation of these species.

## Pseudolycoriella macrotegmenta Mohrig, 1999

(Fig. 48)
Literature. Pseudolycoriella macrotegmenta Mohrig, 1999: Mohrig \& Jaschhof (1999): 41-43, Fig. 22 a-f.-Macfarlane et al. (2010): 441.

Material studied. Holotype male. New Zealand: North Island, Stratford, Mount Egmont National Park, East Egmont, altitude 650 m , Podocarpus wood with rotten wood and a dense layer of herbs, sweep net, 09.01.1993, leg. M. Jaschhof (PWMP). New records. $1 \jmath^{\Uparrow}$ North Island, Ruapehu, Tongariro NP, Mangawhero River Valley 3 km NE Ohakune, altitude 690 m , mixed podocarp/broadleaf forest, sweep net, 03-04.02.2002, leg. M. Jaschhof (SDEI, SDEI-Dipt-0000859). $2 \widehat{\jmath}$ same locality as previous, Malaise trap, 26.11-28.12.2002, leg. M. \& C. Jaschhof \& U. Kallweit (NZAC; SDEI, SDEI-Dipt-0000757).

Redescription. Male. Head brown; eye bridge three facets wide; lateral border of eye bridge two facets wide. Scape and pedicel brown, concolourous with flagellomeres; fourth flagellomere 2.9-3.6 times as long as wide; necks of flagellomeres differentiated; surface of flagellomeres rough with deep pits; sensilla of two different lengths present; setae fine and curved, approximately as long as flagellomere width. Prefrons and clypeus bulging. Maxillary palp three-segmented; first palpomere longest, second palpomere shortest; first palpomere with four bristles and a patch of sensilla. Thorax brown, laterally a brighter yellowish-brown. Posterior pronotum bare. Anterior pronotum with five to six bristles. Episternum 1 with seven to eleven bristles. Mesonotum with five to six robust lateral bristles and several longer bristles among the dorsocentrals. Frontal part of mesonotum with a few arcostichal bristles. Scutellum with four robust and some shorter bristles. Katepisternum as long as high. Wing. Length 2.4-2.5 mm; width/length ratio $0.36-0.38$. Membrane transparent and without macrotrichia, anal area present; all posterior veins distinct and without macrotrichia; apical quarter to two-thirds of $\mathrm{R}_{5}$ with additional macrotrichia ventrally; bM and $\mathrm{r}-\mathrm{m}$ bare, exceptionally one specimen with one seta on r-m on one wing; $\mathrm{R}_{1}$ short, $0.50-0.67$ times as long as R ; M -fork longer than stem of M ; $\mathrm{c} / \mathrm{w}$ ratio $0.70-0.74$; r-m/bM ratio $0.8-1.1$. Haltere pale brown. Legs pale brown; trochanter darker. Fore tibia with none or one to two bristles among the vestiture. Tibial organ comprising more than half of tibial apex, consisting of a patch of bristles surrounded by a robust circular border. Mid tibia with two to four anterioventral and two to three posterioventral robust bristles among the vestiture. Hind tibia with several very robust bristles on anterioventral, posterior and posterioventral sides and a longitudinal row of robust bristles, which comprises two thirds to three quarters of the tibia length. Basal tarsomere of all legs with conspicuous robust bristles. All tibial spurs equal in length. Claws with one robust and three minor teeth. Abdomen brown like the thorax or slightly paler, with dark bristles. Hypopygium (Fig. 48) brown. Gonocoxites wider than long; basally not fused, without a basal lobe of bristles. Gonostylus slender, slightly curved inwards, 2.7-3.0 times longer than wide; thickest part in the middle, apically with dense setae and a group of approximately six small spines, spines more or less longitudinally directed; subapically with one short whip-lash hair, which is two times as long as the apical spines. Tegmen wider than long and sclerotized, base very strongly sclerotized with short and robust parameral apodeme; lateral margin of tegmen with a sclerotized edge, which bears several small teeth on middle third; apex straight, sclerotisation widely interrupted; dorsal structures absent. Area of teeth consists of more than 20 conspicuous teeth. Ejaculatory apodeme short, broad, and very dark, with a long, broad base. Posterior portion of gonocoxal apodeme broad and brown. Anterior portion of gonocoxal apodeme small, roundish, darker than posterior portion.

Body size: 1.8-2.4 mm.
Female. Unknown.
Genetic distances. Both available COI sequences are identical and do not show any distance. The nearest neighbour is Psl. wernermohrigi, diverging by a minimum of $1.84 \%$. The one available 28 S sequence is identical to those of Psl. frederickedwardsi, Psl. huttoni, Psl. jaschhofi, Psl. plicitegmenta, Psl. robustotegmenta, Psl. subtilitegmenta, Psl. tonnoiri, and Psl. wernermohrigi.

Distribution. New Zealand.
Discussion. Pseudolycoriella macrotegmenta belongs to those species of the Psl. macrotegmenta complex which bear lateral teeth on the tegmen. Among these species Psl. macrotegmenta is the only species that lacks the dorsal structures on the tegmen and has a gonostylus with a subapical lobe on the inner side.

The author of Psl. macrotegmenta had a wide concept of this species. Accordingly, the paratype series contained two other species: three specimens of Psl. wernermohrigi and one specimen of Psl. frederickedwardsi. The fifth paratype which is deposited in the NZAC (NZAC02016074) was not studied and remains unrevised. This broad concept of Psl. macrotegmenta led to an inaccuracy in the original description. Mohrig \& Jaschhof (1999) gave a length to width ratio of the fourth flagellomere of 2.4 , which is much shorter than the value of 2.9-3.6 revealed by the present study.

## Pseudolycoriella plicitegmenta sp. n.

(Fig. 49)
urn:Isid:zoobank.org:act:NomenclaturalActs/256F08AF-B9EB-40A7-8D03-E59A5DF412F0
Material studied. Holotype male. New Zealand: South Island, Clutha, Catlins, Purakaunui Scenic Reserve, mixed podocarp/broadleaf/Nothofagus forest, Malaise trap, 27.01-05.03.2002, leg. M. \& C. Jaschhof (NZAC, SDEI-Dipt0001158). Paratypes. $4 \widehat{\jmath}$ same locality and same date as holotype ( 2 x NZAC, SDEI-Dipt- 0000560 \& SDEI-Dipt-

0001155; 2x SDEI, SDEI-Dipt-0001130 \& SDEI-Dipt-0001153). $10^{\text {§ }}$ South Island, Clutha, Catlins Coastal Rain Forest Park, Catlins River Valley, silver beech forest, Malaise trap, 27.01-05.03.2002, leg. M. \& C. Jaschhof (SDEI, SDEI-Dipt-0000799).

Description. Male. Head brown; eye bridge three facets wide, two at margin. Flagellomeres brown, concolourous with scape and pedicel; fourth flagellomere 2.3-2.9 times as long as wide; necks of flagellomeres differentiated; surface of flagellomeres rough with deep pits; sensilla of two different lengths present; setae sparsely scattered, approximately as long as flagellomere width, bases of setae slightly raised. Prefrons and clypeus moderately bulging. Maxillary palp three-segmented; first palpomere longest, third longer than second; first palpomere with three to four bristles and an inconspicuous patch of sensilla. Thorax brown, paler than head; laterally brighter. Posterior pronotum bare. Anterior pronotum with seven to eleven bristles. Episternum 1 with five to nine bristles. Mesonotum with five to six longer bristles among the lateral bristles; row of dorsocentral bristles well developed. Scutellum with two robust and some shorter bristles. Katepisternum bicoloured, basal part pale brown, apically brown; as long as high. Wing. Length $2.1-2.3 \mathrm{~mm}$; width/length ratio $0.36-0.40$. Membrane transparent and without macrotrichia, anal area present; all posterior veins distinct, all without macrotrichia; apical fourth of $\mathrm{R}_{5}$ with additional macrotrichia on ventral side; bM bare; r-m with one macrotrichium or without macrotrichia; $\mathrm{R}_{1}$ short, $0.46-0.54$ times as long as R ; M-fork longer than M stem; $\mathrm{c} / \mathrm{w}$ ratio $0.67-0.74 ; \mathrm{r}-\mathrm{m} / \mathrm{bM}$ ratio $1.0-1.3$. Haltere pale brown. Legs pale brown; coxal bristles dark. Fore tibia with one to four bristles among vestiture; tibial organ comprising half of tibial apex, consisting of a transverse patch of bristles surrounded by a robust circular border. Mid tibia with seven to nine robust bristles among vestiture, located anterio- and posterioventrally. Hind tibia with twelve to twenty robust bristles at anterioventral, posterior and posterioventral sides and a longitudinal row of robust bristles, which comprises three-fifths of tibial length. Basal tarsomere of all legs with conspicuous robust bristles. All tibial spurs equal in length. Claws with one robust tooth and several minor teeth. Abdomen. Pale brown, with dark bristles. Hypopygium (Fig. 49) brown. Gonocoxites wider than long, ventrally u-shaped and not fused, without a basal lobe of bristles. Gonostylus elongate, slightly curved inwards, 3.1-3.4 times longer than wide, inner base with a protruding angle ( $b a$ in Fig. 49); apically with dense setae and a group of approximately five to six spines; subapically with two or three medium-sized whip-lash hairs; dorsal side of apex of gonostylus slightly constricted. Tegmen wider than long, base very strongly sclerotized with short, robust parameral apodeme; lateral margin of tegmen with a sclerotized edge, scattered with several small teeth, apically not fused; dorsal structures on tegmen well developed, as long as two-thirds of tegmen length. Area of teeth with 17 to 28 conspicuous teeth. Ejaculatory apodeme short, broad, and very dark, with a broad, u-shaped base. Posterior portion of gonocoxal apodeme broad and brown. Anterior portion of gonocoxal apodeme small.

Body size: 2.1-2.3 mm.
Female. Unknown.
Genetic distances. The maximum p-distance between all five available COI sequences is $0.15 \%$. Some COI sequences are identical with Psl. gonotegmenta and Psl. subtilitegmenta. Both available 28S sequence are identical and do not show any differences to those of Psl. frederickedwardsi, Psl. gonotegmenta, Psl. huttoni, Psl.jaschhofi, Psl. macrotegmenta, Psl. robustotegmenta, Psl. subtilitegmenta, Psl. tonnoiri, and Psl. wernermohrigi.

Etymology. The epithet of the new species is composed of the two Latin words plica (crease) and tegmen (cover; technical term for a part of the sciarid hypopygium).

Distribution. New Zealand.
Discussion. Pseudolycoriella plicitegmenta belongs to those species of the Psl. macrotegmenta complex that bear several teeth on the lateral tegmen margin. This sclerotized margin is apically interrupted. This combination can be also found in Psl. macrotegmenta, Psl. robustotegmenta, and Psl. subtilitegmenta and some specimens of Psl. gonotegmenta. Pseudolycoriella plicitegmenta can be distinguished from Psl. macrotegmenta by the different shape of the gonostylus and the presence of the dorsal structure on the tegmen. Pseudolycoriella gonotegmenta and Psl. robustotegmenta have a $\mathrm{R}_{1} / \mathrm{R}$ ratio greater than 0.6 while the $\mathrm{R}_{1} / \mathrm{R}$ ratio of Psl. plicitegmenta is less than 0.6 . Pseudolycoriella robustotegmenta differs significantly because it has a massive tegmen margin, long dorsal structures on the tegmen, and a long ejaculatory apodeme.

