

Descriptions of genus *Downsiomyia* Vargas (Diptera: Culicidae: Aedini) and its type species *Do. nivea* (Ludlow)

JOHN F. REINERT¹ & RALPH E. HARBACH²

¹Center for Medical, Agricultural and Veterinary Entomology (CMAVE), United States Department of Agriculture, Agricultural Research Service, 1600/1700 S.W. 23rd Drive, Gainesville, Florida 32608-1067 USA, and collaborator; Walter Reed Biosystematics Unit (WRBU), National Museum of Natural History, Smithsonian Institution, Washington, DC. E-mail: jreinert@gainesville.usda.ufl.edu

²Department of Entomology, The Natural History Museum (NHM), Cromwell Road, London, SW7 5BD, UK. E-mail: r.harbach@nhm.ac.uk

Abstract

Genus *Downsiomyia* and its type species, *Do. nivea* (Ludlow), are described in detail. The 30 species included in the genus are listed. An extensive list of previous literature pertaining to the genus is provided.

Key words: *Aedes*, *Downsiomyia*, *Finlaya*, Albonivea Group, Nivea Group, *Do. nivea*, mosquitoes

Introduction

Recently, Reinert *et al.* (2004), as a result of cladistic analysis of the tribe Aedini, reinstated *Downsiomyia* Vargas to generic rank for species previously placed in the Niveus Group of *Finlaya* Theobald. These authors proposed extensive changes to the generic classification of Aedini based on the analyses of morphological data from eggs, fourth-instar larvae, pupae, and adults of all previously recognized genera, subgenera, and major groups of the tribe. The very brief original description (in Spanish) of *Downsiomyia* by Vargas (1950) is inadequate for distinguishing generic-level taxa of Culicidae, therefore the genus and its type species, *Do. nivea* (Ludlow), are described in detail below. The genus includes 30 species (listed herein). In accordance with provisions of the *International Code of Zoological Nomenclature* (International Commission on Zoological Nomenclature 1999), the terminations of some species names were changed to agree in gender with *Downsiomyia*.

Morphological terminology used in the descriptions follows Harbach & Knight (1980, 1982) and Reinert (1990, 1999b, 2000b). The recommended two-letter abbreviation for *Downsiomyia* is *Do*. A “previous usage” section is provided for tracking the published generic-level history of species included in *Downsiomyia*.

Genus *Downsiomyia* Vargas, 1950

Type species: *Stegomyia nivea* Ludlow, 1903

Previous usage

Stegomyia Theobald, of Ludlow (1903), in part of Theobald (1903, 1905a, 1907, 1908, 1910), Blanchard (1905), Ludlow (1905), Leicester (1908), Stone (1957b).

Scutomyia Theobald of Ludlow (1911), in part of Theobald (1905b, 1907, 1910), Banks (1906), Brunetti (1907, 1912).

Ochlerotatus Lynch Arribalzaga, of Shriram *et al.* (2005), in part of Edwards (1913), Moulton (1914), Brunetti (1920), Senior-White (1923).

Aedes (*Ochlerotatus*) Group *Finlaya*, in part of Edwards (1917).

Finlaya Theobald, in part of Barraud (1923, 1924), Senior-White (1927).

Armigeres of Edwards (1926).

Aedes (*Finlaya*) of Knight & Chamberlain (1948), Peters & Dewar (1956), Stone (1957a), Knight (1978a), Luh & Li (1980), Huang & Rueda (1998), in part of Dyar (1920), Edwards (1921, 1922a, 1922b, 1928, 1929), Brug & Haga (1923), Haga (1924), Dyar & Shannon (1925), Brug (1926, 1931, 1934, 1939), Borel (1930), Brug & Edwards (1931), Barraud (1934), Li & Wu (1935), Bonne-Wepster & Brug (1937, 1939), Causey (1937), Hu (1937), Feng (1938a, 1938b, 1958), Wu (1940), Bohart (1945, 1946), Hsiao (1945), Bohart & Ingram (1946), Brug & Bonne-Wepster (1947), LaCasse (1948), LaCasse & Yamaguti (1948, 1950), Sasa (1948), Chow (1949a, 1949b, 1950), Carter (1950), Knight & Hull (1951), Monchadskii (1951), Bonne-Wepster (1954a, b), Chow *et al.* (1954), Horsfall (1955), Stone & Knight (1956), Hsieh & Liao (1956), Hara (1957), Macdonald (1957), Chu (1957), Wattal *et al.* (1958), Thurman (1959, 1963), Stone *et al.* (1959), Stone (1961, 1963, 1967, 1970), Omori (1962), Lien (1962, 1968), Kurihara (1963, 1978, 1981, 1999), Delfinado *et al.* (1963), Loy & Rowland (1963), Rozeboom & Cabrera (1964), Mackie (1964), Joshi *et al.* (1965), Macdonald *et al.* (1965), Army Mosquito Project (1965), Scanlon & Esah (1965), Scanlon & Peyton (1965), Stone *et al.* (1966), Gould *et al.* (1968), Ramachandran *et al.* (1970), Aslamkhan (1971), Tanaka (1971, 2002), Basio (1971), Grothaus *et al.* (1971), Qutubuddin (1972), Rahman *et al.* (1973), Stone & Delfinado (1973), Ramalingam & Pillai (1973), Ramalingam (1974), Matsuo *et al.* (1974), Gutsevich *et al.* (1974), Baisas (1974), Lien *et al.* (1975), Bhat (1975), Tanaka *et al.* (1975), Sasa *et al.* (1977), Lien *et al.* (1977), Knight & Stone (1977), Knight (1978b), Lee *et al.* (1980, 1982), Gutsevich & Dubitsky (1981), Jayasekera & Chelliah (1981),

O'Connor & Sopa (1981), Ma (1982), Chau (1982), Lu & Li (1982), Gould *et al.* (1982), Cai (1984), Ward (1984, 1992), Miyagi *et al.* (1985), Tsukamoto & Horio (1985), Lee & Egan (1985), Toma & Miyagi (1986), Apiwathnasorn (1986), Rudnick *et al.* (1986), Malhotra *et al.* (1987), Nagpal & Sharma (1987), Lee (1987), Tsukamoto *et al.* (1987), Lee & Zorka (1987), Lu & Su (1987), Ahmed (1988), Lu *et al.* (1988), Debenham & Hicks (1989), Evenhuis & Gon (1989), Townsend *et al.* (1990), Darsie & Pradhan (1990), Harrison *et al.* (1991), Darsie *et al.* (1992, 1993), Malhotra & Mahanta (1994), Stojanovich & Scott (1995, 1996a, 1996b), Reinert (1999a), Peyton *et al.* (1999), Tanaka (2003), Kaur (2003), Kurihara *et al.* (2004), Rajavel *et al.* (2005b).

Aedes (Ochlerotatus) Globus Finlaya, in part of Martini (1931).

Aedes (Finlaya) Group H, in part of Edwards (1932), Bohart (1957).

Aedes Meigen, in part of Riley (1932), Yamada (1932), Hsiao & Bohart (1946), Sasa *et al.* (1947), Barnett & Toshioka (1951), Osima (1952), Ori & Shimogama (1953), Chu (1957), Barnett (1962), Cabrera & Rozeboom (1964), Sakakibara (1965), Stojanovich & Scott (1965, 1966), Lee & Lien (1970), Basio *et al.* (1973), Ree *et al.* (1973), Harinasuta *et al.* (1974), Wada *et al.* (1976), Pae *et al.* (1976), Sarkar *et al.* (1981), Malhotra *et al.* (1982), Gandahusada *et al.* (1984), Lee *et al.* (1984), Xu (1984), Khamboonruang *et al.* (1987), Hawley (1988), Darsie *et al.* (1991), Service (1993), Gerberg *et al.* (1994), Mogi (1996), Strickman *et al.* (2000), Ruang-Areerate *et al.* (2003).

Aedes (Finlaya) niveus complex of Traub & Macdonald (1963).

Aedes (Finlaya) Group H (Genuiculatus-Group), Subgroup I, Niveus of Knight & Marks (1952), Colless (1958, 1959), Macdonald & Traub (1960).

Aedes (Finlaya) niveus group of Wharton (1962), Traub & Macdonald (1963), Rao & Rajagopalan (1957), Colless (1957), Harinasuta *et al.* (1970), White (1979), Lu (1981), Rajput & Singh (1987).

Aedes (Finlaya) niveus subgroup of Sasa & Kakahashi (1952), Knight (1946, 1969), Nakato & Matuo (1960), van Peenen *et al.* (1975), Furumizo & Rudnick (1979), Amerasinghe (1982), Knight & Harrison (1988), Kulasekera *et al.* (1990), Rattarithikul & Panthusiri (1994), Tewari & Hiriyani (1995), Rueda (2004).

Aedes (Finlaya) nipponicus group of Tanaka *et al.* (1979).

Aedes (Finlaya) niveus group, *niveus* subgroup of Lu & Ji (1997).

Ochlerotatus (Finlaya), in part of Reinert (2000a), Zagaria & Savioli (2002), Rajavel *et al.* (2005a), Rattarithikul *et al.* (2005).

Ochlerotatus (Finlaya) Niveus Assemblage, of Reinert (2002).

Downsiomyia Vargas, of Reinert *et al.* (2004).

Description

FEMALES. *Head*: Vertex with broad, decumbent scales; occiput with numerous erect, forked scales; ocular line narrow, with pale scales; eyes above antennal pedicels contiguous or separated by 1 eye facet or less; interocular space small, with few pale

scales, normally 2 dark, interocular setae; antennal pedicel with few small scales and short, dark setae mesally; maxillary palpus and proboscis dark-scaled. *Thorax*: Scutum with dark integument, covered with narrow, dark scales except bare, median prescutellar space; narrow pale (normally silvery or white) scales on anterior promontory, antedorsocentral area, scutal fossa (normally entire area, rarely pale scales extending posteriorly onto antealar and anterior margin of supraalar area, e.g. *Do. nipponica* (LaCasse & Yamaguti)), anterior 0.30–0.70 of acrostichal and dorsocentral areas (some species, e.g. *Do. harinasutai* (Knight), *Do. litorea* (Colless), *Do. nipponica* and *Do. nivea* (Ludlow)); prescutellar space mesal to setae rarely with pale scales (e.g. *Do. mohani* (Knight), *Do. nipponica*, *Do. saperoi* (Knight) and *Do. watteni* (Lien)), and normally antealar area, other areas dark-scaled; acrostichal (anterior and posterior) and dorsocentral (anterior and posterior) areas without setae; scutellum with broad scales (normally dark) and several setae on all lobes; mesopostnotum bare; paratergite wide, bare; anteprepronota widely separated, with broad, pale scales, several setae; postpronotum with few broad scales normally restricted to posterodorsal area, few posterior setae; prespiracular setae absent; postspiracular area without scales, with several setae; hypostigmal and subspiracular areas bare; upper proepisternum with broad scales, several setae; lower proepisternum bare; mesokatepisternum with upper and lower posterior patches of broad, pale scales, few upper and posterior setae; prealar area with patch of broad, pale scales on lower area extending dorsally onto lower part of upper area (except absent in few species, e.g. *Do. ganapathi* (Knight), *Do. pseudonivea* (Theobald), *Do. pexa* (Colless) and *Do. vana* (Colless)); mesepimeron with 1 moderately large patch on upper area and extending onto middle, several upper setae, lower setae absent; metameron bare. *Wing*: Entirely dark-scaled; remigium with 1–3 setae distally on dorsal surface; upper calypter with several setae on margin; alula with row of narrow scales on margin; vein R_2 longer than R_{2+3} ; anal vein terminating distal to juncture of CuA and mCu. *Legs*: Hindfemur dark-scaled distally, with broad, pale-scaled areas on proximal parts of anterior and posterior surfaces; tibiae and tarsi dark-scaled; fore- and midtarsus each with 1 tooth, hindtarsus simple. *Abdomen*: Terga with basolateral, pale-scaled patches; segments VII and VIII laterally compressed.