## Pseudolycoriella subtilitegmenta sp. n.

(Fig. 53)
urn:lsid:zoobank.org:act:NomenclaturalActs/7E7F889A-66DB-46DB-8B60-FB995BB62FC9

Material studied. Holotype male. New Zealand: South Island, Clutha, Catlins, Purakaunui Scenic Reserve, mixed podocarp/broadleaf/Nothofagus forest, Malaise trap, 27.01-05.03.2002, leg. M. \& C. Jaschhof (NZAC, SDEI-Dipt-
 Dipt-0001133). $1 \circlearrowleft^{\Uparrow}$ North Island, Ruapehu, Tongariro NP, Mangawhero River Valley 3 km NE Ohakune, altitude 690 m , mixed podocarp/broadleaf forest, Malaise trap, 26.11-28.12.2002, leg. M. \& C. Jaschhof \& U. Kallweit (SDEI).

Description. Male. Head brown; eye bridge three facets wide, two at margin. Flagellomeres brown, concolourous with scape and pedicel; fourth flagellomere 2.5-2.8 times as long as wide; necks of flagellomeres differentiated; surface of flagellomeres rough with deep pits; sensilla of two different lengths present; setae sparsely scattered, approximately as long as flagellomere width, bases of setae on small elevations. Prefrons and clypeus slightly bulging. Maxillary palp three-segmented; first palpomere longest, third longer than second; first palpomere with two to five bristles and an inconspicuous patch of sensilla; one of the bristles on first and second palpomere robust and elongated. Thorax brown, concolourous with head. Posterior pronotum bare. Anterior pronotum with six to nine long bristles. Episternum 1 with five to nine long bristles. Mesonotum with five to six longer bristles among the lateral bristles; row of dorsocentral bristles well developed. Scutellum with two robust and some shorter bristles. Katepisternum slightly longer than high. Wing. Length $2.3-2.4 \mathrm{~mm}$; width/length ratio $0.36-0.38$. Membrane transparent and without macrotrichia, anal area present; all posterior veins distinct, all without macrotrichia; apical $25 \%-40 \%$ of $\mathrm{R}_{5}$ with additional macrotrichia on ventral side; bM bare, $\mathrm{r}-\mathrm{m}$ with one or two macrotrichia; $\mathrm{R}_{1}$ short, $0.64-0.71$ times as long as R ; M-fork longer than M stem; $\mathrm{c} / \mathrm{w}$ ratio $0.67-0.70$; r-m/bM ratio $1.0-1.2$. Haltere pale brown. Legs pale brown, trochanter slightly darker; coxal bristles dark. Fore tibia with one or two bristles among the vestiture; tibial organ comprising half of tibial apex, consisting of a transverse patch of bristles surrounded by a robust circular border. Mid tibia with seven to eight robust bristles among vestiture, located anterio- and posteriorventrally. Hind tibia with 16 to 22 robust bristles on anterioventral, posterior and posterioventral sides and a longitudinal row of robust bristles, which extends along two-thirds of the length of the tibia. Basal tarsomere of all legs with conspicuous robust bristles. All tibial spurs equal in length. Claws with one robust tooth and several minor teeth. Abdomen. Brown, with dark bristles. Hypopygium (Fig. 53) brown. Gonocoxites wider than long, ventrally v-shaped and not fused, without a basal lobe of bristles. Gonostylus elongated, apex slightly angled and slightly tapered; without an angle at the base of the gonostylus, 2.7-3.2 times longer than wide; apically with dense setae and a group of approximately five to seven spines; subapically with two to three medium-sized whip-lash hairs. Tegmen wider than long, base very strongly sclerotized with medium-sized, robust parameral apodeme; lateral margin of tegmen with a fine sclerotized edge, which is separated into four fragments: one lateral fragment with a few lateral teeth and one apicolateral fragment (the paratype from the North Island shows fused fragments of each tegmen side); dorsal structures on tegmen developed, as long as half of the length of the tegmen. Area of teeth absent. Ejaculatory apodeme short, broad, and dark, with a broad and extended v-shaped base. Posterior portion of gonocoxal apodeme broad and brown. Anterior portion of gonocoxal apodeme broad.

Body size: 2.2-2.6 mm.
Female. Unknown.
Genetic distances. All three available COI sequences are identical and do not show any pairwise distance. These COI sequences are identical with that of Psl. gonotegmenta. One available 28 S sequence is identical to those of Psl. frederickedwardsi, Psl. gonotegmenta, Psl. huttoni, Psl. jaschhofi, Psl. macrotegmenta, Psl. plicitegmenta, Psl. robustotegmenta, Psl. tonnoiri, and Psl. wernermohrigi.

Etymology. The epithet of the new species is composed of the two Latin words subtilis (fine, delicate) and tegmen (cover; technical term for a part of the sciarid hypopygium).

Distribution. New Zealand.
Discussion. Pseudolycoriella subtilitegmenta is the only species among the species of the Psl. macrotegmenta complex with lateral teeth and a very thin sclerotized margin on the tegmen. Furthermore, the area of teeth is absent, which is also only found in Psl. tonnoiri and Psl. breviseta. Pseudolycoriella subtilitegmenta is the only species in this group with a tapered gonostylus apex.


FIGURES 48-53. Pseudolycoriella macrotegmenta complex, hypopygia. 48. Pseudolycoriella macrotegmenta Mohrig 49. Pseudolycoriella plicitegmenta sp. n. 50. Pseudolycoriella wernermohrigi sp. n. 51. Pseudolycoriella huttoni sp. n. 52. Pseudolycoriella gonotegmenta sp. n. 53. Pseudolycoriella subtilitegmenta $\mathbf{~ s p . ~ n . ~ A b b r e v i a t i o n s : ~ b a ~}=$ basal angle; ds $=$ dorsal structure on the tegmen; $1 \mathrm{lt}=$ teeth on lateral side of the tegmen.

## Pseudolycoriella gonotegmenta sp. n.

(Fig. 52)
urn:lsid:zoobank.org:act:9BD9CFA2-AA84-4340-BA03-C94C77EC9748

Material studied. Holotype male. New Zealand: South Island, Buller, Rahu Scenic Reserve, 12 km NW Springs Junction, altitude 550 m , mixed Nothofagus/podocarp forest, Malaise trap, 27.11-25.12.2001, leg. M. \& C. Jaschhof (NZAC, SDEI-Dipt-0001495). Paratypes. 2 §̄龴 North Island, Ruapehu, Tongariro NP, Mangawhero River Valley 3 km NE Ohakune, altitude 690 m , mixed podocarp/broadleaf forest, Malaise trap, 26.11-28.12.2002, leg. M. \& C. Jaschhof \& U. Kallweit (all SDEI). $2{ }^{\top} 0^{\lambda}$ North Island, Masterton, Tararua Forest Park, Blue Range Hut Track 10 km SW Mt. Bruce, mixed Nothofagus/podocarp/broadleaf forest, sweep net, 02.02.2002, leg. M. Jaschhof (NZAC; SDEI). $1 \circlearrowleft$ South Island, Buller, Ahaura, Granville State Forest, altitude 170-250 m, Nothofagus truncata forest, Malaise trap, 01.12.1994, leg. J. Hutcheson (NZAC). $2{ }^{\top} 0^{\lambda}$ South Island, Buller, 5 km W Maruia Springs, mixed Nothofagus forest, Malaise trap, 26.11-25.12.2001, leg. M. \& C. Jaschhof (NZAC, SDEI-Dipt-0000977 \& SDEI, SDEI-Dipt-0000984). $1 \circlearrowleft^{\wedge}$ South Island, Buller, Lake Daniells Track 7 km E Springs Junction, mixed red/silver beech forest, Malaise trap, 24.11-26.12.2001, leg. M. \& C. Jaschhof (SDEI).

Description. Male. Head brown; eye bridge three facets wide, two at margin. Flagellomeres brown, concolourous with scape and pedicel; fourth flagellomere 2.6-2.9 times as long as wide; necks of flagellomeres differentiated; surface of flagellomeres rough with deep pits; sensilla of two different lengths present; setae sparse, approximately as long as flagellomere width, bases of setae with slightly raised coronas. Prefrons and clypeus moderately bulging. Maxillary palp three-segmented; first palpomere longest, third longer than second; first palpomere with two to six bristles and an inconspicuous patch of sensilla; usually one of the bristles on first and second palpomere elongated. Thorax brown, laterally extensively brightened. Posterior pronotum bare. Anterior pronotum with five to seven long bristles. Episternum 1 with six to eleven long bristles. Mesonotum with five to six longer bristles among the lateral bristles; row of dorsocentral bristles well developed. Scutellum with two to four robust and some shorter bristles. Katepisternum as long as high. Wing. Length $2.0-2.3 \mathrm{~mm}$; width/length ratio $0.35-0.41$. Membrane transparent and without macrotrichia, anal area present; all posterior veins distinct, all without macrotrichia; apical $25 \%-40 \%$ of $\mathrm{R}_{5}$ with additional macrotrichia on ventral side; bM and r-m bare; $\mathrm{R}_{1}$ short, $0.64-0.77$ times as long as R ; M-fork longer than stem of M or-in one case-as long as stem of M ; $\mathrm{c} / \mathrm{w}$ ratio $0.66-0.71$; r-m/bM ratio 0.7-1.2. Haltere pale brown. Legs pale brown; coxal bristles dark. Fore tibia with one to three bristles among vestiture; tibial organ comprising half of tibial apex, consisting of a transverse patch of bristles surrounded by a robust circular border. Mid tibia with four to six robust bristles among vestiture, located anterio- and posterioventrally. Hind tibia with seven to 17 robust bristles on anterioventral, posterior and posterioventral sides and a longitudinal row of robust bristles, which extends two-thirds to four-fifths of tibial length. Basal tarsomere of all legs with conspicuous robust bristles. All tibial spurs equal in length. Claws with one robust tooth and several minor teeth. Abdomen. Pale brown, with dark bristles. Hypopygium (Fig. 52) brown. Gonocoxites wider than long, ventrally u-shaped and not fused, without a basal lobe of bristles. Gonostylus elongate, apically curved inwards, apex slightly tapered, without an angle at the base of the gonostylus, 2.6-3.3 times longer than wide; apically with dense setae and a group of approximately five to six spines; subapically with one to three medium-sized whip-lash hairs. Tegmen wider than long, base very strongly sclerotized with short, robust parameral apodeme; lateral margin of tegmen with a sclerotized edge, scattered with several small teeth, apically mostly fused (two exceptions with membranous gaps at apex); dorsal structures on tegmen well developed, as long as half tegmen length. Area of teeth with seven to 40 conspicuous teeth. Ejaculatory apodeme short, broad, and dark. Posterior portion of gonocoxal apodeme broad and brown. Anterior portion of gonocoxal apodeme broad and short.

Body size: 1.7-2.4 mm.
Female. Unknown.
Genetic distances. The maximum p-distance between all three available COI sequences is $0.20 \%$. Some of these COI sequences are identical with that of Psl. plicitegmenta. The one available 28 S sequence is identical to those of Psl. frederickedwardsi, Psl. huttoni, Psl. jaschhofi, Psl. macrotegmenta, Psl. plicitegmenta, Psl. robustotegmenta, Psl. subtilitegmenta, Psl. tonnoiri, and Psl. wernermohrigi.

Etymology. The epithet of the new species is composed of the Greek word $\gamma \omega v o \zeta$ (gonos, edge) and the Latin word tegmen (cover; technical term for a part of the sciarid hypopygium).

Distribution. New Zealand.
Discussion. With the margin of the tegmen apically fused, Psl. gonotegmenta resembles Psl. huttoni and Psl.
wernermohrigi. Pseudolycoriella huttoni differs by having a u-shaped base of the ejaculatory apodeme, a lower $\mathrm{R}_{1} / \mathrm{R}$ ratio ( 0.56 vs. $0.64-0.77$ ), and a smaller anterior portion of gonocoxal apodeme. Pseudolycoriella wernermohrigi has a much longer ejaculatory apodeme, a longer parameral apodeme, and a longer anterior portion of gonocoxal apodeme. In specimens where fragmentation of the tegmen margin has occurred, as for example in two paratypes of Psl. gonotegmenta, the base of the ejaculatory apodeme has to be examined to enable separation from Psl. plicitegmenta. In Psl. gonotegmenta this base is long and broadly v-shaped while Psl. plicitegmenta has a short a u-shaped base. In addition Psl. plicitegmenta possess a basal angle on the gonostylus, which is lacking in Psl. gonotegmenta.

## Pseudolycoriella robustotegmenta sp. n.

(Figs 7 \& 54)
urn:lsid:zoobank.org:act:NomenclaturalActs/3346FE28-2271-4E55-8DB1-4CA3ED714633
Material studied. Holotype male. New Zealand: North Island, Bay of Plenty, Katikati, Uretara mangrove area, Malaise trap, 04.09-22.10.2016, leg. P.A. Maddison (NZAC, SDEI-Dipt-0001358). Paratype. 1 ${ }^{\top}$, North Island, Bay of Plenty, Katikati, Te Mania catchment, Malaise trap, 19.09-27.10.2016, leg. P.A. Maddison (SDEI, SDEI-Dipt-0001371).

Description. Male. Head (Fig. 7) brown; eye bridge two to three facets wide. Flagellomeres brown, concolourous with scape and pedicel; fourth flagellomere 2.7-2.9 times as long as wide; necks of flagellomeres differentiated; surface of flagellomeres rough with deep pits; sensilla of two different lengths present; setae sparse, approximately as long as flagellomere width, bases of setae with coronas. Prefrons and clypeus moderately bulging. Maxillary palp three-segmented; first palpomere longest, third longer than second; first palpomere with three to four bristles and an inconspicuous patch of sensilla; first and second palpomere each with one longer bristle. Thorax brown, laterally brighter. Posterior pronotum bare. Anterior pronotum with four to seven bristles. Episternum 1 with five to six bristles. Mesonotum with five longer lateral bristles; row of dorsocentral bristles present. Scutellum with two robust and some shorter bristles. Katepisternum brown, basally slightly brightened; as long as high. Wing. Length $2.0-2.2 \mathrm{~mm}$; width/length ratio $0.37-0.38$. Membrane transparent and without macrotrichia, anal area present; all posterior veins distinct, all without macrotrichia; apical fourth of $R_{5}$ with five to six additional macrotrichia on ventral side; bM bare; r-m with one macrotrichium (exceptionally the left wing of the holotype shows two macrotrichia on r-m); $\mathrm{R}_{1}$ short, $0.55-0.57$ times as long as R ; M-fork shorter than M stem; $\mathrm{c} / \mathrm{w}$ ratio $0.68-0.70$; r-m 0.8 times as long as bM. Haltere pale brown. Legs pale brown; coxal bristles dark; trochanter darkened. Fore tibia with one robust bristle among the vestiture; tibial organ comprising more than half of the tibial apex, consisting of a patch of bristles surrounded by a robust circular border. Mid tibia with two anterioventral and two posterioventral robust bristles among the vestiture. Hind tibia with several robust bristles on anterioventral, posterior and posterioventral sides and a longitudinal row of robust bristles, which extends along four fifths of the tibial length. Basal tarsomere of all legs with conspicuous robust bristles. All tibial spurs equal in length. Claws with one robust tooth and several minor teeth. Abdomen. Pale brown, with long dark bristles. Hypopygium (Fig. 54) brown. Gonocoxites wider than long, ventrally not fused, without a basal lobe of bristles. Gonostylus long and evenly curved, 2.8-3.1 times longer than wide; apically with dense setae and a group of approximately four to five spines; subapically with two to three medium-sized whip-lash hairs; inner base of gonostylus with an angle. Tegmen strongly sclerotized, wider than long, base very strongly sclerotized with long parameral apodeme; lateral margin of tegmen with a broad sclerotized edge; laterally scattered with several small teeth; sclerotized edge interrupted apically; dorsal structures well developed, reaching apex of the tegmen. Area of teeth with ten to twenty conspicuous teeth. Ejaculatory apodeme medium-sized, broad, and very dark, with a broad, u-shaped base. Posterior portion of gonocoxal apodeme broad and brown. Anterior portion of gonocoxal apodeme broad and short.

Body size: 2.2-2.3 mm.
Female. Unknown.
Genetic distances. Both available COI sequences are identical and do not show any p-distances. The nearest neighbour is Psl. gonotegmenta, diverging by a minimum of $0.61 \%$. Both available 28 S sequence are identical and do not show any differences to those of Psl. frederickedwardsi, Psl. gonotegmenta, Psl. huttoni, Psl.jaschhofi, Psl. macrotegmenta, Psl. plicitegmenta, Psl. subtilitegmenta, Psl. tonnoiri, and Psl. wernermohrigi.

Etymology. The name of the newly described species is composed of the two Latin words robustus (robust) and tegmen (cover; technical term for a part of the sciarid hypopygium).

Distribution. New Zealand.
Discussion. Pseudolycoriella robustotegmenta can be distinguished from the remaining species of this complex by the strongly developed margin of the tegmen, robust dorsal structures on the tegmen reaching the apex, and the base of vein M longer than the M -fork.


FIGURE 54. Pseudolycoriella robustotegmenta sp. n., hypopygium.

## Pseudolycoriella huttoni sp. n.

(Fig. 51)
urn:lsid:zoobank.org:act:6434BD19-1F69-47FD-A8FB-5A73560A1082

Material studied. Holotype male. New Zealand: North Island, Taupo, Tongariro NP, Mangawhero River Valley 3 km NE Ohakune, altitude 690 m , podocarp/broadleaf forest; Malaise trap, 26.11-28.12.2002, leg. M. \& C. Jaschhof \& U. Kallweit (NZAC, SDEI-Dipt-0000621).

Description. Male. Head brown; eye bridge two to three facets wide. Flagellomeres brown, concolourous with scape and pedicel; fourth flagellomere 2.9 times as long as wide; necks of flagellomeres differentiated; surface of flagellomeres rough with deep pits; sensilla of two different lengths present; setae sparse, approximately as long as flagellomere width, bases of setae with slightly raised coronas. Prefrons and clypeus moderately bulging. Maxillary palp three-segmented; first and third palpomere equal in length, longer than second; first palpomere with five bristles and an inconspicuous patch of sensilla; first and second palpomere with one longer outward orientated bristle. Thorax brown, lateral slightly brightened. Posterior pronotum bare. Anterior pronotum with five bristles. Episternum 1 with nine bristles. Mesonotum with five longer lateral bristles; row of dorsocentral bristles present. Scutellum with two robust and some shorter bristles. Katepisternum nearly as long as high. Wing. Length 2.3 mm ; width/length ratio 0.38 . Membrane transparent and without macrotrichia, anal area present; all posterior veins distinct, except weak stem of $M$, all without macrotrichia; apical third of $R_{5}$ with nine additional macrotrichia on ventral side; bM bare; left r-m bare, right r-m with one macrotrichium; $\mathrm{R}_{1}$ short, 0.56 times as long as R ; M-fork longer than M stem; c/w ratio 0.70 ; r-m 0.8 times as long as bM. Haltere paler brown than thorax. Legs pale brown; coxal bristles dark; trochanter slightly darker. Fore tibia with one robust bristle among vestiture; tibial organ comprising more than half of tibial apex, consisting of a patch of bristles surrounded by a robust circular border. Mid tibia with six robust bristles among the vestiture, located on anterioventral and posterioventral side. Hind tibia with several robust bristles at anterioventral, posterior and posterioventral sides and a longitudinal row of robust bristles, which comprises one half of tibial length. Basal tarsomere of all legs with conspicuous robust bristles. All tibial spurs equal in length. Claws with one robust tooth and several minor teeth. Abdomen. Pale brown, with long dark bristles.

Hypopygium (Fig. 51) brown. Gonocoxites wider than long, ventrally not fused, without a basal lobe of bristles. Gonostylus long and apically curved, giving it a hump-backed appearance, 2.8 times longer than wide; apically with dense setae and a group of approximately five spines; subapically with three medium-sized whip-lash hairs; inner base of gonostylus without a remarkable angle. Tegmen strongly sclerotized, slightly wider than long, base very strongly sclerotized with long parameral apodemes; lateral margin of tegmen with a sclerotized edge; laterally scattered with several small teeth; sclerotized edge surrounds apex of tegmen; dorsal structures well developed, as long as two-thirds of tegmen. Area of teeth present, comprising fifteen teeth. Ejaculatory apodeme short, broad, and very dark, with a long, u-shaped, pale brown base. Posterior portion of gonocoxal apodeme broad and brown. Anterior portion of gonocoxal apodeme very short.

Body size: 2.6 mm .
Female. Unknown.
Genetic distances. Only one COI sequence was available. The nearest neighbour is Psl. tonnoiri, diverging by a minimum of $2.74 \%$. The one available 28S sequence is identical to those of Psl. frederickedwardsi, Psl.jaschhofi, Psl. macrotegmenta, Psl. plicitegmenta, Psl. robustotegmenta, Psl. subtilitegmenta, Psl. tonnoiri, and Psl. wernermohrigi.

Etymology. The epithet was selected to honour Frederick Wollaston Hutton (1836-1905), who described the first New Zealand sciarid fly in 1881.

Distribution. New Zealand.
Discussion. Pseudolycoriella huttoni is morphologically very similar to Psl. wernermohrigi, but shows a considerable difference in the genetic analysis (Fig. 61). As reliable distinguishing characters the length of the ejaculatory apodeme and the length of the anterior portion of gonocoxal apodeme can be used.

## Pseudolycoriella wernermohrigi sp. n.

(Fig. 50)
urn:Isid:zoobank.org:act:NomenclaturalActs/F62E7CA7-79FB-44C4-AC2C-FD9403DDAD27
Literature. Pseudolycoriella macrotegmenta Mohrig, 1999: Mohrig \& Jaschhof (1999): 41-43 [misidentification].

Material studied. Holotype male. New Zealand: North Island, Western Bay of Plenty, Katikati, Te Mania catchment, Malaise trap, 19.09-27.10.2016, leg. P.A. Maddison (NZAC, SDEI-Dipt-0001223). Paratypes. 10 §§ same locality and same date as holotype (5x NZAC, SDEI-Dipt-0000670, SDEI-Dipt-0000674, SDEI-Dipt-0001227, SDEI-Dipt-0001238 \& SDEI-Dipt-0001245; 5x SDEI, SDEI-Dipt-0000665, SDEI-Dipt-0001234, SDEI-Dipt0001334 , SDEI-Dipt-0001235 \& SDEI-Dipt-0001373). $1 \precsim$ North Island, Auckland, Pukororo Miranda, Te Kama forest remnant, litter, 13.11.2014, leg. P.A. Maddison (NZAC, SDEI-Dipt-0001270). 2 § ${ }^{\lambda}$ North Island, ThamesCoromandel, Coromandel Range, 5 km east of Coromandel, dense Podocarpus secondary wood with tree ferns, sweep net, 28.12.1992, leg. M. Jaschhof (PWMP; SDEI [both specimens previously misidentified, published as paratypes of Psl. macrotegmenta in Mohrig \& Jaschhof 1999]). $1 \sigma^{\lambda}$ § North Island, Wairoa, Urewera National Park, Huiarau Range 30 km southeast of Murupara, altitude $600-1,000 \mathrm{~m}$, Podocarpus-Nothofagus wood, sweep net, 23.12.1992, leg. M. Jaschhof (PWMP [previously misidentified, published as paratype of Psl. macrotegmen$t a$ in Mohrig \& Jaschhof 1999]). $1 \sigma^{\lambda}$ North Island, Ruapehu, Tongariro NP, Mangawhero River Valley 3 km NE Ohakune, altitude 690 m , mixed podocarp/broadleaf forest, malaise trap, 26.11-28.12.2002, leg. M. \& C. Jaschhof \& U. Kallweit (SDEI). $7 \delta^{\lambda} \delta^{\lambda}$ North Island, Taupo, Pureora Forest Park, Waipapa Reserve, altitude 600 m , mixed podocarp/broadleaf forest, sweep net, 04-05.02.2002, leg. M. Jaschhof (3x NZAC, SDEI-Dipt-0000824; 4x SDEI, SDEI-Dipt-0000822 \& SDEI-Dipt-0000825). Additional material. $2 \delta^{\lambda}$ § same locality and same date as holotype (all SDEI, SDEI-Dipt-0000681 \& SDEI-Dipt-0001232).