Female genitalia. *Tergum VIII*: Proximal 0.20–0.40 retracted into segment VII; moderately to heavily pigmented, base broad and moderately concave; apex broadly rounded; numerous broad scales covering distal 0.70–0.85; apical margin with number of long, stout setae; several short setae on distal 0.35–0.69; VIII-Te index 0.50–0.76 (0.84 in *Do. nipponica*); length 0.26–0.36 mm; width 0.38–0.56 mm. *Sternum VIII*: Moderately to heavily pigmented; base straight; apex with minute (0.01–0.06 of VIII-S length), median emargination separating pair of broad, flattened lobes or with pair of broad, flattened lobes with apicolateral areas angled posterolaterally; broad scales forming moderately large patches on median, lateral areas on distal 0.76–0.92; numerous short setae on distal 0.84–0.97; apical margin with numerous short, lanceolate setae with apices curved; seta

2-S inserted posterior to 1-S; intersegmental membrane between segments VII and VIII short; VIII-S index 0.81–0.93 (0.74 in *Do. inermis* (Colless)); length 0.32–0.46 mm; width 0.38–0.50 mm. *Tergum IX*: Heavily pigmented; comprised of 2 narrow, finger-like, lateral lobes connected by very narrow, basal strip; 1–5 (usually 2–4) moderately long, stout setae apically on each lobe; IX-Te index 0.60–1.07; length 0.09–0.14 mm; width 0.10–0.16 mm. *Insula*: Covered with short spicules; lightly to moderately pigmented; short; wide; liplike; with 3–8 moderately long setae. *Lower vaginal lip*: Moderately pigmented; narrow, lower vaginal sclerite absent. *Upper vaginal lip*: Heavily pigmented; narrow; median caudal area flattened; upper vaginal sclerite very small, comprised of narrow, heavily pigmented strip along basolateral area of lip. *Spermathecal eminence*: Membranous; comprised of few wrinkled, circular folds. *Postgenital lobe*: Spicules along lateral margins longer, stout and many with basal denticles; short; moderately broad; apex flattened or broadly rounded but usually with minute (0.02–0.08 of dorsal length), median emargination; 12–39 total setae; ventral PGL index 1.42–2.00; ventral length 0.11–0.15 mm (0.17 mm in *Do. pexa*). *Cercus*: Short; broad; apical margin broad and oblique with 5–7 (usually 6) stout setae, setae long on mesal area and tapering in length to moderately long on lateral area, several short, lanceolate setae with apices curved; dorsal surface with scales absent, 2,3 long and few short setae on distal 0.37–0.65; cercus index 1.45–1.89; length 0.14–0.18 mm; width 0.08–0.11 mm. *Spermathecal capsules*: Nivea Group with one large capsule, Albonivea Group with one large and 2 medium capsules; numerous small spermathecal capsule pores near orifice.

MALES. *Head*: Antenna about 0.75 length of proboscis, flagellar whorls with numerous long setae directed dorsally and ventrally, distal 2 flagellomeres elongate; maxillary palpus dark-scaled, slightly shorter to approximately equal to proboscis length, palpomere 1 small, palpomere 2 long, narrow, palpomere 3 long, narrow, slightly upturned, with few to several long setae apically on ventral surface, palpomere 4 short, thin, with several to numerous moderately long setae on ventrolateral margins, setae projecting anteroventrally, few moderately long setae dorsoapically, palpomere 5 short, with several short to moderately long setae mainly on ventral and apical areas, palpomeres 4,5 normally slightly down-turned; proboscis longer than forefemur. *Thorax*: Anterior area of scutum with silvery scales more extensive than in female. *Legs*: Fore- and midtarsi with ungues unequal, each with 1 tooth, hindtarsus with ungues equal, both simple.

Male genitalia. *Tergum IX*: Relatively short, bearing 2 small lobes caudally, each with few to several stout setae. *Gonocoxite*: Somewhat triangular in outline, proximal part relatively broad, distal part narrow, ventral surface with several to numerous long, broad, fusiform scales on mesal area (except *Do. mikrokopion* (Knight & Harrison)); dorsal surface with numerous short, slender setae on mesal area, and with narrow, thumblike lobe on basomesal area projecting caudoventrally and bearing few to several short setae; dorsal and lateral surfaces with numerous relatively long, dark, broad, spatulate scales.

Gonostylus: Narrow, relatively short, with long, narrow gonostylar claw attached apically. *Proctiger*: Paraproct heavily pigmented and terminating in short, curved, beaklike point; few short, cercal setae. *Phallosome*: Aedeagus simple, scooplike, apex concave (except *Do. axitiosa* (Kulasekera, Knight & Harbach), see Kulasekera *et al.* 1990) with one or usually 2 or more small teeth on each side laterally; paramere narrow, shorter than aedeagus; basal piece short and broad. *Claspette*: With moderately long columnar stem bearing 1–3 short, thin setae, with narrow, basolateral extension connected to ventral area of basomesal, thumblike lobe of gonocoxite; claspette filament long, somewhat leaflike, longitudinally striated, expanded on middle part and narrow distally. *Sternum IX*: With few, normally moderately long setae caudomesally.

PUPAE. *Cephalothorax*: Seta 1-CT long, moderately stout to stout, normally single (occasionally 2-branched); 2,3-CT short, slender; 5-CT longer than 4-CT, 5-CT often long, moderately stout to stout, single; 11-CT stout, single (distal part rarely forked). *Trumpet*: With basal tracheoid area weakly developed. *Abdomen*: Seta 3-I long, single, longer than 6,7-I; 6-I single (rarely 2-branched), longer than 7-I; 7-I normally 2-4 branched (rarely single); 9-III-VI short; 6-VII short, slender, with 2–7 branches, inserted posterior and slightly mesal to 9-VII; 9-VII moderately long, stout, aciculate, normally with 2,3 branches (rarely single on 1 side); 9-VIII long, stout, multiple-branched, aciculate, inserted on posterolateral corner of segment. *Paddle*: Without fringe of long spicules; midrib well developed, reaching or nearly reaching apex; seta 1-Pa often single (with 2,3 branches in some species, e.g. *Do. novonivea*).

FOURTH-INSTAR LARVAE. *Head*: Seta 1-C single, distal part attenuate (*Do. axitiosa* split distally, see Kulasekera *et al.* 1990); 4-7-C well developed, multiple-branched, aciculate; 4-C slightly shorter than 5-7-C, inserted mesal to 6-C; 5-7-C approximately equal in length; 5-C inserted posterior to 4,6,7-C; 13-C single (single or 2-branched in *Do. mikrokopion*, see Knight & Harrison 1987), long, inserted lateral to 12-C; 18-C present; 19-C absent; antenna with spicules, 1-A multiple-branched, aciculate; 6-Mx branched; labiogula width greater than length; ventromedian cervical sclerite present. *Thorax*: Setae 1-3-P normally inserted on common setal support plate; 8-P branched, shorter than 4-P; 5-M longer than 7-M; 6-T single. *Abdomen*: Seta 12-I present; 6-I-VI long, 6-II,III branched, 6-II shorter than 6-III; 7-I long, stout, single or 2-branched; 7-II short, multiple-branched; 8-II branched; 2-VI at same level or mesal to 1-VI (2-VI slightly lateral to 1-VI in *Do. litorea*); 1-VII long; 2-VII inserted near 1-VII; 3-VII branched; 4,10,12-VII, 2,4-VIII single; comb scales in single row (except small patch in *Do. axitiosa*); segment X with saddle incomplete ventrally, normally with several moderately long to long, stout spicules on posterior margin dorsal to insertion of 1-X, acus absent, 1-X inserted on saddle, 2-X normally with few branches, much shorter than 3-X, 3-X long, single, ventral brush with several setae, posterior setae normally with several long

branches arising from short stem, inserted on grid normally with transverse and lateral bars (lateral bar weakly developed or absent in some species, e.g. *Do. harinasutai* and *Do. mohani*), 2,3 shorter precratal setae. *Siphon*: Pecten comprised of numerous, evenly spaced spines, inserted on approximately proximal 0.50 of siphon, acus present, seta 1-S branched, aciculate, inserted distal to pecten, 6,8-S short, 9-S short, slightly curved.

EGGS. Matsuo *et al.* (1974) provided photographs and the following description of the egg of *Do. albolateralis* (Theobald). *Size*: Length 470–510 μ ($495 \pm 2 \mu$); width 120–170 μ ($151 \pm 3 \mu$). *Outer chorion*: Reticulation composed of pattern of quadrilateral, pentagonal, or sometimes hexagonal cells (at 200x magnification), ridge of reticulation high, hence cells appearing concave, many small papillae in each cell, irregular in shape, occasionally confluent with each other and ridge (at 1,000x magnification).

Species included in Downsiomyia

Specimens examined in parenthesis, F = female, Fg = female genitalia, M = male, Mg = male genitalia, P = pupa, and L = fourth-instar larva. Nivea Group: *Do. albolateralis* (F, Fg, M, Mg, P, L), *Do. axitiosa*, *Do. dorseyi* (Knight) (F, Fg, M, Mg, L), *Do. ganapathi* (F, Fg, M, Mg, P, L), *Do. harinasuti* (F, Fg, M, Mg, P, L), *Do. idjenensis* (Brug), *Do. inermis* (F, Fg, M, Mg, P, L), *Do. lactea* (Knight) (F, L), *Do. laoagensis* (Knight) (F, Fg, M, P, L), *Do. leonis* (Colless) (F, Fg, M, Mg, P, L), *Do. litorea* (F, Fg, M, Mg, P, L), *Do. mikrokopion* (F, Fg, M), *Do. mjoebergi* (Edwards) (F, Fg), *Do. mohani* (F, Fg, M, P, L), *Do. nipponica* (F, Fg, M, Mg, P, L), *Do. nippononivea* (Sasa & Nakahashi) (F, M), *Do. nishikawai* (Tanaka, Mizusawa & Saugstad) (F, M, P, L), *Do. niveoides* (Barraud) (F, Fg, M, Mg, P, L), *Do. nivea* (F, Fg, M, Mg, P, L), *Do. novonivea* (Barraud) (F, Fg, M, Mg, P, L), *Do. omorii* (Lien) (F, M, Mg, P, L), *Do. pexa* (F, Fg, M, Mg, P, L), *Do. pseudonivea* (F, Fg, M, Mg, P, L), *Do. shehzadae* (Qutubuddin), *Do. sinensis* (Chow) (M, Mg), *Do. subnivea* (Edwards) (F, Fg, M, Mg, P, L), *Do. vana* (F, Fg, M, Mg, P, L), *Do. watteni* (F, M, Mg, P, L), and Albonivea Group: *Do. albonivea* (Barraud) (F, Fg, M, Mg) and *Do. saperoi* (F, Fg, M, Mg, P, L).

Bionomics

Immatures of *Downsiomyia* species normally inhabit water in treeholes but have been collected occasionally from bamboo stumps, fallen bamboo and bamboo internodes (Macdonald & Traub 1960). Macdonald (1957) reported *Fl. albolateralis* biting humans in the forest canopy (75 feet high) and less commonly at ground level in Malaysia. *Finlaya niveoides* and *Fl. vana* were collected biting humans in a swamp-forest in Malaysia (Wharton 1962) and Macdonald *et al.* (1965) reported small numbers of *Fl. nivea*-subgroup species biting humans by day and early evening in Malaysia. Shriram *et al.* (2005) found 96 of 3,625 *Fl. nivea* (identification ?, see discussion below under *Fl. nivea*) naturally infected with *Wuchereria bancrofti* filarial parasites in the Andaman and Nicobar Islands of India.

Distribution

Species of the genus occur in the Oriental Region and adjoining areas of the Australasian and Palaearctic Regions.