Description. Male. Head brown; eye bridge two to three facets wide. Flagellomeres brown, concolourous with scape and pedicel; fourth flagellomere 2.5-2.9 times as long as wide; necks of flagellomeres differentiated; surface of flagellomeres rough with deep pits; sensilla of two different length present; setae sparse, approximately as long as flagellomere width, bases of setae with slightly raised coronas. Prefrons and clypeus bulging. Maxillary palp threesegmented; first palpomere longest, third longer than second; first palpomere with two to six bristles and an inconspicuous patch of sensilla; first and second palpomeres each with one longer outward orientated bristle. Thorax brown, laterally brighter. Posterior pronotum bare. Anterior pronotum with four to seven bristles. Episternum 1 with
five to seven bristles. Mesonotum with five to seven longer lateral bristles; row of dorsocentral bristles present. Scutellum with two robust and some shorter bristles. Katepisternum brown; as long as high. Wing. Length $2.0-2.3 \mathrm{~mm}$; width/length ratio $0.35-0.42$. Membrane transparent and without macrotrichia, anal area present; all posterior veins distinct, all without macrotrichia; apical quarter of $\mathrm{R}_{5}$ with two to five additional macrotrichia on ventral side; bM and r-m bare (exceptionally one of the misidentified paratypes from the type series of Psl. macrotegmenta bears one macrotrichium on $\mathrm{r}-\mathrm{m}$ on the right wing); $\mathrm{R}_{1}$ short, $0.51-0.71$ times as long as R ; M-fork longer than M stem; $\mathrm{c} / \mathrm{w}$ ratio $0.68-0.74$; r-m $0.8-1.1$ times as long as bM . Haltere brown. Legs pale brown; coxal bristles dark; trochanter darkened. Fore tibia mostly with one to two robust bristles among the vestiture, two exceptions with no bristles among vestiture; tibial organ comprising more than half of tibial apex, consisting of a patch of bristles surrounded by a robust circular border. Mid tibia with five to six robust bristles among vestiture, located on anterioventral and posterioventral side. Hind tibia with several robust bristles on anterioventral, posterior and posterioventral sides and a longitudinal row of robust bristles, which comprises three quarters of tibial length. Basal tarsomere of all legs with conspicuous robust bristles. All tibial spurs equal in length. Claws with one robust tooth and several minor teeth. Abdomen. Pale brown, with long dark bristles. Hypopygium (Fig. 50) brown. Gonocoxites wider than long, ventrally not fused, without a basal lobe of bristles. Gonostylus long and apically curved, giving it a hump-backed appearance, 2.6-3.2 times longer than wide; apically with dense setae and a group of approximately five to seven spines; subapically with two medium-sized whip-lash hairs; inner base of gonostylus without a noticeable angle. Tegmen strongly sclerotized, slightly wider than long, base very strongly sclerotized with long parameral apodemes; lateral margin of tegmen with a sclerotized edge; laterally scattered with several small teeth; sclerotized edge surrounds apex of tegmen, exceptionally with a small interruption apically; dorsal structures well developed, at least as long as two-thirds of tegmen or reaching apex of the tegmen. Area of teeth mostly impoverished, without teeth or with a few tiny teeth (maximally up to fifteen; one exception with twenty teeth). Ejaculatory apodeme long, broad, and very dark, with a long base. Posterior portion of gonocoxal apodeme broad and brown. Anterior portion of gonocoxal apodeme narrow and long.

Body size: 2.0-2.5 mm.
Female. Unknown.
Genetic distances. The maximum p-distance between all twelve available COI sequences is $1.82 \%(0.61 \%$, if the specimens SDEI-Dipt-0000681 \& SDEI-Dipt-0001232 are omitted). The nearest neighbour is Psl. macrotegmenta, diverging by a minimum of $1.84 \%$. The maximum p-distance between all twelve available 28 S sequences is $0.06 \%$. Some of these sequences are identical to those of Psl. frederickedwardsi, Psl. gonotegmenta, Psl. huttoni, Psl. jaschhofi, Psl. macrotegmenta, Psl. plicitegmenta, Psl. robustotegmenta, Psl. subtilitegmenta, and Psl. tonnoiri.

Etymology. The newly described species is named after Prof. Dr Werner Mohrig (1937-2019), who had fruitfully worked on sciarid taxonomy for over five decades.

Distribution. New Zealand.
Discussion. Three of the paratypes of Psl. macrotegmenta, designated by Mohrig \& Jaschhof (1999), actually represent Psl. wernermohrigi. Both cryptic species share the remarkable lateral teeth on the tegmen margin with five other species (Psl. dentitegmenta, Psl. huttoni, Psl. plicitegmenta, Psl. gonotegmenta, and Psl. subtilitegmenta). However, unlike Psl. macrotegmenta, the sclerotized margin of the tegmen of Psl. wernermohrigi is broad and surrounds the apex without any gap. This character can also be found in Psl. huttoni, but compared to this species and the other species with lateral teeth on the tegmen, Psl. wernermohrigi has the longest ejaculatory apodeme. Only Psl. dentitegmenta has an ejaculatory apodeme of a similar length, but has a tegmen with major differences.

Two specimens (SDEI-Dipt-0000681 \& SDEI-Dipt-0001232) were identified as Psl. wernermohrigi based on morphology. However, their genetic data differ from other conspecific specimens, which were also barcoded. Their highest p-distance for the COI gene with regard to other Psl. wernermohrigi specimens is $1.82 \%$, which is a high value in this species complex. Consequently, both specimens are in a sister-group relationship with respect to the remaining Psl. wernermohrigi specimens (Fig. 61). Thus, the existence of a further species cannot be completely ruled out. Both specimens were treated as additional material and not included in the type series.

## Pseudolycoriella dentitegmenta sp. n.

(Fig. 60)
urn:Isid:zoobank.org:act:4C5B54A5-1BA7-478F-9879-5C3DCCEF1815

Material studied. Holotype male. New Zealand: North Island, Taupo, Pureora Forest, Select Loop Road, Malaise trap, 15.07-23.08.2001, leg. M. \& C. Jaschhof (NZAC, SDEI-Dipt-0000594).

Description. Male. Head brown; eye bridge one to three facets wide. Flagellomeres brown, paler than head, concolourous with scape and pedicel; fourth flagellomere 3.1 times as long as wide; necks of flagellomeres differentiated; surface of flagellomeres rough with deep pits; sensilla of two different lengths present; setae sparse, approximately as long as flagellomere width, bases of setae with slightly raised coronas. Prefrons and clypeus moderately bulging. Maxillary palp three-segmented; first palpomere longest, third slightly longer than second; first palpomere with three bristles and an inconspicuous patch of sensilla. Thorax brown, slightly paler than head. Posterior pronotum bare. Anterior pronotum with seven bristles. Episternum 1 with ten bristles. Mesonotum with five longer bristles among the lateral bristles; row of dorsocentral bristles well developed. Scutellum with two robust and some shorter bristles. Katepisternum bicoloured, basal part pale brown, apically brown; as long as high. Wing. Length 2.9 mm ; width/length ratio 0.38 . Membrane transparent and without macrotrichia, anal area present; all posterior veins distinct, all without macrotrichia; apical third of $\mathrm{R}_{5}$ with additional macrotrichia on ventral side; bM bare; r-m with one macrotrichium; $\mathrm{R}_{1}$ short, 0.71 times as long as R ; M -fork longer than M stem; $\mathrm{c} / \mathrm{w}$ ratio 0.71 ; r-m and $b M$ equal in length. Haltere pale brown. Legs pale brown; coxal bristles dark. Fore tibia without bristles among vestiture; tibial organ comprising half of tibial apex, consisting of a transverse patch of bristles surrounded by a robust circular border. Mid tibia with two robust bristles; one anterioventrally and one posterioventrally among the vestiture. Hind tibia with several robust bristles on anterioventral, posterior and posterioventral sides and a longitudinal row of robust bristles, which extends two-thirds of tibial length. Basal tarsomere of all legs with conspicuous robust bristles. All tibial spurs equal in length. Claws with one robust tooth and several minor teeth. Abdomen. Pale brown, with dark bristles. Hypopygium (Fig. 60) brown. Gonocoxites wider than long, ventrally v-shaped and not fused, without a basal lobe of bristles. Gonostylus long and slender, curved slightly inwards, 3.9 times longer than wide; apically with dense setae and a group of approximately six spines; subapically with two medium-sized whiplash hairs. Tegmen longer than wide, base very strongly sclerotized with medium-sized, robust parameral apodeme; lateral margin of tegmen with a sclerotized margin; several small teeth scattered on lateral and subapical margin; apex of tegmen membranous; dorsal structures on tegmen indicated. Area of teeth with 15 conspicuous teeth. Ejaculatory apodeme medium-sized, broad, and very dark, with a broad, horseshoe-shaped base. Posterior portion of gonocoxal apodeme broad and brown. Anterior portion of gonocoxal apodeme not present.

Body size: 2.8 mm .
Female. Unknown.
Genetic distances. No genetic data available.
Etymology. The epithet of the new species is composed of the two Latin words dentis (teeth) and tegmen (cover; technical term for a part of the sciarid hypopygium).

Distribution. New Zealand.
Discussion. Due to the presence of several small spines and two whip-lash hairs on the gonostylus Psl. dentitegmenta can easily be recognized as a species of the Psl. macrotegmenta group. However, it is unique because of the long tegmen with many scattered subapical teeth. Furthermore Psl. dentitegmenta has the longest body length, wing length, and length-width ratio of the gonostylus in this species complex.

## Pseudolycoriella breviseta Mohrig, 1999

(Fig. 59)
Literature. Pseudolycoriella breviseta Mohrig, 1999: Mohrig \& Jaschhof (1999): 41, Fig. 21 a-f.-Macfarlane et al. (2010): 441.

Material studied. Holotype male. New Zealand: North Island, Northland, Waipoua Forest, 45 km northwest of Dargaville, sweep net, 31.12.1992, leg. M. Jaschhof (PWMP). Paratypes. $3 \delta^{\lambda} \widehat{\pi}$ same location and same date as holotype (all PWMP). $2 \AA^{\Uparrow}{ }^{\lambda}$ North Island, Northland, Puketi Forest, Waipapa River Valley, sweep net, 01-02.01.1993, leg. M. Jaschhof (PWMP \& SDEI).

Description. See Mohrig \& Jaschhof (1999).
Genetic distances. No genetic data available.
Remarks. Due to the numerous short spines on the gonostylus and the short whip-lash hairs Psl. breviseta belongs to the Psl. macrotegmenta complex. The lateral projections on the tegmen are unique among the species of the genus Pseudolycoriella. Unfortunately, no new records could be obtained; consequently, no genetic information for this species is available. According to the New Zealand Arthropod Collection (2019) the sixth paratype of Psl. breviseta (NZAC02016075) that was not examined in the present study is labelled with a different sampling locality compared to Mohrig \& Jaschhof (1999). Instead of Puketi Forest the sampling locality is given as East Egmont (North Island, Taranaki, Mount Egmont National Park), which would be the southernmost locality of this species.

## Pseudolycoriella frederickedwardsi Köhler, 2016

(Fig. 57)
Literature. Pseudolycoriella macrotegmenta Mohrig, 1999: Mohrig \& Jaschhof (1999): 41-43 [misidentification].-Pseudolycoriella frederickedwardsi Köhler, 2016: Köhler \& Mohrig (2016): 101-103, Fig. 5 A-D.

Material studied. Previously published material: $1 \delta$ New Zealand: North Island, Thames-Coromandel, Coromandel Range, 5 km east of Coromandel, Podocarpus wood, 28.12.1992, leg. M. Jaschhof (PWMP [misidentified, wrongly designated as paratype Psl. macrotegmenta in Mohrig \& Jaschhof (1999)]). New records. 7§̊ North Island, Ruapehu, Tongariro NP, Mangawhero River Valley 3 km NE Ohakune, altitude 690 m , mixed podocarp/ broadleaf forest [type locality], sweep net, 03-04.02.2002, leg. M. Jaschhof (3x NZAC, SDEI-Dipt-0000945, SDEI-Dipt-0000947 \& SDEI-Dipt-0001530; 4x SDEI, SDEI-Dipt-0000772, SDEI-Dipt-0001531 \& SDEI-Dipt0001535). 7 § ${ }^{\text {§ }}$ North Island, same locality as previous, Malaise trap, 26.11-28.12.2002, leg. M. \& C. Jaschhof \& U. Kallweit (3x NZAC, SDEI-Dipt-0001221, SDEI-Dipt-0001224 \& SDEI-Dipt-0001537; 4x SDEI, SDEI-Dipt0000756, SDEI-Dipt-0001237, SDEI-Dipt-0001241 \& SDEI-Dipt-0001247). 5 § §, North Island, Taupo, Pureora Forest, Select Loop Road, altitude 500 m , mixed mature podocarp forest, Malaise trap, 15.07-23.08.2001, leg. M. \& C. Jaschhof (2x NZAC, SDEI-Dipt-0000568 \& SDEI-Dipt-0001507; 3x SDEI, SDEI-Dipt-0001508, SDEI-Dipt0001509 \& SDEI-Dipt-0001510). $4 \circlearrowleft^{\top}$ North Island, Taupo, Pureora Forest Park, Waipapa Reserve, altitude 600 m , mixed podocarp/broadleaf forest, sweep net, 04-05.02.2002, leg. M. Jaschhof (1x NZAC; 3x SDEI, SDEI-Dipt0000823).

Description. See Köhler \& Mohrig (2016).
Genetic distances. The maximum p-distance between all eleven available COI sequences is $0.46 \%$. The nearest neighbour is Psl. huttoni, diverging by a minimum of $2.75 \%$. The maximum p-distance between all twelve available 28 S sequences is $0.17 \%$. Some of these sequences are identical to those of Psl. gonotegmenta, Psl. huttoni, Psl. jaschhofi, Psl. macrotegmenta, Psl.plicitegmenta, Psl. robustotegmenta, Psl. subtilitegmenta, Psl. tonnoiri, and Psl. wernermohrigi.

Remarks. Among the paratypes of Psl. macrotegmenta, designated by Mohrig \& Jaschhof (1999), there was one specimen of Psl. frederickedwardsi (see Psl. macrotegmenta). Dorsal structures on the tegmen, which occur in several species of this complex, are absent.

## Pseudolycoriella tonnoiri Köhler, 2016

(Fig. 58)
Literature. Pseudolycoriella tonnoiri Köhler, 2016: Köhler \& Mohrig (2016): 104-106, Fig. 6 A-D.
Material studied. New records. $6{ }^{\lambda} \delta^{\lambda}$ New Zealand: North Island, Ruapehu, Tongariro NP, Mangawhero River Valley 3 km NE Ohakune, altitude 690 m , mixed podocarp/broadleaf forest, sweep net, 03-04.02.2002, leg. M. Jaschhof (3x NZAC, SDEI-Dipt-0000813, SDEI-Dipt-0000856 \& SDEI-Dipt-0001532; 3x SDEI, SDEI-Dipt0001534 ). $3 \delta^{\Uparrow}{ }^{\text {® }}$ same locality as previous, Malaise trap, 26.11-28.12.2002, leg. M. \& C. Jaschhof \& U. Kallweit (1x NZAC, SDEI-Dipt-0001536; 2x SDEI, SDEI-Dipt-0000597 \& SDEI-Dipt-0000980). 2 § ${ }^{\Uparrow}$ South Island, Selwyn,


FIGURES 55-60. Pseudolycoriella macrotegmenta complex, hypopygia. 55. Pseudolycoriella fiordlandia sp. n. 56. Pseudolycoriella jaschhofi sp. n. 57. Pseudolycoriella frederickedwardsi Köhler 58. Pseudolycoriella tonnoiri Köhler 59. Pseudolycoriella breviseta Mohrig 60. Pseudolycoriella dentitegmenta sp. n.

Cass，Middle Bush，Nothofagus solandri forest，stream site，Malaise trap，17．11．1998，leg．P．M．Johns（NZAC， SDEI－Dipt－0000719；SDEI，SDEI－Dipt－0000722）． $3{ }^{\top} \circlearrowleft^{\lambda}$ South Island，Clutha，Catlins，Purakaunui Scenic Reserve， mixed podocarp／broadleaf／Nothofagus forest，Malaise trap，27．01－05．03．2002，M．\＆C．Jaschhof（1x NZAC，SDEI－ Dipt－0000531；2x SDEI，SDEI－Dipt－0001140 \＆SDEI－Dipt－0001152）．

Description．See Köhler \＆Mohrig（2016）．
Genetic distances．The maximum p－distance between all eleven available COI sequences is $0.15 \%$ ．The nearest neighbour is Psl．gonotegmenta，diverging by a minimum of $2.75 \%$ ．All four available 28 S sequences are identical and do not show any differences to those of Psl．frederickedwardsi，Psl．gonotegmenta，Psl．huttoni，Psl．jaschhofi， Psl．macrotegmenta，Psl．plicitegmenta，Psl．robustotegmenta，Psl．subtilitegmenta，and Psl．wernermohrigi．

Remarks．The presence of the dorsal structures on the tegmen has to be added to the description of Köhler \＆ Mohrig（2016）．These structures do not exceed half the length of the tegmen．

## Pseudolycoriella jaschhofi sp．n．

（Fig．56）
urn：Isid：zoobank．org：act：6010CE99－5267－410E－8C40－D522E7D378B3

Material studied．Holotype male．New Zealand：South Island，Buller，Lewis Pass，altitude 850 m ，tussock grassland at beech forest edge，Malaise trap，26．11－25．12．2001，leg．M．\＆C．Jaschhof（NZAC，SDEI－Dipt－0001199）．Para－ types． $12 \widehat{\sigma}^{\lambda}$ d same locality and same date as holotype（6x NZAC，SDEI－Dipt－0001036，SDEI－Dipt－0001207，SDEI－ Dipt－0001209 \＆SDEI－Dipt－0001210；6x SDEI，SDEI－Dipt－0001198，SDEI－Dipt－0001201，SDEI－Dipt－0001203 \＆SDEI－Dipt－0001217）．5 đ ${ }^{\top}$ South Island，Southland，Fiordland，Fiordland NP， 4 km E Milford Sound，mixed broadleaf／Nothofagus forest，sweep net，07．01．2002，leg．M．Jaschhof（3x NZAC，SDEI－Dipt－0001332，SDEI－Dipt－ 0001341 \＆SDEI－Dipt－0001342；2x SDEI，SDEI－Dipt－0000892 \＆SDEI－Dipt－0000896）． 8 § § South Island，South－ land，Fiordland，Fiordland NP，Hollyford River Valley，Moraine Creek Track，mixed podocarp／Nothofagus forest， Malaise trap，05－24．01．2002，M．\＆C．Jaschhof（4x NZAC，SDEI－Dipt－0001343，SDEI－Dipt－0001345，SDEI－ Dipt－0001512，SDEI－Dipt－0001513；4x SDEI，SDEI－Dipt－0001344，SDEI－Dipt－0001511，SDEI－Dipt－0001514 \＆ SDEI－Dipt－0001515）． 8 入入 South Island，Southland，Otago Lakes，Fiordland NP，Eglinton River Valley，Deer Flat， Nothofagus forest，Malaise trap，04－24．01．2002，leg．M．\＆C．Jaschhof（ $4 x$ NZAC；4x SDEI）． $1 \delta^{\text {§ }}$ same locality as previous，Malaise trap，01－20．12．2002，leg，M．\＆C．Jaschhof \＆U．Kallweit（SDEI，SDEI－Dipt－0000962）．6 § § South Island，Southland，Otago Lakes，Fiordland NP，Eglinton River Valley，opposite Earl Mountain Tracks，Not－ hofagus forest，Malaise trap，05－24．01．2002，leg．M．\＆C．Jaschhof（3x NZAC，SDEI－Dipt－0000848，SDEI－Dipt－ 0000850 \＆SDEI－Dipt－0000851；3x SDEI，SDEI－Dipt－0000852，SDEI－Dipt－0000868 \＆SDEI－Dipt－0000872）． $4{ }^{\top}$ ふ South Island，Southland，Otago Lakes，Fiordland NP，Hollyford River Valley S Divide Creek，mixed Nothofa－ $g u s / b r o a d l e a f ~ f o r e s t, ~ M a l a i s e ~ t r a p, ~ 06-24.01 .2002, ~ l e g . ~ M . ~ \& ~ C . ~ J a s c h h o f ~(2 x ~ N Z A C ; ~ 2 x ~ S D E I) . ~$.