Discussion

Downsiomyia is diagnosed and distinguished from other generic-level taxa of Aedini by the following combinations of characters: females by (1) vertex with only broad, decumbent scales whereas erect, forked scales restricted to occiput, (2) antennal pedicel with few small scales and short setae mesally, (3) eyes contiguous or separated by one eye facet or less, (4) maxillary palpus and proboscis dark-scaled, (5) acrostichal and dorsocentral areas without setae, (6) scutal fossa entirely covered (or rarely nearly covered) with narrow, curved, pale scales, (7) supraalar area dark-scaled (rarely pale scales extending to anterior margin of supraalar area, e.g. *Do. nipponica*), (8) scutellum with broad scales on all lobes, (9) paratergite, postspiracular area, subspiracular area and metameron without scales, (10) wing entirely dark-scaled, and (11) hindtibia, hindtarsus and distal part of hindfemur dark-scaled; female genitalia by (1) both tergum VIII and sternum VIII with numerous broad scales, (2) tergum IX comprised of two narrow, lateral lobes connected by narrow, basal strip and each lobe with 1–5 setae apically, (3) insula liplike with few long setae in lateral patches, (4) upper vaginal sclerite small, (5) cercus short, broad, with lateral part of distal margin oblique and bearing 5–7 stout setae that decrease in length laterally, and (6) only single large spermathecal capsule (Nivea Group) or one large and two smaller spermathecal capsules (Albonivea Group); males by (1) flagellar whorls of antenna with numerous long setae directed dorsally and ventrally and distal two flagellomeres elongate, (2) maxillary palpus slender, dark-scaled and slightly shorter to approximately equal to length of proboscis, (3) fore- and midtarsi with unequal ungues, each with one tooth, whereas hind ungues are equal, simple; male genitalia by (1) gonocoxite somewhat triangular in outline, proximal part relatively broad, distal part narrow and bearing several to numerous long, broad, fusiform scales (except *Do. mikrokopion*) on mesal area of ventral surface, and basomesal area of dorsal surface with narrow, thumblike lobe bearing several short setae, (2) gonostylus relatively narrow, short and with long, narrow gonostylar claw apically, (3) aedeagus simple, scooplike and with one or usually two or more small teeth apically on each side, and (4) claspette developed as moderately long column bearing one long, somewhat leaf-like, longitudinally striated, claspette filament apically; pupae by (1) seta 1-CT long, moderately stout to stout and normally single whereas setae 2,3-CT are short and slender, (2) seta 5-CT noticeably longer than seta 4-CT and often long and single, (3) seta 11-CT long, stout and single, (4) seta 3-I long and stout, longer than seta 6-I, (5) seta 6-VII short, with few slender branches, inserted posterior to seta 9-VII which is moderately long, stout and normally with 2 or 3 aciculate branches, (6) seta 9-VIII long, stout, multiple-branched, aciculate and inserted on posterior lateral corner of segment, and (7) paddle without hairlike spicules on

margin and midrib well developed; and fourth-instar larvae by (1) seta 1-C single with distal part attenuate, (2) setae 4-7-C well developed, multiple-branched and aciculate, (3) antenna with spicules and seta 1-A with multiple aciculate branches, (4) seta 12-I present, (5) seta 7-I long, stout, single or 2-branched whereas seta 7-II is short and multiple-branched, (6) seta 6-II shorter than seta 6-III, (7) setae 2-VIII and 4-VIII both single, (8) segment X with saddle incomplete ventrally and seta 1-X attached, and (9) pecten on siphon with numerous, evenly spaced spines and seta 1-S branched, aciculate and inserted distal to pecten.

Illustrations and descriptions of *Downsiomyia* species are found in articles listed above in the “previous usage” section.

Downsiomyia females have large, pale-scaled patches covering the scutal fossae, which also occur in females of the Gubernatoris Group of “*Ochlerotatus*” (“*Finlaya*”). However, the latter group is easily distinguished by the presence of an elongate, transverse patch of pale scales anterior to the wing base that extends mesally. In contrast, the supraalar area is dark-scaled in *Downsiomyia*. The pale-scaled scutal fossa of *Downsiomyia* is also superficially similar to some *Stegomyia*, as evidenced by the type species, *Do. nivea*, which was originally assigned to that genus. *Stegomyia* are readily distinguished by possessing white-scaled areas on the supraalar area of the scutum, apex of the maxillary palpus and the hindtarsomeres, as well as numerous differences in the other life stages. The simple aedeagus bearing small teeth apically in *Downsiomyia* males is somewhat similar to some species of *Ochlerotatus* Lynch Arribalzaga, e.g. *Oc. cantans* (Meigen) and *Oc. excrucians* (Walker), but these species differ in numerous other characters.

We conducted an extensive search of the literature on Culicidae, but were unable to find the formal synonymy of *Downsiomyia* with *Finlaya*. This synonymy was listed, without explanation in *A Synoptic Catalog of the Mosquitoes of the World* by Stone *et al.* (1959), and again in Knight & Stone (1977). *Downsiomyia*, as the Niveus Subgroup, was last reviewed by Colless (1958, 1959), who treated 19 species. Knight (1946) previously reviewed nine species, including *Do. nivea*. Reinert (2002) indicated that the female genitalia could be easily separated into two assemblages of species that are similar but distinguished as follows: “Niveus Assemblage” with a single, large, spermathecal capsule, and tergum IX usually with three or four setae on each lateral lobe apically (at least on one lateral lobe); and the “Alboniveus Assemblage” with one large and two medium-sized spermathecal capsules, and tergum IX with only one or two setae apically on each lateral lobe. These two “Assemblages” of species are treated herein as Species Groups. Reinert (2002) separated species treated here from other groups of *Finlaya* based on features of the female genitalia. Reinert (1981, 2000c) noted that groups of species in *Paraedes* Edwards also possess different numbers of spermathecal capsules.

Rozeboom & Cabrera (1964) incriminated *Do. nivea* as a vector of nocturnally periodic *Wuchereria bancrofti* (Cobbold) in the Philippine Islands and Gould *et al.* (1982)

indicated that *Do. harinasutai* was the primary vector of subperiodic *Wuchereria bancrofti* in Thailand. Zagaria & Savioli (2002) also list these two species as vectors of filariasis. Rudnick *et al.* (1986) reported that species of *Downsiomyia* (as “Niveus Subgroup”) were canopy vectors of dengue virus in Malaysia.

Description of type species, *Downsiomyia nivea* (Ludlow), 1903

FEMALE. *Head:* Maxillary palpus dark brown-scaled, 0.18 length of proboscis; proboscis dark brown-scaled, 1.13–1.20 length of forefemur; clypeus brown, bare; antenna brown, 0.80 length of proboscis, pedicel brown with few small, dark brown scales and short brown setae mesally; eyes contiguous above antennal pedicels; ocular line narrow, covered with overlapping, broad, silvery scales; ocular setae numerous; vertex covered with overlapping, broad, dark brown, decumbent scales; occiput with numerous blackish-brown, erect, forked scales; postgena covered with overlapping, broad, silvery scales. *Thorax:* Scutum covered with narrow scales except narrow, bare, median area of prescutellar space, anterior 0.65–0.70 covered with silvery scales extending to anterior and lateral margins, remainder of scutum with dark brown scales, setae as follow: few short on anterior promontory, several on antedorsocentral area, few along lateral margin of scutal fossa, few on antealar area, numerous dark ones on supraalar area with those anterior to wing base shorter, 3–5 laterally on each side of prescutellar area, and 1,2 on parascutellar area; scutellum with overlapping, broad, dark brown scales on each lobe, median lobe with 4,5 long and 6,7 short setae, lateral lobe with 2,3 long and 5,6 short setae; mesopostnotum bare; anteprepronotum with numerous setae on dorsal and lateral surfaces, patch of overlapping, broad, silvery scales posterior to setae; pleural areas with all scales overlapping, broad, silvery except postpronotum; proepisternum with 7–10 setae and patch of scales on upper area, lower area bare; postpronotum with 4,5 posterior setae, narrow band of several moderately broad to broad, brown scales on dorsal margin; prespiracular area, subspiracular area, paratergite, mesomeron, metameron and metepisternum bare; postspiracular area with 7–13 setae; mesokatepisternum with 4,5 setae in row on upper margin and 11–15 along posterior margin, one near middle very long, large upper and small lower posterior patches of scales; prealar area with 10–13 setae on upper area and patch of scales on lower area; mesepimeron with 13–17 short, upper posterior setae, large triangular patch of scales extending from near upper part to near lower part of anterior margin caudal to area in front of setae; metameron with upper margin well above base of hindcoxa. *Legs:* Ante- and postprocoxal membranes bare; fore-, mid- and hindcoxa with large patch of overlapping, broad, silvery scales on anterior surface, forecoxa with patch extending over lateral surface, midcoxa also with several broad, brown scales ventral to silvery scales; fore- and midfemur with anterior surface dark brown-scaled, forefemur with broad, white-scaled patch extending from near base to near midlength of posterior surface, midfemur with short, white-scaled stripe near base

ventrally on posterior surface, hindfemur with distal part dark brown-scaled, anterior surface with proximal 0.78 white-scaled except few brown scales on basal margin, posterior surface similar to anterior surface but proximal white-scaled area slightly shorter; fore-, mid- and hindtibiae and fore-, mid- and hindtarsi dark brown-scaled; fore- and midungues each with 1 tooth; hindungues simple. *Wing*: Dark brown-scaled; vein R_2 longer than R_{2+3} ; remigium with 2 short, dark, posterior setae mainly obscured by scales; alula with row of brown scales on posterior margin; upper calypter with row of numerous setae on margin; halter with stem pale, knob dark brown-scaled. *Abdomen*: Terga I-VIII with several setae on lateral and posterior margins, dark brown-scaled, I-VII each with large, basolateral patch of overlapping, silvery scales, IV with band incomplete, VIII with median, dorsobasal patch of silvery scales; sterna II-VII with broad, brown scales, basolateral areas with patches of broad, silvery scales that connect to form narrow, basal bands on posterior 3,4 sterna; segments VI, VII laterally compressed.

Female genitalia. *Tergum VIII*: Proximal 0.20–0.35 retracted into segment VII; numerous broad, spatulate scales covering distal 0.73–0.79, both dark and pale scales present; setae on distal 0.51–0.65; VIII-Te index 0.53–0.59; length 0.28–0.31 mm; width 0.51–0.56 mm. *Sternum VIII*: Apex with minute (0.02–0.04 of VIII-S length), median emargination separating broad, flattened lobe with lateral portion slightly angled posterolaterally on each side; broad, spatulate scales on distal 0.76–0.81; setae on distal 0.94–0.97; VIII-S index 0.88–0.91; length 0.39–0.42 mm; width 0.44–0.47 mm. *Tergum IX*: With 3–5 setae on each lateral lobe apically, 6–9 total setae; length 0.10–0.12 mm; width 0.11–0.14 mm. *Insula*: With 5–8 setae. *Postgenital lobe*: Apex broadly rounded or flat; 10–17 setae on each side of midline, 21–32 total setae; ventral PGL/cercus index 0.78–0.87; dorsal PGL index 1.00–1.16; ventral PGL index 1.50–1.71; ventral length 0.13–0.14 mm. *Cercus*: Oblique, apical margin with 6 stout setae; dorsal surface with setae on distal 0.47–0.52; cercus index 1.45–1.68; cercus/dorsal PGL index 1.59–1.91; length 0.16–0.17 mm; width 0.09–0.11 mm. *Spermathecal capsule*: One large capsule.

MALE. *Head*: Antenna brown, 0.74–0.75 length of proboscis; maxillary palpus dark brown-scaled, 0.96 length of proboscis, palpomere 3 long, narrow, slightly upturned and bearing few long, brown setae ventroapically, palpomeres 4,5 slightly down-turned; proboscis 1.26 length of forefemur; vertex with broad scales creamy-brown. *Thorax*: Scutum with anterior, silvery-scaled area more extensive, extending posteriorly nearly to prescutellar area and covering most of supraalar area, median posterior margin nearly straight or with small, blunt extension; postpronotum with only few broad, brown scales on upper posterior area. *Legs*: Fore- and midungues unequal, each with 1 tooth, hindungues equal, both simple. *Abdomen*: Sterna VI, VII with several semierect, brown scales.