Description．Male．Head brown；eye bridge one to three facets wide．Flagellomeres concolourous with scape and pedicel；fourth flagellomere 2．6－3．1 times as long as wide；necks of flagellomeres differentiated；surface of flagellomeres rough with deep pits；sensilla of two different length；setae sparsely scattered，approximately as long as flagellomere width，bases of setae with slightly raised coronas．Prefrons and clypeus moderately bulging．Maxil－ lary palp three－segmented；first palpomere longest，third slightly longer than second；first palpomere with up to five bristles and an inconspicuous small patch of sensilla．Thorax pale brown，paler than head．Posterior pronotum bare． Anterior pronotum with two to five bristles．Episternum 1 with four to ten bristles．Mesonotum with several longer bristles among the laterals and among the dorsocentral bristles．Scutellum with four robust and some shorter bristles． Katepisternum bicoloured，basal part pale brown，apical third brown；as long as high．Wing．Length $1.9-2.2 \mathrm{~mm}$ ； width／length ratio $0.35-0.41$ ．Membrane transparent and without macrotrichia，anal area present；all posterior veins distinct except faint stem of $M$ ，all without macrotrichia；apical quarter of $R_{5}$ with two to seven additional macro－ trichia on ventral side； bM and r－m bare， $\mathrm{r}-\mathrm{m}$ exceptionally with one to two macrotrichia； $\mathrm{R}_{1}$ short， $0.67-0.80$ times as long as R ； M －fork longer than stem of M ； $\mathrm{c} / \mathrm{w}$ ratio $0.67-0.72$ ；r－m and bM equal in length．Haltere pale brown． Legs pale brown；coxal bristles dark．Fore tibia with one to two bristles among vestiture on ventral side；tibial organ comprising more than half of tibial apex，consisting of a transverse patch of bristles surrounded by a robust circular border．Mid tibia with one or two anterioventral and two to four posterioventral robust bristles among vestiture． Hind tibia with several robust bristles on anterioventral，posterior and posterioventral sides and a longitudinal row of robust bristles，which extends along two－thirds of length of tibia．Basal tarsomere of all legs with conspicuous robust
bristles. All tibial spurs equal in length. Claws with teeth. Abdomen. Pale brown, with dark bristles. Hypopygium (Fig. 56) as pale brown as abdomen. Gonocoxites wider than long; ventrally v-shaped and not fused, without a basal lobe of bristles. Gonostylus slender, apical quarter curved inwards, 3.3-4.0 times longer than wide; apically with dense setae and a group of approximately eight small spines; subapically one short whip-lash hair, which is two times as long as the apical spines. Tegmen broad and membranous, only strongly sclerotized on lateral and basal margins; wider than long, with short parameral apodemes; apically membranous with a robust hood-like structure; dorsal structures absent. Area of teeth with six to 20 conspicuous teeth. Ejaculatory apodeme short and very dark with a large base. Posterior portion of gonocoxal apodeme long and brown. Anterior portion of gonocoxal apodeme apodeme long.

Body size: 1.8-2.5 mm.
Female. Unknown.
Genetic distances. The maximum p-distance between all 13 available COI sequences is $1.83 \%$. The nearest neighbour is Psl. teo, diverging by a minimum of $4.92 \%$. The maximum p-distance between all three available 28 S sequences is $0.49 \%$ (this high p-distance results from some ambigious positions at the beginning of one sequence [SDEI-Dipt-0001511]). Some of these sequences are identical to those of Psl. frederickedwardsi, Psl. gonotegmenta, Psl. huttoni, Psl. macrotegmenta, Psl. plicitegmenta, Psl. robustotegmenta, Psl. subtilitegmenta, Psl. tonnoiri, and Psl. wernermohrigi.

Etymology. The new species is named after Dr Mathias Jaschhof, to honour his work on the Sciaroidea and to express the author's thanks for his manifold support.

Distribution. New Zealand.
Discussion. Among the species of the Psl. macrotegmenta complex this new species can easily be recognised by the hood-like structure on the tegmen, which otherwise only appears in Psl. aotearoa and Psl. zealandica. Furthermore, Psl.jaschhofi has a very elongated gonostylus, which it only shares with the otherwise unmistakable Psl. dentitegmenta. Pseudolycoriella attrita Mohrig from Papua New Guinea has very similarly shaped gonostyli and tegmen (Mohrig, 2013), but a much smaller body size and shorter flagellomeres (1.8 vs. 2.6-3.1). Additionally, the two gonostylar spines of Psl. attrita are well developed and clearly visible among the apical setae, while in Psl. jaschhofi the spines are more numerous and not so easy to distinguish from the apical setae.

## Pseudolycoriella fiordlandia sp. n.

(Fig. 55)
urn:Isid:zoobank.org:act:0CEEA4EF-C3F2-427F-86D4-B4A203964228
Material studied. Holotype male. New Zealand: South Island, Southland, Fiordland, Fiordland NP, 3 km E Milford Sound, mixed podocarp/broadleaf forest, Malaise trap, 01-21.12.2002, leg. M. \& C. Jaschhof \& U. Kallweit (NZAC). Paratypes. 50 o South Island, Southland, Otago Lakes, Fiordland NP, Eglinton River Valley, Deer Flat, Nothofagus forest, Malaise trap, 01-20.12.2002, leg. M. \& C. Jaschhof \& U. Kallweit (2x NZAC; 3x SDEI).

Description. Male. Head brown; eye bridge two to three facets wide. Scape and pedicel concolourous with flagellomeres; fourth flagellomere 2.4-2.8 times as long as wide; necks of flagellomeres differentiated; surface of flagellomeres rough with deep pits; sensilla of two different lengths; setae sparsely scattered, approximately as long as flagellomere width, more curved towards the apex; some setae on the basal flagellomeres with conspicuous bases. Prefrons and clypeus moderately bulging. Maxillary palp three-segmented; first palpomere longest, third slightly longer than second; first palpomere with three to four bristles and an inconspicuous patch of sensilla. Thorax pale brown, slightly paler than head. Posterior pronotum bare. Anterior pronotum with five to eight bristles. Episternum 1 with six to eight bristles. Mesonotum with several longer bristles among the lateral bristles and among the dorsocentral bristles. Scutellum with four robust and some shorter bristles. Katepisternum bicoloured, basal part pale brown, apical third brown; as long as high. Wing. Length $1.8-2.0 \mathrm{~mm}$; width/length ratio $0.39-0.42$. Membrane transparent and without macrotrichia, anal area well developed; all posterior veins distinct except faint stem of M, all without macrotrichia; apical quarter of $R_{5}$ with macrotrichia on the ventral and dorsal side; bM and r-m bare, $\mathrm{r}-\mathrm{m}$ exceptionally with one to two macrotrichia (in three of the paratypes, only developed on one wing each); $\mathrm{R}_{1}$ short, $0.58-0.80$ times as long as R ; M-fork longer than stem of M ; c/w ratio $0.60-0.75$; r-m/bM $0.6-0.8$ (not easy to measure due to faint stem of M). Haltere brown. Legs pale brown, except mid and hind coxae which are slightly darker; coxal bristles dark. Fore tibia with one to two spines among vestiture on ventral side; tibial organ comprising
more than half of tibial apex, consisting of a patch of bristles surrounded by a robust circular border. Mid tibia with three to four robust anterioventral and up to two posterioventral spines among the vestiture. Hind tibia with a dorsal, posterioventral and a ventral row of robust spines; the dorsal one three-quarters of the tibia length. Basal tarsomere of all legs with conspicuous robust spines. All tibial spurs equal in length. Claws with two teeth. Abdomen. Pale brown, with dark bristles. Hypopygium (Fig. 55) pale brown, like the abdomen. Gonocoxites longer than wide; ventrally v-shaped and not fused, without a basal lobe of bristles; apex of gonocoxites stretched. Gonostylus curved, bean shaped, 2.8-3.4 times longer than wide; in basal third with a robust insertion area for the adductor tendon; apically with dense setae and a group of approximately ten small spines, which are not easy to distinguish; subapically one to two long whip-lash hairs. Tegmen large and strongly sclerotized; as long as wide, with very robust parameral apodemes; margin of tegmen laterally sclerotized and with a prominent ledge on each side; sclerotized margin apically interrupted by a small membranous area; dorsal structures present. Area of teeth with approximately ten tiny teeth. Ejaculatory apodeme very dark with a large slightly paler base. Posterior portion of gonocoxal apodeme apodeme broad and brown. Anterior portion of gonocoxal apodeme apodeme medium-sized and dark.

Body size: 1.6-1.9 mm.
Female. Unknown.
Genetic distances. No genetic data available.
Etymology. The new species is named after the Fiordland National Park where the type locality and the second sampling site are located.

## Distribution. New Zealand.

Discussion. Pseudolycoriella fiordlandia has a unique tegmen with conspicuous lateral ledges and very robust parameral apodemes. In addition, the gonostyli are more curved than in the other species of this complex.

## Phylogenetic tree

According to the software Modelfinder (implemented in IQ-TREE), the best DNA substitution model for the three concatenated nuclear and mitochondrial genes is $\mathrm{GTR}+\mathrm{F}+\mathrm{R} 4$ (general time reversible model with unequal rates and unequal base frequencies (Tavaré 1986)). The phylogenetic tree based on this model is shown in Figure 61. The Log-likelihood of this tree is $-18,016.987$. The calulation strongly supports the monophyly of the Sciaridae but does not yield an adequately supported UFBootstrap value for the Pseudolycoriella cluster. The first internal branching results in two major subclades both with high UFBoot support. One major subclade contains an undescribed Psl. species from China and the New Zealand Psl. macrotegmenta complex, whose status as a monophyletic group is highly supported. The first internal branching of the Psl. macrotegmenta complex is Psl. jaschhofi and remaining species of this complex. The remaining species do not show high genetic distances. The clusters of specimens belonging to Psl. wernermohrigi and Psl. frederickedwardsi are supported by high UFBootstrap values ( $\geq 99 \%$ ). Another highly supported cluster contains a mixture of four species-Psl. gonotegmenta, Psl. plicitegmenta, Psl. subtilitegmenta and Psl. robustotegmenta. None of these four species can be discriminated by the genetic markers used.

The second major subclade of Pseudolycoriella consists of Psl. cavatica + (an undescribed Psl. species from Korea + (the remaining New Zealand representative of Pseudolycoriella + (Psl. horribilis + another undescribed Psl. species from Korea))). The clade of the remaining representatives of Pseudolycoriella shows an unsolved trichotomy of the supported Psl. jejuna complex, a supported branch of the Psl. zealandica complex and an unsupported branch of Psl. kaikoura + Psl. bispina complex. The branch of the Psl. jejuna complex and nearly all its internal nodes with only two exceptions are highly supported. The two internal nodes of the Psl.zealandica complex are supported by the UFBootstrap approximation. Nearly all internal nodes of the Psl. bispina complex are also highly supported. Only the position of Psl. teo within the Psl. bispina complex remains uncertain.

0.02 substitutions per site

FIGURE 61. Phylogenetic tree of the New Zealand Pseudolycoriella species inferred from Maximum Likelihood analysis based on three concatenated genes (COI, 16S, and 28S; total length $3,053 \mathrm{bp}$ ) applying the model GTR $+\mathrm{F}+\mathrm{R} 4$. Ultrafast bootstrap values (UFBoot; 1,000 replicates) are given if these values are greater than or equal to $95 \%$. Intraspecific UFBoot values are not specified. Pseudolycoriella species without available genetic data are also inserted and assigned to a species complex according to the morphological results.

## Discussion

The present study demonstrates a high diversity of New Zealand Pseudolycoriella species. The number of species has increased from seven to 36 . Twenty-eight species were described as new for science and one species-the widely distributed Psl. cavatica-is newly recorded for New Zealand. Two further species are only represented by female specimens and have not yet been described. Hopefully future projects will record and recognise their male counterparts by barcode matching. These undescribed species and the circumstance that it was not possible to provide new records for Psl. breviseta indicates that even more, still unknown New Zealand Pseudolycoriella species are to be expected. The existence of such a large proportion of new species is not unusual in sciarids. For instance, Savage et al. (2019) stated that 129 described Canadian sciarid species exist, compared to 2,863 BINs. Even in the comparatively well studied Western Palaearctic the use of molecular techniques has revealed hitherto unknown cryptic species complexes (Heller et al. 2016). Subsequently, the small community of sciarid taxonomists still faces a mammoth task.

Interestingly, apart from two species-the probably introduced Psl. cavatica and the autochthonous Psl. kai-koura-nearly all New Zealand Psl. species are grouped in only four different species complexes. In the present study the term "species complex" is used instead of "species group". The reason is that in sciarid taxonomy "species groups" are used for a provisional clustering of similar species mostly for determination purposes and without detailed consideration of the underlying phylogenetic relationship. Another reason for avoiding the term "species group" is that three of the discovered species complexes could be incorporated into the Psl. bruckii group without any problems. Only the Psl. macrotegmenta complex can be considered as a separate group of species, which is not covered by the currently used species group concepts for Pseudolycoriella. Among these Pseudolycoriella taxa, the applicability of molecular markers, in particular the frequently used COI gene, differs greatly. Of the New Zealand members of the Psl. bruckii group (represented by Psl. cavatica and Psl. kaikoura and the species complexes grouped around Psl. bispina, Psl.jejuna, and Psl. zealandica) the genetic distances are sufficient enough to allow an allocation of a sequence to a known species. The lowest p-distance between two specimens of different species ranges from $3.51 \%$ (between Psl. tewaipounamu and Psl. whakahara) to $12.21 \%$ between (Psl. aotearoa and Psl. zealandica). Only the intraspezific p-distance of Psl. puhihi is in this range (4.44\%). Otherwise this value lies between $0 \%$ and $3.07 \%$ in Psl. bispina. The median of the lowest interspecific p-distances is $7.14 \%$. The median of the intraspecific p-distances is $0.76 \%$. Contrary to the Psl. bruckii group, the COI p-distances between species of the Psl. macrotegmenta complex are lower and do not always allow a clear delimitation of the species. This issue is best illustrated by Psl. gonotegmenta, Psl. plicitegmenta, and Psl. subtilitegmenta, which partly show identical sequences. Among the remaining species the value of the lowest p-distance ranges between $0.61 \%$ (between Psl. gonotegmenta and Psl. robustotegmenta) to $4.92 \%$ between (Psl.jaschhofi and Psl. teo). The median is $1.84 \%$. The intraspecific p-distances range from $0 \%$ to $1.83 \%$ (Psl. jaschhofi), with a median of $0.15 \%$. Such inapplicability of COI barcoding has also been observed in other insect groups, e.g. in some sawflies (Prous et al. 2017) and in some hoverflies (Haarto \& Ståhls 2014). It is to be assumed that the Psl. macrotegmenta complex has undergone a recent radiation and that this speciation process is not yet reflected by the nuclear 28 S gene. In the mitchondrial COI gene the radiation is only partially mirrored, e.g. three species that are morphologically well separated show identical COI sequences, possibly due to incomplete lineage sorting. Introgression may also be a factor. In this context, it is noteworthy that two of these probably recently diverged species-Psl. plicitegmenta and Psl. subtilitegmen$t a$-were caught at the same locality, i.e., occur sympatrically. This implies the existence of adequate reproductive barriers. Some other sibling species pairs were caught at the same localities, e.g. Psl. jejuna/Psl. tuakana and Psl. aoteraoa/Psl. zealandica. However, in these cases at least the COI sequences support the morphological species delimitation.

A temporary result of the present study is a predominance of the genus Pseudolycoriella with 36 species compared to a total of 82 New Zealand sciarid species. This highly unnatural situation will change with further taxonomic work in the other Sciarid genera of New Zealand. It is highly likely that the genera Corynoptera Winnertz and Ctenosciara Tuomikoski also contain several undescribed species and cryptic species complexes in New Zealand.

## Acknowledgment

The author is indebted to the DGaaE (German Society of general and applied Entomology) for their generous funding and to Frank Menzel (Müncheberg, Germany) for the possibility to work with these puzzling and challenging gnats at the SDEI. The extensive collections of Catrin and Mathias Jaschhof (both Färjestaden, Sweden), and Peter A. Maddison (Katikati, New Zealand) were an elementary and indispensable resource for this study. Other collectors, who kindly caught sciarids in New Zealand, were Ian Hogg, John Hutcheson, Peter M. Johns, Uwe Kallweit, Roderick P. Macfarlane, and Andreas Stark. Werner Mohrig, who passed away on $26^{\text {th }}$ April 2019, and Duncan Sivell (formerly London, UK) are thanked for loans of type material from collections curated by them. Pekka Vilkamaa (Helsinki, Finland) also kindly provided some specimens for this study. Valerie Levesque-Beaudin (Guelph, Canada) gave some helpful information about the specimens held in the Centre for Biodiversity Genomics. Katja Kramp (Müncheberg) introduced the author into molecular lab techniques and the methodology of genetic analyses. Furthermore, she and Marko Prous (Müncheberg) discussed and answered several questions about these issues. Without the assistance of Christian Kutscher (Müncheberg) no SEM investigation would have been possible. Andrew Liston (Müncheberg) proofread the manuscript and improved the language significantly. Finally, Adam Broadley (Melbourne, Australia) and Hans-Georg Rudzinski (Schwanewede, Germany) carried out the extremely helpful task of reviewing the manuscript. The author would like to express his sincere thanks to everyone.

## References

Boevé, J.-L., Blank, S.M., Meijer, G. \& Nyman, T. (2013) Invertebrate and avian predators as drivers of chemical defensive strategies in tenthredinid sawflies. BMC evolutionary biology, 13. https://doi.org/10.1186/1471-2148-13-198
BOLD (2019a) Public data portal - BIN page. BIN BOLD:ABW3602. http://dx.doi.org/10.5883/BOLD:ABW3602
BOLD (2019b) Public data portal - BIN page. BIN BOLD:ACP1302. http://dx.doi.org/10.5883/BOLD:ACP1302
Broadley, A., Kauschke, E. \& Mohrig, W. (2016) Revision of the types of male Sciaridae (Diptera) described from Australia by F.A.A. Skuse. Zootaxa, 4193 (3), 401-450. http://doi.org/10.11646/zootaxa.4193.3.1
Coleman, C.O. (2003) "Digital inking": How to make perfect line drawings on computers. Organisms Diversity \& Evolution, 3, 1-14.
Coleman, C.O. (2009) Drawing setae the digital way. Zoosystematics and Evolution, 85 (2), 305-310. http://doi.org/10.1002/zoos. 200900008
Davies, T.H. (1988) List of Mycetophilidae and Sciaridae (Diptera) collected in Hawkes Bay. New Zealand Entomologist, 11 (1), 12-14. https://doi.org/10.1080/00779962.1988.9722529
Folmer, O., Black, M., Hoeh, W., Lutz, R. \& Vrijenhoek, R. (1994) DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. Molecular marine biology and biotechnology, 3 (5), 294-299.
Friedrich, M. \& Tautz, D. (1997) Evolution and phylogeny of the Diptera: a molecular phylogenetic analysis using 28S rDNA sequences. Systematic Biology, 46 (4), 674-698. https://doi.org/10.1093/sysbio/46.4.674
Haarto, A. \& Ståhls, G. (2014) When mtDNA COI is misleading: congruent signal of ITS2 molecular marker and morphology for North European Melanostoma Schiner, 1860 (Diptera, Syrphidae). ZooKeys, 431, 93-134. https://doi.org/10.3897/zookeys.431.7207
Hardy, D.E. (1960) Diptera: Nematocera - Brachycera. Insects of Hawaii, 10, 1-268.
Hebert, P.D.N., Penton, E.H., Burns, J.M., Janzen, D.H. \& Hallwachs, W. (2004) Ten species in one: DNA barcoding reveals cryptic species in the neotropical skipper butterfly Astraptes fulgerator. Proceedings of the National Academy of Sciences of the United States of America, 101 (41), 14812-14817. https://doi.org/10.1073/pnas. 0406166101
Heller, K. (2012) A new species of the genus Pseudolycoriella (Diptera: Sciaridae) bred from an ornamental plant. Heteropterus Revista de Entomología, 12 (2), 195-199.
Heller, K., Köhler, A., Menzel, F., Olsen, K.M. \& Gammelmo, Ø. (2016) Two formerly unrecognized species of Sciaridae (Diptera) revealed by DNA barcoding. Norwegian Journal of Entomology, 63, 96-115.
Hippa, H., Vilkamaa, P. \& Heller, K. (2010) Review of the Holarctic Corynoptera Winnertz, 1867, s. str. (Diptera, Sciaridae). Zootaxa, 2695 (1), 1-197.
https://doi.org/10.11646/zootaxa.2695.1

Hoang, D.T., Chernomor, O., von Haeseler, A., Minh, B.Q. \& Vinh, L.S. (2018) UFBoot2: Improving the Ultrafast Bootstrap Approximation. Molecular Biology and Evolution, 35 (2), 518-522.
https://doi.org/10.1093/molbev/msx281
Hutton, F.W. (1881) Catalogues of the New Zealand Diptera, Orthoptera, Hymenoptera; with descriptions of the species. Wellington: G. Didsbury, Government Printer.
Jaschhof, M. \& Jaschhof, C. (2003a) Wood midges of New Zealand (Cecidomyiidae, Lestremiinae). Part I: Introductory notes and tribes Lestremiini, Strobliellini, Campylomyzini and Pteridomyiini Jaschhof trib. nov. Studia dipterologica, 10 (1), 97-132.
Jaschhof, M. \& Jaschhof, C. (2003b) Wood midges of New Zealand (Cecidomyiidae, Lestremiinae). Part II: Tribes Micromyini and Aprionini. Studia dipterologica, 10 (2), 423-440.
Jaschhof, M. \& Jaschhof, C. (2004) Wood midges of New Zealand (Cecidomyiidae, Lestremiinae). Part III: Tribe Peromyiini and remarks on the composition, origin and relationships of the fauna as a whole. Studia dipterologica, 11 (1), 75-127.
Jaschhof, M. \& Jaschhof, C. (2009) The Wood Midges (Diptera: Cecidomyiidae: Lestremiinae) of Fennoscandia and Denmark. Studia dipterologica Supplement, 18, Ampyx-Verlag, Halle (Saale), 333 pp.
Kalyaanamoorthy, S., Minh, B.Q., Wong, T.K.F., von Haeseler, A. \& Jermiin, L.S. (2017) ModelFinder: fast model selection for accurate phylogenetic estimates. Nature methods, 14 (6), 587-589. https://doi.org/10.1038/nmeth. 4285
Katoh, K., Misawa, K., Kuma, K. \& Miyata, T. (2002) MAFFT: a novel method for rapid multiple sequence alignment based on fast Fourier transform. Nucleic Acids Research, 30 (14), 3059-3066. https://doi.org/10.1093/nar/gkf436
Katoh, K., Rozewicki, J. \& Yamada, K.D. (2017) MAFFT online service: multiple sequence alignment, interactive sequence choice and visualization. Briefings in Bioinformatics, bbx108, 1-7. https://doi.org/10.1093/bib/bbx108
Katoh, K. \& Standley, D.M. (2013) MAFFT Multiple Sequence Alignment Software Version 7: Improvements in Performance and Usability. Molecular Biology and Evolution, 30 (4), 772-780. https://doi.org/10.1093/molbev/mst010
Köhler, A. \& Menzel, F. (2013) New records of Black Fungus Gnats (Diptera: Sciaridae) from New Caledonia, with the description of two new Bradysia species and an updated checklist. Zootaxa, 3718 (1), 63-72. https://doi.org/10.11646/zootaxa.3718.1.5
Köhler, A. \& Mohrig, W. (2016) Additions to the New Zealand fauna of black fungus gnats (Diptera: Sciaridae), with descriptions of six new species. New Zealand Entomologist, 39 (2), 91-109. https://doi.org/10.1080/00779962.2016.1153233
Kück, P. \& Longo, G.C. (2014) FASconCAT-G: extensive functions for multiple sequence alignment preparations concerning phylogenetic studies. Frontiers in Zoology, 11 (1), 81. https://doi.org/10.1186/s12983-014-0081-x
Kumar, S., Stecher, G. \& Tamura, K. (2016) MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. Molecular Biology and Evolution, 33 (7), 1870-1874.
https://doi.org/10.1093/molbev/msw054
Leblanc, L., Rubinoff, D. \& Vargas, R.I. (2009) Attraction of Nontarget Species to Fruit Fly (Diptera: Tephritidae) Male Lures and Decaying Fruit Flies in Traps in Hawaii. Environmental Entomology, 38 (5), 1447-1461. https://doi.org/10.1603/022.038.0513
Macfarlane, R.P., Maddison, P.A., Andrew, I.G., Berry, J.A., Johns, P.M., Hoare, R.J.B., Larivière, M.-C., Greenslade, P., Henderson, R.C., Smithers, C.N., Palma, R.L., Ward, J.B., Pilgrim, R.L.C., Towns, D.R., Mclellan, I., Teulon, D.A.J., Hitchings, T.R., Eastop, V.F., Martin, N.A., Fletcher, M.J., Stufkens, M.A.W., Dale, P.J. Burckhardt, D. Buckley, T.R. \& Trewick, S.A. (2010) 9. Phylum Arthropoda, Subphylum Hexapoda, Protura, springtails, Diplura, and insects. In: D.P. Gordon (Ed), New Zealand Inventory of Biodiversity. Vol. 2. Kingdom Animalia: Chaetognatha, Ecdysozoa, Ichnofossils. Canterbury Univ. Press, Christchurch, pp. 233-467.
Menzel, F. \& Heller, K. (2007) Bemerkungen zur Nomenklatur der Sciariden (Diptera, Bibionomorpha: Sciaridae). Studia dipterologica, 13 (2), 209-229. [in German]
Menzel, F. \& Mohrig, W. (1998) Beiträge zur Taxonomie und Faunistik der paläarktischen Trauermücken (Diptera, Sciaridae). Teil VI - Neue Ergebnisse aus Typenuntersuchungen und die daraus resultierenden taxonomisch-nomenklatorischen Konsequenzen. Studia dipterologica, 5 (2), 351-378. [in German]
Menzel, F. \& Mohrig, W. (2000) Revision der paläarktischen Trauermücken (Diptera: Sciaridae). Studia dipterologica Supplement 6. Ampyx-Verlag, Halle (Saale), 761 pp. [in German]
Menzel, F. \& Smith, J.E. (2009) Family Sciaridae. In: Gerlach, J. (Ed.), The Diptera of the Seychelles Islands. Pensoft Publisher, Sofia \& Moscow, pp. 19-45.
Menzel, F. \& Smith, J.E. (2017) 21 Sciaridae (Black Fungus Gnats). In: Kirk-Spriggs, A.H. \& Sinclair, B.J. (Eds.), Manual of Afrotropical Diptera. Vol. 2. Nematocerous Diptera and lower Brachycera. Suricata 5. SANBI Graphics \& Editing, Pretoria, pp. 557-580.
Menzel, F., Vilkamaa, P. \& Smith, J.E. (2013) Overview of the Black Fungus Gnats from the Tristan da Cunha archipelago, including a redescription of Hyperlasion viridiventris (Frey) (Diptera: Sciaroidea: Sciaridae). Contributions to Entomology,