Male genitalia. *Tergum IX*: Heavily pigmented; length relatively short; pair of narrow, short, pointed lobes on posterior margin each bearing 3–5 long, stout, slightly curved

setae, with longitudinal fold on lateral side of each lobe; anterior margin concave mesally. *Gonocoxite*: Heavily pigmented but with mesal surface lightly pigmented and membranous; moderately long; proximal part of dorsal surface moderately wide with narrow, thumblike lobe on basomesal margin, lobe with several short, relatively slender setae extending from base to apex, elongate patch of numerous short, slender setae on most of mesal area, patch of several long, narrow, fusiform scales on basolateral area, several relatively long, broad, dark spatulate scales on lateral area distal to these scales; lateral surface with numerous broad scales similar to those of dorsal surface and extending over entire ventral surface except on small basomesal area, several long, stout setae on distal portion of lateral and few on apical area of ventral surface; ventral surface with proximal part wide and distal part narrow presenting triangular outline, with double row of long, broad, fusiform scales on distal 0.54 of mesal margin, few moderately long setae on mesal margin proximal to fusiform scales, 5,6 short, slender setae on basomesal area. *Gonostylus*: Short, 0.39 length of gonocoxite; narrow throughout length but distal part slightly narrower, curved mesally and bearing one short, fine seta subapically on mesal surface; gonostylar claw attached apically to gonostylus, narrow, long, 0.53 length of gonostylus, distal part slightly curved, apex truncate. *Proctiger*: Paraproct heavily pigmented, narrow, terminating in short, curved, beaklike point, basal area curved and without sternal arm; 1,2 short, thin, cercal setae. *Tergum X*: Heavily pigmented, narrow; extending from base of paraproct tergomesally, apex attached to ventral surface of tergum IX lobe. *Phallosome*: Aedeagus moderately long, simple, scooplike, lateral margins gently convex with distal portion narrower, apex concave with 2 or 3 small teeth on each side laterally; paramere heavily pigmented, narrow, 0.89 length of aedeagus; basal piece heavily pigmented, short, moderately broad to broad. *Claspette*: With moderately long, narrow, spiculate, columnar stem bearing 2 short, slender setae at about midlength, basal area with narrow lateral extension connected to ventral area of basomesal, thumblike lobe of gonocoxite, mesal area connected to its mate by spiculate, troughlike aedeagal guide; claspette filament attached apically to stem, long, somewhat leaf-like, longitudinally striated, expanded on middle part and narrowing to point distally. *Sternum IX*: Moderately to heavily pigmented, 2,3 moderately long, moderately stout setae on posteromesal area.

PUPA. *Cephalothorax*: Lightly pigmented with some moderately pigmented areas; seta 1-CT very long, stout, with 2 branches; 2-CT relatively short, slender, single; 3-CT short, slender, with 2 branches; 4-CT short, slender, with 4,5 branches; 5-CT very long, stout, single; 6-CT short, slender, single; 7-CT moderately long, moderately stout, with 2-5 branches; 8-CT moderately long, slender, with 2,3 branches; 9-CT moderately long, slender, single; 10-CT moderately long, slender, single to 4-branched; 11-CT long, stout, single; 12-CT long, slender, with 3-5 branches; 11-CT > 12-CT > 10-CT length. *Trumpet*: Moderately long with apex relatively broad; moderately pigmented; tracheoid weakly developed at base; index 2.91-3.10; pinna 0.37-0.39 of trumpet length. *Abdomen*: Terga

and sterna covered with numerous minute spicules in short rows; punctures present on terga III–V, located posterior to seta 4; sterna II–VII each with transverse, curved ridge near base; seta 1-I moderately long, stout, fanlike, with multiple, brush-tipped branches; 2-I short, slender, single or 2-branched; 3,6-I long, stout, single; 4-I short, slender, with 3–6 branches; 5-I short, slender, with 4–7 branches; 7-I moderately long, slender, with 2,3 branches; 9-I short, slender, single; 10-I short, slender, single or 2-branched; 0-II minute, single; 1-II moderately long, slender, with 4–9 branches; 2-II short, moderately stout, single, inserted anterior and lateral to 1,3-II; 3-II long, stout, single, inserted lateral to 1-II; 4-II short, slender, with 5,6 branches; 5-II short, slender, with 2,3 branches, inserted posterior and slightly lateral to 4-II; 6-II relatively long, slender, single to 3-branched; 7-II moderately long, slender, with 2 branches; 9-II short, slender, single; 0,14-III minute, single; 1-III moderately long, slender, with 3–6 branches; 2-III short, moderately stout, single, inserted anterior and approximately in line with 3-III; 3-III long, stout, single, inserted anterior and slightly mesal to 1-III; 4-III short, slender, with 2–4 branches; 5,7-III short, slender, with 2,3 branches; 6-III moderately long, slender, single; 8-III short, slender, with 3–5 branches; 9,11-III short, slender, single; 10-III moderately long, slender, with 2 branches; 0,14-IV minute, single; 1-IV short, slender, with 2–4 branches; 2-IV short, moderately stout, single, inserted anterior and mesal to 1-IV; 3-IV short, slender, with 3–5 branches; 4,9,11-IV short, slender, single; 5-IV very long, greater than 1.5 times length of tergum IV, stout, single; 6-IV moderately long, slender, single; 7-IV short, slender, with 2–4 branches; 8-IV short, slender, with 3,4 branches; 10-IV moderately long, slender, single to 3-branched; 0,14-V minute, single; 1-V short, slender, single or 2-branched; 2-V short, moderately stout, single, inserted anterior and mesal to 1,3-V; 3-V moderately long, slender, single or 2-branched, inserted anterior and very slightly mesal to 1-V; 4-V short, slender, with 3 branches; 5-V very long, greater than 1.5 times length of tergum V, stout, single; 6-V moderately long, slender, single or 2-branched; 7-V moderately long, slender, single to 6-branched; 8-V short, slender, with 3,4 branches; 9,11-V short, slender, single; 10-V moderately long, slender, single; 0,14-VI minute, single; 1-VI short, slender, single or 2-branched; 2-VI short, moderately stout, single, inserted anterior and very slightly mesal to 1-VI; 3,9,11-VI short, slender, single; 4-VI short, slender, with 3 branches; 5-VI long, approximately equal to length of tergum VII, stout, single; 6-VI moderately long, slender, with 2 branches; 7,10-VI moderately long, slender, single; 8-VI short, slender, with 3 branches; 0,14-VII minute, single; 1,4-VII short, slender, with 2 branches; 2-VII short, moderately stout, single, inserted anterior and mesal to 1-VII; 3-VII short, slender, single or 2-branched; 5-VII moderately long, moderately stout, single; 6-VII short, slender, with 5,6 branches, inserted posterior and slightly mesal to 9-VII; 7,10-VII moderately long, slender, single; 8-VII short, slender, with 4,5 branches; 9-VII moderately long, stout, with 5–7 aciculate branches; 11-VII short, slender, single; 0-VIII minute, single; 4-VIII moderately long, slender, single; 9-VIII moderately long, stout, with 8–10 aciculate branches, inserted near posterolateral corner; 14-VIII

short, slender, single. *Paddle*: Broadly ovoid; short, stout spicules on outer margin except basal area and proximal 0.35; index 1.27–1.36; midrib extending to apex; seta 1-Pa short, moderately stout, with 2,3 branches.

FOURTH-INSTAR LARVA. *Head*: Seta 1-C relatively slender, distal part attenuate, single; 2-C absent; 3-C short, single; 4-C moderately long, fanlike, with 7–11 aciculate branches; 5-C long, with 6–12 aciculate branches; 6-C long, fanlike, with 8–10 aciculate branches; 7-C long, fanlike, with 9–12 aciculate branches; $4-C < 5-C < 6-C < 7-C$ length, but 5,6-C approximately equal length, and 7-C slightly longer; 4-7-C noticeably posterior to anterior margin of cranium; 4-C mesal and slightly posterior to 6-C; 5-C posterior to 4,6-C, lateral to 4-C, slightly mesal to 6-C; 7-C mesal and slightly posterior to antennal base, lateral to 4-6-C, slightly anterior to 4-C, anterior to 5-C, approximately in line with 6-C; 8-C short, slender, with 2,3 branches; 9-C short, slender, with 4,5 branches; 10-C short, slender, with 2 branches; 11-C moderately long, fanlike, with 15–21 lightly aciculate branches; 12-C short, slender, with 3,4 branches, mesal to 13-C; 13-C moderately long, slender, single (2-forked on 1 side of 1 specimen); 14-C short, stellate, with 4–10 branches; 15-C short, moderately stout, with 2–4 branches, inserted on anterior 0.20 of labiogula; 18-C short, single. *Antenna*: Moderately long; lightly pigmented with approximately distal 0.50 slightly darker; several scattered spicules, more numerous on proximal 0.50; seta 1-A moderately long, extending beyond apex of antennal shaft, with 6–10 lightly aciculate branches, inserted 0.54 from apex. *Mouthparts*: Lateral palatal brush with mesal filaments comb-tipped, simple filaments laterally; seta 1-Mx short, stellate, with 4–9 branches; dorsomentum with 15–17 teeth. *Thorax*: Seta 0-P short, slender, with 9,10 branches; 1-P moderately long, with 5,6 aciculate branches; 2-P moderately long, lightly aciculate, single; 3-P short, stellate, with 7–14 branches; 1-3-P inserted on moderately pigmented, common setal support plate; $1-P > 2-P > 3-P$ length; 4-P moderately long, slender, single or 2-branched; 5-P moderately long, moderately stout, with 3,4 aciculate branches; 6-P moderately long, moderately stout, aciculate, single; 6-P greater than 5-P length; 7-P moderately long, moderately stout, with 4,5 aciculate branches; 5-7-P each inserted on separate, moderately pigmented, small, setal support plate; 8-P short, stellate, with 6,7 branches; 9-P moderately long, with 4,5 branches; 10-P moderately long, single; 10-P less than 12-P length; 12-P moderately long, single (rarely 2-branched); 11-P short, with 2,3 branches; 9-12-P on moderately pigmented, common tubercle; 14-P short, slender, with 2–4 branches; 1-M short, stellate, with 6–9 branches, 1,2-M approximately equal length; 2,3-M short, single; 4-M short, slender, with 2 branches; 5,7-M long, moderately stout, aciculate, 5-M inserted on moderately pigmented, small tubercle, $5-M > 7-M$ length; 6-M long, moderately stout, fanlike, with 5–7 aciculate branches, 6,7-M inserted on moderately pigmented, common tubercle; 8-M long, moderately stout, fanlike, with 7,8 aciculate branches, inserted on moderately pigmented tubercle; 9-M long, moderately stout, with 7–9 aciculate branches; 10,12-M single,

aciculate, 10-M stout, long, longer than 9,12-M; 11-M very short, slender, with 3,4 branches; 9-11-M inserted on moderately pigmented, common tubercle with well developed lateral spine; 13-M short, slender, with 10-13 branches; 14-M short, stellate, with 8-13 branches; 1-T short, stellate, with 7-11 branches; 2-T moderately long, slender, single; 3-T short, stellate, with 9-14 branches; 4-T short, slender, with 4,5 branches; 5-T short, single or with 2 stiff branches; 6-T moderately long, slender, single; 7-T long, stout, fanlike, with 10-14 aciculate branches, inserted on moderately pigmented tubercle; 8-T short, slender, with 7-12 branches; 9-T long, stout, with 5-7 aciculate branches; 10-T very long, stout, aciculate, single; 10-T greater than 9,12-T length; 11-T short, slender, with 3,4 branches; 12-T short, slender, single; 9-12-T inserted on moderately pigmented, common tubercle with short lateral spine; 14-T short, stellate, with 5-11 branches. *Abdomen*: Setae 0,14-II-VIII minute, single, except 14-V,VI single or 2-branched, 14-VII with 2,3 branches; 1-I-VI, 2-I-VII, 4-I,II, 5-II-VI, 7-IV, 9-I,III,IV, 11-I, 13-I-VI stellate, 2-I-VI with stouter branches, 9-II,V,VI with stiff branches; 6-I-VI, 7-I long, stout, aciculate, each attached to small, moderately pigmented, tubercle; 1-I with 6-16 branches; 2-I with 6-8 branches; 3-I moderately long, slender, with 2 branches; 4-I with 8-11 branches; 5-I short, slender, with 5-11 branches; 6-I with 4-7 branches; 7-I single; 9-I with 3,4 branches; 10-I moderately long, slender, single; 11-I with 5,6 branches; 12-I short, slender, single; 13-I with 5-7 branches; 1-II with 5-8 branches; 2-II with 4-6 branches; 3-II moderately long, with 2 branches; 4-II with 5-9 branches; 5-II with 4,5 branches; 6-II with 4,5 branches; 7-II with 4-6 branches; 8-II short, slender, with 2-4 branches; 9-II with 2-4 branches; 10,12-II moderately long, slender, single; 11-II short, slender, with 2 branches; 13-II with 7-10 branches; 1-III with 5-9 branches; 2-III with 5-8 branches; 3-III moderately long, slender, with 2 branches (rarely single); 4-III short, slender, with 3-5 branches; 6-III with 2 branches; 7-III short, slender, with 8-10 branches; 8-III short, slender, single; 9-III with 3-5 branches; 10-III moderately long, slender, single; 11,12-III short, slender, with 2,3 branches; 13-III with 5-9 branches; 1-IV with 3,4 branches; 2-IV with 6,7 branches; 3-IV moderately long, slender, with 2 branches; 4-IV short, slender, with 2-4 branches; 5-IV moderately long, with 4-6 branches; 6-IV with 2 branches; 7-IV with 6-8 branches; 8-IV short, slender, single; 9-IV with 3-6 branches; 10-IV moderately long, slender, single; 11,12-IV short, slender, with 2,3 branches; 1-V with 3-5 branches; 2-V with 4-8 branches; 3-V moderately long, slender, single (rarely 2-branched); 4-V short, slender, with 3-6 branches; 5-V moderately long, with 3-6 branches; 6-V with 2 branches; 7-V short, slender, with 7-10 branches; 8-V short, slender, single; 9-V with 2-5 branches; 10-V moderately long, slender, single; 11-V short, slender, with 2,3 branches; 12-V short, slender, with 2 branches; 13-V with 4-6 branches; 1-VI with 3,4 branches; 2-VI with 5-7 branches; 3-VI moderately long, single or 2-branched; 4-VI moderately long, slender, with 2 branches; 5-VI moderately long, with 3,4 branches; 6-VI with 2 branches; 7-VI short, slender with 3-5 branches; 8-VI short, slender, with 3-6 branches; 9-VI with 2-4 branches; 10-VI moderately long, slender, single; 11-VI short, slender with 2 branches;

12-VI moderately long, slender, single to 3-branched; 13-VI with 6–12 branches; 1-VII long, moderately stout, with 3,4 branches; 2-VII with 2,3 branches; 3-VII short, slender, with 3,4 branches; 4-VII moderately long, slender, single; 5-VII moderately long, slender, with 3,4 branches; 6-VII short, slender, with 6–10 branches; 7-VII short, slender, with 3,4 branches; 8-VII short, slender, with 8–16 branches; 9-VII short, slender, with 3–5 branches; 10,12-VII moderately long, slender, single; 11-VII very short, slender, single or 2-branched; 13-VII moderately long, with 4,5 branches; 1-VIII moderately long, moderately stout, with 4,5 branches; 2,4-VIII moderately long, slender, single; 3-VIII moderately long, moderately stout, inserted on moderately pigmented tubercle, with 3–6 aciculate branches; 5-VIII moderately long, moderately stout, with 7–9 aciculate branches; comb with 14–16 scales in curved row, scales comprised of long, heavily pigmented, pointed spine with minute spicules along much of lateral margins extending to subapical area; segment X with dorsal saddle heavily pigmented, anterior margin darker, moderate size, incomplete ventrally, acus absent, with numerous, moderately long, stout spicules on posterior margin between setae 1-X and 3-X, numerous rows of minute spicules scattered over remainder of surface, dorsal siphon/dorsal saddle index 2.12–2.20, seta 1-X relatively long (longer than dorsal saddle length), moderately stout, with 2 aciculate branches, 2-X long, moderately stout, with 8,9 branches, 3-X very long, stout, single, ventral brush with 10,11 setae, fanlike with short stems and 7–11 branches, anterior 2 setae precratal, remainder with transverse grid bar, and posterior 5 or 6 setae with well developed, lateral grid bars, 4 anal papillae relatively short, broad, with apices pointed. *Siphon*: Heavily pigmented; acus small, detached; index 2.00–2.06 (dorsal length/width at midlength); pecten on proximal 0.52–0.57 of siphon, with 16–21 evenly spaced spines, each with tooth moderately long, relatively stout, with stout subbasal tooth and usually with 2–4 minute spicules proximally, and apex pointed, proximal 2,3 spines shorter; seta 1-S moderately long, moderately stout, with 7–10 aciculate branches, inserted distal and ventral to last pecten spine; 2-S relatively long, single; 6-S short, single; 8-S short, slender, with 2,3 branches; 9-S very short, single; spiracular apodeme relatively short, ventral arm narrow, with moderately narrow, uniformly wide lobe extending proximally, apex bluntly rounded.

Bionomics

The larval specimens examined by us were collected from treeholes and cut bamboo in the Philippine Islands.

Discussion

The above description of *Do. nivea* is based on our examination of the lectotype female, one paralectotype female, two females, two males, nine pupal exuviae, and nine larval exuviae collected in the Philippine Islands. These specimens were identified as *Ae. (Fin.) nivea* by the late Kenneth L. Knight and their identity confirmed by us. Huang & Rueda (1998) discussed the type series of this species.

Published illustrations of *Do. nivea* include the adult female (Huang & Rueda 1998: Fig. 1), the female genitalia (Reinert 2002: Fig. 8), the male genitalia (Knight 1946: Figs. 5 and 9), the pupa (Knight & Chamberlain 1948: Fig. 33; Baisas 1974: Fig. 9e,g), and the larva (Knight 1946: Fig. 16; Knight & Hull 1951: Fig. 12; Baisas 1974: Fig. 9a–d,f).

We agree with Knight (1946) who stated “Because of the number of closely related species, literature records of *nivea* must be considered with caution, unless careful descriptions are appended.” This was further emphasized by Huang & Rueda (1998) who considered that the species described and illustrated by Tewari & Hiriyan (1995, as *Ae. niveus*) from the Andaman and Nicobar Islands of India is not conspecific with the type specimens from the Philippine Islands. We agree with Huang & Rueda. This is obvious from our descriptions of the larva and pupa of *Do. nivea* from the Philippine Islands, which differ considerably from the illustrations of Tewari & Hiriyan.

Acknowledgments

Appreciation is expressed to Kenneth J. Linthicum (CMAVE) for providing research facilities to JFR; to Rampa Rattanarithikul (Museum of World Insects and Natural Wonders, Chiang Mai, Thailand) and Graham B. White (Department of Entomology and Nematology, University of Florida, Gainesville, FL) for reviewing the manuscript; to James E. Pecor and Thomas V. Gaffigan (WRBU), Theresa M. Howard (NHM), and the late Kenneth L. Knight, for the loan of specimens.

References

- Ahmed, T.U. (1988) Checklist of the mosquitoes of Bangladesh. *Mosquito Systematics* (1987), 19, 187–200.
- Amerasinghe, F.P. (1982) Observations on the mosquitoes (*Diptera: Culicidae*) [sic] of Udawat-takele Forest, Sri Lanka. *Journal National Science Counsel, Sri Lanka*, 10, 81–97.
- Apiwathnasorn, C. (1986) *A list of mosquito species in Southeast Asia*. Museum Reference Centre, SEAMEO-TROPED National Centre Thailand, Mahidol University, Bangkok, Thailand, 73 pp.
- Army Mosquito Project. (1965) *Preliminary Keys to the Mosquitoes of Vietnam*. Department of Entomology, Smithsonian Institution, U.S. National Museum, Washington, DC, 76 pp. + pl. I–IV.
- Aslamkhan, M. (1971) The mosquitoes of Pakistan I. A checklist. *Mosquito Systematics Newsletter*, 3, 147–159.
- Baisas, F.E. (1974) *The Mosquito Fauna of Subic Bay Naval Reservation Republic of the Philippines*. Technical Report Number 72-2, Headquarters, First Medical Service Wing (PACAF), APO San Francisco 96274, 170 pp. + figs. 1–79.
- Banks, C.S. (1906) A list of Philippine Culicidae with descriptions of some new species. *Philippine Journal of Science*, 1, 977–1005.
- Barnett, H.C. (1962) The incrimination of arthropods as vectors of disease. *XI Internationaler Kon-*

- gress fur Entomologie, Wien* (1960), 2, 341–345.
- Barnett, H.C. & Toshioka, S. (1951) *The Bloodsucking Insects, Mites and Ticks of Korea and their Relation to Disease Transmission*. Japan Logistical Command, 406th Medical General Laboratory, APO 500, 25 pp.
- Barraud, P.J. (1923) A revision of the culicine mosquitoes of India. Part VI. Some Indian species of the genus *Finlaya* Theo. adult stage. *Indian Journal of Medical Research*, 11, 475–493 + pls. XXII–XXXIII.
- Barraud, P.J. (1924) A revision of the culicine mosquitoes of India. Part VIII. Further descriptions of Indian species of *Finlaya* Theo. *Indian Journal of Medical Research*, 11, 845–865 + pls. XLVIII–LVII.
- Barraud, P.J. (1934) *The Fauna of British India, Including Ceylon and Burma. Diptera, Vol. V. Family Culicidae. Tribes Megarhinini and Culicini*. Taylor and Francis, London, United Kingdom, 463 pp. + pls. I–VIII.
- Basio, R.G. (1971) *The Mosquito Fauna of the Philippines (Diptera Culicidae)*. Monograph No. 4. National Museum of the Philippines, Manila, Philippines, 198 pp.
- Basio, R.G., Corcega, A.V. & Madriga, M.E. (1973) On Philippine mosquitoes, XI. The species at the Manila International Airport and its environs with notes on their medical importance (Diptera: Culicidae). *Philippine Scientist*, 10, 11–34.
- Bhat, H.R. (1975) A survey of haematophagous arthropods in western Himalayas, Sikkim and Hill Districts of West Bengal: Records of mosquitoes collected from Himalayan Region of Uttar Pradesh with ecological notes. *Indian Journal of Medical Research*, 63, 1583–1608.
- Blanchard, R. (1905) *Les Moustiques Histoire Naturelle et Medicale*. F. R. de Rudeval, Imprimeur-Editeur, Paris, France, 673 pp.
- Bohart, R.M. (1945) *A Synopsis of the Philippine Mosquitoes* NAVMED 580. U.S. Naval Medical Research Unit Number 2, 88 pp. + figs. 1–91.
- Bohart, R.M. (1946) *A key to the Chinese Culicine Mosquitoes*. NAVMED 961, Bureau of Medicine and Surgery, Navy Department, Washington, DC, 23 pp.
- Bohart, R.M. (1957) Insects of Micronesia, Diptera: Culicidae. *Insects of Micronesia, Bernice P. Bishop Museum, Honolulu, HI* (1956), 12(1), 1–85.
- Bohart, R.M. & Ingram, R.L. (1946) *Mosquitoes of Okinawa and Islands in the Central Pacific*. NAVMED 1055, Bureau of Medicine and Surgery, Navy Department, Washington, DC, 110 pp.
- Bonne-Wepster, J. (1954a) Synopsis of a hundred common non-anopheline mosquitoes of the Greater and Lesser Sundas, the Moluccas and New Guinea. *Documenta de Medicina Geographica et Tropica*, 6, 208–246.
- Bonne-Wepster, J. (1954b) *Synopsis of a Hundred Common Non-anopheline Mosquitoes of the Greater and Lesser Sundas, the Moluccas and New Guinea*. Royal Tropical Institute Amsterdam Special Publication Number CXI, Department of Tropical Hygiene & Geographical Pathology Number 20. Elsevier Publishing Company, Amsterdam, The Netherlands, 147 pp.
- Bonne-Wepster, J. & Brug, S.L. (1937) Nederlandsch-Indische Culicinen. *Geneeskundig Tijdschrift Nederlandsch-Indie*, 77, 1–105.
- Bonne-Wepster, J. & Brug, S.L. (1939) Larven van Nederlandsch-Indische Culicinen. *Geneeskundig Tijdschrift voor Nederlandsch-Indie*, 79(20), 1218–1279.
- Borel, E. (1930) *Les Moustiques de la Cochinchine et du Sud-Annam*. Masson et cie, Editeurs, Libraires de l'Academie de Medecine, Paris, France, 423 pp.
- Brug, S.L. (1926) The geographical distribution of mosquitoes in the Malayan Archipelago. *Mededeelingen van den dienst der Volksgezondheid in Nederlandsch-Indie*, Part 4, 471–482.
- Brug, S.L. (1931) XXXII. Culiciden der Deutschen Limnologischen Sunda-Expedition. In: *Sonder-Abdruck aus dem Archiv fur Hydrobiologie, Supplementne IX. Tropische Binnengewasser*, 2, 1–42.