63 (2), 283-296.
https://doi.org/10.21248/contrib.entomol.63.2.283-296
Miller, D. (1950) Catalogue of the Diptera of the New Zealand Sub-region. New Zealand Department of Scientific and Industrial Research Bulletin, 100, 1-194.
Minh, B.Q., Nguyen, M.A.T. \& von Haeseler, A. (2013) Ultrafast Approximation for Phylogenetic Bootstrap. Molecular Biology and Evolution, 30 (5), 1188-1195.
https://doi.org/10.1093/molbev/mst024
Mohrig, W. (2013) Die Trauermücken (Diptera: Sciaridae) von Papua-Neuguinea. Teil III - Gattungen Ctenosciara und Pseudolycoriella. Studia dipterologica, 20 (1), 123-168. [in German]
Mohrig, W. \& Blasco-Zumeta, J. (1996) The sciarid fauna (Diptera, Sciaridae) of a Juniperus thurifera L. forest of the Monegros region (Zaragoza, Spain) with description of ten new species. Miscellanea Zoologica, 18, 99-116.
Mohrig, W., Heller, K., Hippa, H., Vilkamaa, P. \& Menzel, F. (2013) Revision of the Black Fungus Gnats (Diptera: Sciaridae) of North America. Studia dipterologica, 19 (1-2), 141-286.
Mohrig, W. \& Jaschhof, M. (1999) Sciarid flies (Diptera, Sciaridae) of New Zealand. Studia dipterologica Supplement 7. Ampyx-Verlag, Halle (Saale), 101 pp.
Mohrig, W. \& Kauschke, E. (2019) New Black Fungus Gnats (Diptera, Sciaridae) of North America. Part V. Genera Pseudolycoriella Menzel \& Mohrig and Phytosciara Frey. Zootaxa, 4543 (2), 261-283. https://doi.org/10.11646/zootaxa.4543.2.5
Mohrig, W., Kauschke, E. \& Broadley, A. (2016) Pseudolycoriella skusei sp. nov. (Diptera: Sciaridae), a new dark-winged fungus gnat from Norfolk Island and Australia. Zootaxa, 4097 (1), 139-142. http://dx.doi.org/10.11646/zootaxa.4097.1.11
Mohrig, W., Kauschke, E. \& Broadley, A. (2018) New black fungus gnats (Diptera: Sciaridae) from Eastern Australia. Zootaxa, 4450 (2), 203-241. https://doi.org/10.11646/zootaxa.4450.2.3
Mohrig, W., Kauschke, E. \& Broadley, A. (2019) Revision of black fungus gnat species (Diptera, Sciaridae) described from the Hawaiian Islands by D.E. Hardy and W.A. Steffan, and a contribution to the knowledge of the sciarid fauna of the Galápagos Islands. Zootaxa, 4590 (4), 401-439. https://doi.org/10.11646/zootaxa.4590.4.1
Mohrig, W., Röschmann, F. \& Rulik, B. (2004) The fauna of scarid flies from the Dominican Republic (Diptera, Sciaridae). Beiträge zur Entomologie, 54 (2), 267-331. https://doi.org/10.21248/contrib.entomol.54.2.267-331
Natural History Museum (2014) Dataset: Collection specimens. Resource: Specimens. Natural History Museum Data Portal (data.nhm.ac.uk). https://doi.org/10.5519/0002965
New Zealand Arthropod Collection (2019) Specimen data. Accessed through Systematics Collection Data. Available from: http://scd.landcareresearch.co.nz (accessed 16 January 2019)
Nguyen, L.-T., Schmidt, H.A., von Haeseler, A. \& Minh, B.Q. (2015) IQ-TREE: A Fast and Effective Stochastic Algorithm for Estimating Maximum-Likelihood Phylogenies. Molecular Biology and Evolution, 32 (1), 268-274. https://doi.org/10.1093/molbev/msu300
Palumbi, S., Martin, A., Romano, S., McMillan, W., Stice, L. \& Grabowski, G. (1991) Simple fool's guide to PCR. University of Hawaii, Honolulu, 45 pp .
Prous, M., Heidemaa, M. \& Soon, V. (2011) Empria longicornis species group: taxonomic revision with notes on phylogeny and ecology (Hymenoptera, Tenthredinidae). Zootaxa, 2756 (1), 1-39. https://doi.org/10.11646/zootaxa.2756.1.1
Prous, M., Kramp, K., Vikberg, V. \& Liston, A. (2017) North-Western Palaearctic species of Pristiphora (Hymenoptera, Tenthredinidae). Journal of Hymenoptera Research, 59, 1-190. https://doi.org/10.3897/jhr.59. 12656
Rudzinski, H.-G. (2000) Neue Trauermücken aus der tschechischen und slowakischen Republik (Diptera: Sciaridae). Mitteilungen des Internationalen Entomologischen Vereins, 25 (3/4), 167-184.
Rudzinski, H.-G., Baumjohann, K. \& Wolff, M. (2016) Pseudolycoriella martita sp. nov.: The first species of the genus Pseudolycoriella Menzel \& Mohrig, 1998 from Colombia. Mitteilungen des Internationalen Entomologischen Vereins, 41 (1/2), 5-9.
Savage, J., Borkent, A., Brodo, F., Cumming, J.M., Curler, G. \& Currie, D.C. (2019) Diptera of Canada. ZooKeys, 819, 397450. https://doi.org/10.3897/zookeys.819.27625
Ševčík, J., Kaspřák, D., Mantič, M., Ševčíková, T. \& Tóthová, A. (2014) Molecular phylogeny of the fungus gnat family Diadocidiidae and its position within the infraorder Bibionomorpha (Diptera). Zoologica Scripta, 43 (4), 370-378. https://doi.org/10.1111/zsc. 12059
Ševčík, J., Kaspřák, D. \& Tóthová, A. (2013) Molecular phylogeny of fungus gnats (Diptera: Mycetophilidae) revisited: Position of Manotinae, Metanepsiini, and other enigmatic taxa as inferred from multigene analysis. Systematic Entomology, 38 (4), 654-660.

Shin, S., Jung, S., Menzel, F., Heller, K., Lee, H. \& Lee, S. (2013) Molecular phylogeny of black fungus gnats (Diptera: Sciaroidea: Sciaridae) and the evolution of larval habitats. Molecular Phylogenetics and Evolution, 66 (3), 833-846. https://doi.org/10.1016/j.ympev.2012.11.008
Simon, C., Frati, F., Beckenbach, A., Crespi, B., Liu, H. \& Flook, P. (1994) Evolution, weighting, and phylogenetic utility of mitochondrial gene sequences and a compilation of conserved polymerase chain reaction primers. Annals of the Entomological Society of America, 87 (6), 651-701. https://doi.org/10.1093/aesa/87.6.651
Skuse, F.A.A. (1888) Diptera of Australia. Part II. The Sciaridae. The Proceedings of the Linnean Society of New South Wales (Second series), 3, 657-726.
Steffan, W.A. (1973) Notes on Hawaiian Sciaridae (Diptera) and Descriptions of two new species. Pacific Insects, 15 (3-4), 353-361.
Steffan, W.A. (1989) 11. Family Sciaridae. In: Evenhuis, N.L. (Ed), Catalog of the Diptera of the Australasian and Oceanian Regions. Bishop Museum Press \& E. J. Brill, Honolulu, pp. 146-151.
Tavaré, S. (1986) Some probabilistic and statistical problems in the analysis of DNA sequences. Lectures on mathematics in the life sciences, 17 (2), 57-86.
Tonnoir, A.L. \& Edwards, F.W. (1927) New Zealand fungus gnats (Diptera, Mycetophilidae). Transactions and Proceedings of the New Zealand Institute, 57, 747-878.
Vilkamaa, P., Hippa, H. \& Mohrig, W. (2012) The genus Pseudolycoriella Menzel \& Mohrig (Diptera, Sciaridae) in New Caledonia, with the description of thirteen new species. Zootaxa, 3207 (1), 1-21. https://doi.org/10.11646/zootaxa.3207.1
Whiting, M.F., Carpenter, J.C., Wheeler, Q.D. \& Wheeler, W.C. (1997) The Strepsiptera problem: phylogeny of the holometabolous insect orders inferred from 18S and 28S ribosomal DNA sequences and morphology. Systematic Biology, 46 (1), 1-68. https://doi.org/10.1093/sysbio/46.1.1
Wisely, B. (1959) A Contribution to the Life Histories of Two Fungus Gnats, Scythropochroa nitida Edw., and Sciara annulata Mg., (Diptera, Mycetophilidae, Sciarinae). Transactions of the Royal Society of New Zealand, 86 (1), 59-64.