- Brug, S.L. (1934) Notes on Dutch East Indian mosquitos [sic]. *Bulletin of Entomological Research*, 25, 501–519.
- Brug, S.L. (1939) Notes on Dutch East-Indian mosquitoes. *Overgedrukt uit het Tijdschrift voor Entomologie*, 82, 91–113.
- Brug, S.L. & Bonne-Wepster, J. (1947) The geographical distribution of the mosquitoes of the Malay Archipelago. *Chronica Naturae*, 103, 179–197.
- Brug, S.L. & Edwards, F.W. (1931) Fauna Sumatrensis (Bijdrage Nr. 68), Culicidae (Diptera). *Overgedrukt uit het Tijdschrift voor Entomologie*, 74, 251–261.
- Brug, S.L. & Haga, J. (1923) Aanteekening omtrent muskieten. *Overgedrukt uit het Geneeskundig Tijdschrift Nederlandsch-Indie*, 63, 635–640.
- Brunetti, E. (1907) XXV. — Annotated catalogue of Oriental Culicidae. *Records of the Indian Museum (Calcutta)*, 1, 297–377.
- Brunetti, E. (1912) X. Annotated catalogue of Oriental Culicidae — supplement. *Records of the Indian Museum (Calcutta)*, 4, 403–517.
- Brunetti, E. (1920) I. Catalogue of Oriental and south Asiatic Nematocera. *Records of the Indian Museum (Calcutta)*, 17, 1–300.
- Cabrera, B.D. & Rozeboom, L.E. (1964) Filariasis in Palawan, Philippine Islands. *Nature*, 202(4933), 725–726.
- Cai, H-Q. (1984) Additions and corrections to the checklist of mosquitoes in Fujian Province (Diptera, Culicidae). *Wuyi Science Journal*, 4, 209–217.
- Carter, H.F. (1950) Ceylon mosquitoes: Lists of species and names of mosquitoes recorded from Ceylon. *Ceylon Journal of Science (B)*, 24, 85–115.
- Causey, O.R. (1937) Some anopheline and culicine mosquitoes of Siam with remarks on malaria control in Bangkok. *American Journal of Hygiene*, 25, 400–420.
- Chau, G-W. (1982). *An Illustrated Guide to the Identification of the Mosquitoes of Hong Kong*. Urban Council Public Information Unit, Better Printer Company, Hong Kong, 53 pp.
- Chow, C.Y. (1949a) Culicine mosquitoes collected in western Yunnan, China during 1940–1942. *Proceedings of the Entomological Society of Washington*, 51, 127–132.
- Chow, C.Y. (1949b) Observations on mosquitoes breeding in plant containers in Yunnan. *Annals of the Entomological Society of America*, 42, 465–470.
- Chow, C.Y. (1950) Collection of culicine mosquitoes (Diptera, Culicidae) in Taiwan (Formosa), China, with description of a new species. *Quarterly Journal of the Taiwan Museum, Taipei*, 3, 281–287.
- Chow, C.Y., Thevasagayam, E.S. & Tharumarajah, K. (1954) Insects of public health importance in Ceylon. *Revista Ecuatoriana de Entomologia y Parasitologia*, 2, 105–150.
- Chu, F-I. (1957) Collection of megarhine and culicine mosquitoes from Hainan Island, south China, with description of a new species. *Acta Zootaxonomica Sinica*, 9, 145–163 + pls. I–III.
- Chu, I-H. (1957) Seasonal variations in mosquito density in South Korea. *Korean Journal of Biology* (1956), 2, 1–7.
- Colless, D.H. (1957) Notes on the culicine mosquitoes of Singapore III.—Larval breeding-places. *Annals of Tropical Medicine and Parasitology*, 51, 102–116.
- Colless, D.H. (1958) Notes on the culicine mosquitoes of Singapore IV.—The *Aedes niveus* subgroup (Diptera, Culicidae): Introduction and description of five new species and of one new subspecies. *Annals of Tropical Medicine and Parasitology*, 52, 468–483.
- Colless, D.H. (1959) Notes on the culicine mosquitoes of Singapore V.—The *Aedes niveus* subgroup (Diptera, Culicidae): Previously described species and keys to adults and larvae. *Annals of Tropical Medicine and Parasitology*, 53, 166–179.
- Darsie, R.F., Jr., Courtney, G.W. & Pradhan, S.P. (1993) Notes on the mosquitoes of Nepal: III. Additional new records in 1992 (Diptera: Culicidae). *Mosquito Systematics*, 25, 186–191.
- Darsie, R.F., Jr. & Pradhan, S.P. (1990) The mosquitoes of Nepal: Their identification, distribution

- and biology. *Mosquito Systematics*, 22, 69–130.
- Darsie, R.F., Jr., Pradhan, S.P. & Vaidya, R.G. (1991) Notes on the mosquitoes of Nepal: I. New country records and revised *Aedes* keys (Diptera, Culicidae). *Mosquito Systematics*, 23, 39–49.
- Darsie, R.F., Jr., Pradhan, S.P. & Vaidya, R.G. (1992) Notes on the mosquitoes of Nepal: II. New species records from 1991 collections. *Mosquito Systematics*, 24, 23–28.
- Debenham, M.L. & Hicks, M.M. (1989) *The Culicidae of the Australasian Region Volume 12, Summary of Taxonomic Changes, Revised Alphabetic List of Species, Supplementary Bibliography, Errata and Addenda, Geographic Guide to Species, Synopsis of Disease Relationships, Indexes*. Monograph Series, Entomology Monograph Number 2. University of Queensland and University of Sydney in collaboration with Commonwealth Department of Community Services and Health, Australian Government Publishing Service, Canberra, Australia, 217 pp.
- Delfinado, M.D., Viado, G.B. & Coronel, L.T. (1963) A checklist of Philippine mosquitoes with a larval key to genera (Diptera, Culicidae). *The Philippine Journal of Science* (1962), 91(4), 433–457.
- Dyar, H.G. (1920) A collection of mosquitoes from the Philippine Islands. *Insector Inscitae Menstruus*, 8, 175–186.
- Dyar, H.G. & Shannon, R.C. (1925) The types of Philippine mosquitos [sic] described by Ludlow and other notes on the fauna (Diptera, Culicidae) [sic]. *Insector Inscitae Menstruus*, 13, 66–89.
- Edwards, F.W. (1913) New synonymy in Oriental Culicidae. *Bulletin of Entomological Research*, 4, 221–242.
- Edwards, F.W. (1917) Notes on Culicidae, with descriptions of new species. *Bulletin of Entomological Research*, 7, 201–229.
- Edwards, F.W. (1921) A revision of the mosquitos [sic] of the Palaearctic Region. *Bulletin of Entomological Research*, 12, 263–351.
- Edwards, F.W. (1922a) A synopsis of adult Oriental culicine (including megarhinine and sabethine) mosquitoes. Part I. *Indian Journal of Medical Research*, 10, 249–293.
- Edwards, F.W. (1922b) A synopsis of adult Oriental culicine (including megarhinine and sabethine) mosquitoes. Part II. *Indian Journal of Medical Research*, 10, 430–475.
- Edwards, F.W. (1926) XIV.—Diptera Nematocera from the mountains of Borneo. *Journal of Sarawak Museum*, 3, 243–278 + pls. 9–10.
- Edwards, F.W. (1928) Diptera Nematocera from the Federated Malay States Museums. *Journal of the Federated Malay States Museums*, 14(1), 139 pp. + pls. I–II.
- Edwards, F.W. (1929) Philippine nematoceros Diptera II. *Natulae Entomologicae*, 9, 1–14.
- Edwards, F.W. (1932) *Genera Insectorum. Diptera, Fam. Culicidae*. Fascicle 194, Desmet-Vertheuil, Imprimeur-Editeur, Bruxelles, Belgium, 258 pp. + pl. 1–5.
- Evenhuis, N.L. & Gon, III, S.M. (1989) Family Culicidae. In: Evenhuis, N.L. ed. *Catalog of the Diptera of the Australasian and Oceanica Regions*. Bishop Museum Special Publication 86, Bishop Museum Press, Honolulu, HI, pp. 191–218.
- Feng, L.-C. (1938a) A critical review of literature regarding the records of mosquitoes in China Part II. Subfamily Culicinae, tribes Megarhinini and Culicini. *Peking Natural History Bulletin*, 12, 285–318.
- Feng, L.-C. (1938b) The tree hole species of mosquitoes of Peiping, China. *Chinese Medical Journal*, Supplement 2, pp. 503–525.
- Feng, L.-C. (1958) *Collection of Chinese Culicidae*. Peking, Science Press, 250 pp.
- Furumizo, R.T. & Rudnick, A. (1979) Laboratory observations on the life history of two species of the *Aedes (Finlaya) niveus* subgroup (Diptera: Culicidae) in Malaysia. *Journal of Medical Entomology*, 15, 573–575.
- Gandahasada, D., Dennis, D.T., Saroso, J.S., Simanjuntak, C.H., Olson, J., Lee, V., Nalim, S., Budi-arso, I., Sukaeri, Suryatman, Simanjuntak & Suwasono, H. (1984) Infectious disease risks in

- the transmigration area, Way Abung III, Lampung Province II. *Bulletin Penelitian Kesehatan*, 12, 1–10.
- Gerberg, E.J., Barnard, D.R. & Ward, R.A. (1994) *Manual for Mosquito Rearing and Experimental Techniques*. Bulletin Number 5 (revised), American Mosquito Control Association, Incorporated, Lake Charles, LA, 98 pp.
- Gould, D.J., Bailey, C.L. & Vongpradist, S. (1982) Implication of forest mosquitoes in transmission of *Wuchereria bancrofti* in Thailand. *Mosquito News*, 42, 560–564.
- Gould, D.J., Yuill, T.M., Moussa, M.A., Simasathien, P. & Rutledge L.C. (1968) An insular outbreak of dengue hemorrhagic fever III. Identification of vectors and observations on vector ecology. *The American Journal of Tropical Medicine and Hygiene*, 17, 609–618.
- Grothaus, R.H., Floore, T.G., Stasiak, R.S. & Miner, W.F. (1971) A partial list of the mosquitoes of I Corps, Republic of Vietnam, with notes on bionomics. *U.S. Naval Medical Field Research Laboratory, Camp Lejeune, North Carolina*, 21(16), 1–31.
- Gutsevich, A.V. & Dubitsky, A.M. (1981) New species of mosquitoes in USSR fauna. *Novyye vidy komarov fauny Sovetskogo Soyuz, Nauka*, pp. 97–165.
- Gustevich, A.V., Monchadskii, A.S. & Shtakel'berg, A.A. (1974) *Fauna of the U.S.S.R. Diptera. Mosquitoes [sic] Family Culicidae [sic]*. Academy of Sciences of the USSR, Zoological Institute, New Series Number 100 (translated from Russian, Israel Program for Scientific Translations, Jerusalem), 408 pp.
- Haga, J. (1924) Aanteekening omtrent muskieten (II). *Overgedrukt uit helt Geneeskundig Tijdschrift voor Nederlandsch-Indie*, 5, 815–834.
- Hara, J. (1957) Studies on the female terminalia of Japanese mosquitoes (with 48 original plates). *Japanese Journal of Experimental Medicine*, 27, 45–91.
- Harbach, R.E. & Knight, K.L. (1980) *Taxonomists' Glossary of Mosquito Anatomy*. Plexus Publishing, Incorporated, Marlton, NJ, 415 pp.
- Harbach, R.E. & Knight, K.L. (1982) Corrections and additions to *Taxonomists' Glossary of Mosquito Anatomy*. *Mosquito Systematics* (1981), 13, 201–217.
- Harinasuta, C., Sucharit, S., Deesin, T., Surathin, K. & Vutikes S. (1970) Bancroftian filariasis in Thailand, a new endemic area. *Southeast Asian Journal of Tropical Medicine and Public Health*, 1, 233–245.
- Harinasuta, C., Guptavanij, P. & Vasuvat, C. (1974) Studies on the medical ecological epidemiology in mangrove areas in Thailand. *Southeast Asian Journal of Tropical Medicine and Public Health*, 5, 105–127.
- Harrison, B.A., Rattanarithikul, R., Peyton, E.L. & Mongkolpanya, K. (1991) Taxonomic changes, revised occurrence records and notes on the Culicidae of Thailand and neighboring countries. *Mosquito Systematics* (1990), 22, 196–227.
- Hawley, W.A. (1988) The biology of *Aedes albopictus*. *Journal of the American Mosquito Control Association*, 4 (Supplement), 1–39.
- Horsfall, W.R. (1955) *Mosquitoes Their Bionomics and Relation to Disease*. Ronald Press Company, New York, NY, 723 pp.
- Hsiao, T.Y. (1945) *Epidemiology of Diseases of Naval Importance in China*. U.S. NAVMED 630, Bureau of Medicine and Surgery, Navy Department, Washington, DC, 149 pp. + App. A–I.
- Hsiao, T-Y. & Bohart, R.M. (1946) *The Mosquitoes of Japan and Their Medical Importance*. NAVMED 1095, Bureau of Medicine and Surgery, Navy Department, Washington, DC, 44 pp. + figs. 1–27.
- Hsieh, L-K. & Liao, T-H. (1956) A list of Amoy mosquitoes with the description of a new species and a new variety. *Acta Entomologica Sinica*, 6, 123–127.
- Hu, S.M.K. (1937) A brief mosquito survey of Foochow Region, south China. *Lingnan Science Journal*, 16, 579–584.
- Huang, Y-M. & Rueda, L.M. (1998) Description of a paralectotype female of *Aedes (Finlaya)*

- niveus* (Ludlow) (Diptera: Culicidae). *Proceedings of the Entomological Society of Washington*, 100, 824–827.
- International Commission on Zoological Nomenclature. (1999) *International Code of Zoological Nomenclature*. Fourth Edition. International Trust for Zoological Nomenclature, London, United Kingdom, 306 pp.
- Jayasekera, N. & Chelliah, R.V. (1981) An annotated checklist of mosquitoes of Sri Lanka (Diptera: Culicidae) (sic). Publication Number 8. UNESCO, *Man and the Biosphere National Committee of Sri Lanka*, pp. 1–16.
- Joshi, G., Pradhan, S. & Darsie, R.F., Jr. (1965) Culicine, sabethine and toxorhynchitine mosquitoes of Nepal including new country records (Diptera: Culicidae). *Proceedings of the Entomological Society of Washington*, 67, 137–146.
- Kaur, R. (2003) An update on the distribution of mosquitoes of the tribe Aedini in India (Diptera: Culicidae). *Oriental Insects*, 37, 439–455.
- Khamboonruang, C., Thitasut, P., Pan-in, S., Morakote, N., Choochote, W., Somboon, P. & Keha, P. (1987) Filariasis in Tak Province, northwest Thailand: The presence of subperiodic variant *Wuchereria bancrofti*. *Southeast Asian Journal of Tropical Medicine and Public Health*, 18, 218–222.
- Knight, K.L. (1946) Entomology. *The Aedes (Finlaya) niveus subgroup of Oriental mosquitoes* [sic]. *Journal of the Washington Academy of Sciences*, 36, 270–280.
- Knight, K.L. (1969) A new species of the genus *Aedes* [sic], subgenus *Finlaya* [sic] Theobald, from India (Diptera: Culicidae). *Journal of the Kansas Entomological Society*, 42, 382–386.
- Knight, K.L. (1978a) A new *Aedes (Finlaya)* mosquito from Thailand. *Mosquito Systematics*, 10, 106–116.
- Knight, K.L. (1978b) *Supplement to A Catalog of the Mosquitoes of the World (Diptera: Culicidae)*. The Thomas Say Foundation, Entomological Society of America, College Park, MD, 107 pp.
- Knight, K.L. & Chamberlain, R.W. (1948) A new nomenclature for the chaetotaxy of the mosquito pupa, based on a comparative study of the genera (Diptera: Culicidae). *Proceedings of the Helminthological Society of Washington*, 15, 1–10 + figs. 1–35.
- Knight, K.L. & Harrison, B.A. (1988) A new *Aedes (Finlaya)* of the Niveus-Subgroup (Diptera: Culicidae). *Mosquito Systematics* (1987), 19, 212–236 + (errata) 1988, 20, p. 97.
- Knight, K.L. & Hull, W.B. (1951) The *Aedes* mosquitoes of the Philippine Islands I. Keys to species, subgenera *Mucidus*, *Ochlerotatus*, and *Finlaya* (Diptera, Culicidae). *Pacific Science*, 5, 211–251.
- Knight, K.L. & Marks, E.N. (1952) An annotated checklist of the mosquitoes of the subgenus *Finlaya* [sic], genus *Aedes* [sic]. *Proceedings of the United States National Museum*, 101, 513–574.
- Knight, K.L. & Stone, A. (1977) *A Catalog of the Mosquitoes of the World (Diptera: Culicidae)*. Volume VI. The Thomas Say Foundation, Entomological Society of America. College Park, MD, 611 pp.
- Kulasekera, V., Knight, K.L. & Harbach, R.E. (1990) *Aedes (Finlaya) axitiosus*, a new species of the Niveus Subgroup (Diptera: Culicidae) from east Malaysia. *Mosquito Systematics*, 22, 26–33.
- Kurihara, T. (1963) Comparative studies on the pleural structure of the Japanese mosquitoes. *Japanese Journal of Sanitary Zoology*, 14, 191–207.
- Kurihara, T. (1978) Collection records of mosquitoes in Indonesia. *Teikyo Journal of Medicine*, 1, 333–338.
- Kurihara, T. (1981) Cibarial dome of group-A female *Aedes (Finlaya)* mosquitoes. *Japanese Journal of Sanitary Zoology*, 32, 337–338.
- Kurihara T. (1999) *Database Record of Entomological Collections in Reference Museum / Taxon-*

- omy and Ecology Laboratory, Department of Medical Entomology, National Institute of Infectious Diseases Number 1, Family Culicidae (Insecta: Diptera) / Mosquitoes. Department of Medical Entomology, National Institute of Infectious Diseases, Tokyo, Japan, 34 pp.
- Kurihara, T., Shinohara, A. & Kurahashi, H. (2004) Type specimens of mosquitoes (Diptera, Culicidae) deposited in the National Science Museum, Tokyo. *Bulletin of the National Science Museum, Series A (Zoology)*, 30(1), 45–60.
- LaCasse, W.J. (1948) Mosquito survey data on Japan and their application in the control of mosquito-borne diseases. In: *Mosquito Fauna of Japan and Korea (with 78 Original Plates)*. Part I. Office of the Surgeon, Headquarters I Corps APO 301, pp. 1–183.
- LaCasse, W.J. & Yamaguti, S. (1948) Mosquito fauna of Japan and Korea. In: *Mosquito Fauna of Japan and Korea (with 78 Original Plates)*. Part II. Office of the Surgeon, Headquarters I Corps APO 301, pp. 1–273.
- LaCasse, W.J. & Yamaguti, S. (1950) *Mosquito Fauna of Japan and Korea (with 95 Original Plates)*. Office of the Surgeon, Headquarters 8th Army, APO 343, 213 pp.
- Lee, D.J., Hicks, M.M., Griffiths, M., Russell, R.C. & Marks, E.N. (1980) *The Culicidae of the Australasian Region Volume I*. Monograph Series, Entomology Monograph Number 2. School of Public Health and Tropical Medicine, Australian Government Publishing Service, Canberra, Australia, 248 pp.
- Lee, D.J., Hicks, M.M., Griffiths, M., Russell, R.C. & Marks, E.N. (1982) *The Culicidae of the Australasian Region Volume II, Nomenclature, Synonymy, Literature, Distribution, Biology and Relation to Disease. Genera AEDEOMYIA, Genus AEDES (Subgenera [Aedes], Aedimorphus, Chaetocruimyia, Christophersomyia, Edwardsaedes and Finlaya)*. Monograph Series, Entomology Monograph Number 2. School of Public Health and Tropical Medicine, Australian Government Publishing Service, Canberra, Australia, 286 pp.
- Lee, K.W. (1987) Checklist of mosquitoes (Culicidae) in Korea. *Korean Journal of Parasitology*, 25, 207–209.
- Lee, K.W. & Egan, P.J. (1985) *Illustrated Taxonomic Keys to Genera and Species of Female Mosquitoes of Korea. Part I*. Department of the Army, 5th Preventive Medicine Unit, 18th Medical Command, APO San Francisco 96301, 32 pp.
- Lee, K.W. & Lien, J.C. (1970) Pictorial keys to the mosquitos [sic] of Korea. *World Health Organization VBC 70.196*, pp. 1–7.
- Lee, K.W. & Zorka, T. (1987) *Illustrated Taxonomic Keys to Genera and Species of Mosquito Larvae of Korea*. Department of the Army, 5th Preventive Medicine Unit, 18th Medical Command, APO San Francisco 96301, 26 pp.
- Lee, V.H., Nalim, S., Olson, J.G., Gubler, D.J., Ksiazek, T.G. & Aep, S. (1984) A survey of adult mosquitoes on Lombok Island, Republic of Indonesia. *Mosquito News*, 44, 184–191.
- Leicester, G.F. (1908) *The Culicidae of Malaya*. Studies from Institute for Medical Research, Federated Malay States. Kelly & Walsh, Limited, Printers, Singapore, pp. 18–261.
- Li, F-S. & Wu, S-C. (1935) On the known species of Chinese Culicini, with a few other species. *Entomology and Phytopathology, Hangchow, China*, 3, 44–88.
- Lien, J.C. (1962) Non-Anopheline mosquitoes of Taiwan: Annotated catalog and bibliography. *Pacific Insects*, 4, 615–549.
- Lien, J.C. (1968) New species of mosquitoes from Taiwan (Diptera: Culicidae) Part III. Five new species of *Aedes*. *Tropical Medicine*, 10, 95–115.
- Lien, J.C., Kawengian, B.A., Partono, F., Lami, B. & Cross, J.H. (1977) A brief survey of the mosquitoes of south Sulawesi, Indonesia, with special reference to the identity of *Anopheles barbirostris* (Diptera: Culicidae) from the Margolembu area. *Journal of Medical Entomology*, 13, 719–727.
- Lien, J.C., Kosman, L., Partono, F., Joesoef, A., Kosin, E. & Cross, J.H. (1975) A brief survey of mosquitoes in north Sumatra, Indonesia. *Journal of Medical Entomology*, 12, 233–239.

- Loy, V.A. & Rowland, J.N. (1963) *Keys for the Identification of Mosquitoes of Viet-Nam*. Entomological Bulletin Number II. 20th Medical Laboratory (Preventive Medicine) (Field), Saigon, Viet-Nam, 55 pp.
- Lu, B. (1981) The *Aedes niveus* group of China (Diptera: Culicidae). *Entomotaxonomia*, 3, 255–263.
- Lu, B. & Li, B-S. (1982) Chapter 1, Identification of Chinese mosquitoes. In: *Handbook for the Identification of Chinese Important Medical Zoology*. People's Hygiene Publishing House, Beijing, China, pp. 1–159, 941–944.
- Lu, B., Cheng, H., Xu, R. & Ji, S. (1988) *A Checklist of Chinese Mosquitoes (Diptera: Culicidae)*. Guizhou People's Publishing House, Guiyang, China, 164 pp.
- Lu, B. & Ji, S. (1997) Subgenus *Finlaya* Theobald, 1903. In: Lu, B., Li, B., Ji, S., Chen, H., Meng, Q., Su, L., Qu, F., Gong, Z. & Zhang, Z. eds. *Diptera: Culicidae I. Fauna Sinica, Insecta*. Volume 8. Science Press, Beijing, China, pp. 99–173, figs. 18–60.
- Lu, B-L. & Su, L. (1987) *A Handbook for the Identification of Chinese Aedine Mosquitoes*. Science Press, Beijing, People's Republic of China, 160 pp.
- Ludlow, C.S. (1903) Some Philippine mosquitoes. *Journal of the New York Entomological Society*, 11, 137–144.
- Ludlow, C.S. (1905) Mosquito notes. — No. 3. *Canadian Entomologist*, 37, 94–102, 129–135.
- Ludlow, C.S. (1911) The Philippine mosquitoes. *Psyche*, 18, 125–133.
- Luh, P-L & Li, B-S. (1980) Notes on the mosquitoes new of China. *Acta Entomologica Sinica*, 23, 330.
- Ma, S-F. (1982) Diptera: Culicidae. *Insects of Xizang*, 2, 157–163.
- Macdonald, W.W. (1957) Malaysian parasites, XVI. An interim review of the non-anopheline mosquitoes of Malaya. In: Andy, J.R. ed. *Malaysian Parasites XVI–XXXIV. Study Number 28*. Institute for Medical Research of the Federation of Malaya, Kuala Lumpur, Malaysia, pp. 1–34.
- Macdonald, W.W. & Traub, R. (1960) Malaysian parasites XXXVII. An introduction to the ecology of the mosquitoes of the lowland dipterocarp forest of Selangor, Malaya. In: Macdonald, W.W. ed. *Malaysian Parasites XXXV–XLIX. Studies Number 29*. Institute for Medical Research of the Federation of Malaya, Kuala Lumpur, Malaysia, pp. 79–109.
- Macdonald, W.W., Smith, C.E.G. & Webb, H.E. (1965) Arbovirus infections in Sarawak: observations on the mosquitoes. *Journal of Medical Entomology*, 1, 335–347.
- Mackie, J.B. (1964) *A Guide to the Species of Culicines Which Give Rise to Domestic Mosquito Nuisance in the Colony of Hong Kong with Notes Concerning Them*. Government Press, Hong Kong, 15 pp.
- Malhotra, P.R. & Mahata, H.C. (1994) Check-list of mosquitoes of northeast India (Diptera: Culicidae). *Oriental Insects*, 28, 125–149.
- Malhotra, P.R., Sarkar, P.K. & Bhuyan, M. (1982) Mosquito survey in Nagaland. *Indian Journal of Public Health*, 26, 163–168.
- Malhotra, P.R., Sarkar, P.K., Das, N.G., Hazarika, S. & John, V.M. (1987) Mosquito survey in Tirap and Subansiri Districts of Arunachal Pradesh. *Indian Journal of Malariology*, 24, 151–158.
- Martini, E. (1929–1931) 11 u. 12. Culicidae. In: Linder, E. *Die Fliegen der palaearktischen Region*, 3, pp. 1–144 (1929), 145–320 (1930), 321–398 (1931), Stuttgart.
- Matsuo, K., Yoshida, Y. & Lien, J.C. (1974) Scanning electron microscopy of mosquitoes II. The egg surface structure of 13 species of *Aedes* from Taiwan. *Journal of Medical Entomology*, 11, 179–188.
- Miyagi, I., Toma, T., Tsukamoto, M., Mogi, M., Horio, M., Cabrera, B.D. & Rivera, D.G. (1985) A survey of the mosquito fauna in Palawan, Mindanao and north Luzon, Republic of the Philippines. *Mosquito Systematics*, 17, 133–146.
- Mogi, M. (1996) Overwintering strategies of mosquitoes (Diptera: Culicidae) on warmer islands may predict impact of global warming on Kyushu, Japan. *Journal of Medical Entomology*, 33,

- 438–444.
- Monchadskii, A.S. (1951) The larvae of bloodsucking mosquitoes of the USSR and adjoining countries (subfam. Culicinae). *Opredeliteli Faune SSSR, Moscow Zoologicheskim Institut Akademiyi Nauk SSSR*, 37, 1–290.
- Moulton, J.C. (1914) The mosquitoes of Borneo. *Report Sarawak Museum*, 13, 46–48.
- Nagpal, B.N. & Sharma, V.P. (1987) Survey of mosquito fauna of Northeastern Region of India. *Indian Journal of Malariology*, 24, 143–149.
- Nakata, G. & Matuo, K. (1960) Taxonomic reexamination of the *Aedes (Finlaya) niveus* subgroup found in Japan 1. On the variations of distinctive characters of larvae collected from a stone basin being accompanied by those of adults derived from them. *Sanitary Injurious Insects*, 5, 1–8.
- O'Connor, C.T. & Sopa, T. (1981) *A Checklist of the Mosquitoes of Indonesia*. NAMRU-SP-45, Special Publication, U.S. Naval Medical Research Unit Number 2, Jakarta, Indonesia, 26 pp.
- Omori, N. (1962) Morphology of undescribed male and immature mosquitoes of aedines in Japan. 1. *Aedes (Finlaya) watasei*. *Endemic Diseases Bulletin of Nagasaki University*, 4, 10–14.
- Ori, S. & Shimogama, M. (1953) Seasonal prevalence of mosquitoes in cow-shed at Nagasaki City. I. The results obtained in 1952. *Journal of Nagasaki Medical Society*, 28, 1020–1026.
- Osima, M. (1952) Mosquitoes of Katsura-shima. *Journal of Nagasaki Medical Society*, 27, 275–280.
- Pae, C.M., Frommer, R.L. Chong, C.S. & Endris, R.G. (1976) The distribution and abundance of mosquitoes collected from light traps in the Republic of Korea during 1974 and 1975. *Journal of the Korean Medical Association*, 19, 398–403.
- Peters, W. & Dewar, S.C. (1956) A preliminary record of the megarhine and culicine mosquitoes of Nepal with notes on their taxonomy (Diptera: Culicidae). *Indian Journal of Malariology*, 10, 37–39.
- Peyton, E.L., Pecor, J.E., Gaffigan, T.V., Tripis, M., Rueda, L.M. & Wilkerson, R.C. (1999) The Johns Hopkins University School of Hygiene and Public Health, Lloyd E. Rozeboom mosquito collection. *Journal of the American Mosquito Control Association*, 15, 526–551.
- Qutubuddin, M. (1972) A new species of *Finlaya* [sic] (Diptera: Culicidae) from Papua. *Pakistan Journal of Health*, 22, 119–120.
- Rahman, S.J., Wattal, B.L. & Sharma, I.D. (1973) Ecology of mosquitoes of Nilgiri Hills (Tamil Nadu) with particular reference to vectors of human diseases. *Indian Journal of Entomology*, 35, 228–246.
- Rajavel, A.R., Natarajan, R. & Vaidyanathan, K. (2005a) Mosquito collections in the Jeypore Hill Tracts of Orissa, India, with notes on three new country records, *Culex (Lophoceraomyia) pilifemoralis*, *Culex (Lophoceraomyia) wilfredi*, and *Heizmannia (Heizmannia) chengi*. *Journal of the American Mosquito Control Association*, 21, 121–127.
- Rajavel, A.R., Natarajan, R. & Vaidyanathan, K. (2005b) Mosquitoes of the mangrove forests of India: Part 1–Bhitarkanika, Orissa. *Journal of the American Mosquito Control Association*, 21, 131–135.
- Rajput, K.B. & Singh, T.K. (1987) Day biting mosquitoes (Diptera: Culicidae) of Manipur. *Entomon*, 12, 21–25.
- Ramachandran, C.P., Cheong, W.H., Sivanandam, S., bin Omar, A.H. & Mahadevan, S. (1970) Filariasis in Ulu Trengganu, West Malaysia: Parasitological and entomological observations. *Southeast Asian Journal of Tropical Medicine and Public Health*, 1, 505–515.
- Ramalingam, S. (1974) A brief mosquito survey of Java, report of visit from 18 June to 15 July 1973. *World Health Organization VBC 74.504*, pp. 1–66.
- Ramalingam, S. & Pillai, A.G. (1973) Ten new records of mosquitoes occurring in west Malaysia. *Southeast Asian Journal of Tropical Medicine and Public Health*, 4, 271–272.
- Rao, T.R. & Rajagopalan, P.K. (1957) Observations on mosquitoes of Poona District, India, with

- special reference to their distribution, seasonal prevalence and biology of adults. *Indian Journal of Malariology*, 11, 1–54.
- Rattanaarithikul, R. & Panthusiri, P. (1994) Illustrated keys to the medically important mosquitos [sic] of Thailand. *Southeast Asian Journal of Tropical Medicine and Public Health*, 25(Supplement 1), 1–66.
- Rattanaarithikul, R., Harrison, B.A., Panthusiri, P. & Coleman, R.E. (2005) Illustrated keys to the mosquitoes of Thailand I. Background; geographic distribution; lists of genera, subgenera, and species; and a key to the genera. *Southeast Asian Journal of Tropical Medicine and Public Health*, 36(Supplement 1), 1–80.
- Ree, H.I., Self, L.S., Hong, H.K. & Lee, K.W. (1973) Mosquito light trap surveys in Korea 1969–1971. *Southeast Asian Journal of Tropical Medicine and Public Health*, 4, 382–386.
- Reinert, J.F. (1981) Medical entomology studies - XV. A revision of the subgenus *Paraedes* of the genus *Aedes* (Diptera: Culicidae). *Contributions of the American Entomological Institute (Ann Arbor)*, 18(4), 1–91.
- Reinert, J.F. (1990) Medical entomology studies - XVII. Biosystematics of *Kenknightsia*, a new subgenus of the mosquito genus *Aedes* Meigen from the Oriental Region (Diptera: Culicidae). *Contributions of the American Entomological Institute (Gainesville)*, 26(2), 1–119.
- Reinert, J.F. (1999a) The dorsal apotome of pupae and fourth-instar larvae of Culicidae (Diptera), a structure of phylogenetic significance. *Journal of the American Mosquito Control Association*, 15, 77–83.
- Reinert, J.F. (1999b) Descriptions of *Zavortinkius*, a new subgenus of *Aedes*, and the eleven included species from the Afrotropical Region (Diptera: Culicidae). *Contributions of the American Entomological Institute (Gainesville)*, 31(2), 1–105.
- Reinert, J.F. (2000a) New classification for the composite genus *Aedes* (Diptera: Culicidae: Aedini), elevation of subgenus *Ochlerotatus* to generic rank, reclassification of the other subgenera, and notes on certain subgenera and species. *Journal of the American Mosquito Control Association*, 16, 175–188.
- Reinert, J.F. (2000b) Comparative anatomy of the female genitalia of genera and subgenera in tribe Aedini (Diptera: Culicidae). Part I. Introduction, preparation techniques, and anatomical terminology. *Contributions of the American Entomological Institute (Gainesville)*, 32(2), 1–18.
- Reinert, J.F. (2000c) Comparative anatomy of the female genitalia of genera and subgenera in tribe Aedini (Diptera: Culicidae). Part V. Genus *Aedes* Meigen. *Contributions of the American Entomological Institute (Gainesville)*, 32(3), 1–102.
- Reinert, J.F. (2002) Comparative anatomy of the female genitalia of genera and subgenera in tribe Aedini (Diptera: Culicidae). Part XIII. Genus *Ochlerotatus* Lynch Arribalza. *Contributions of the American Entomological Institute (Gainesville)*, 33(1), 1–112.
- Reinert, J.F., Harbach, R.E. & Kitching, I.J. (2004) Phylogeny and classification of Aedini (Diptera: Culicidae) based on morphological characters of all life stages. *Zoological Journal of the Linnean Society*, 142, 289–368.
- Riley, W.A. (1932) An annotated list of Canton, China, mosquitoes, based on collections by the late C.W. Howard. *Lingnam Science Journal*, 11, 25–35.
- Rozeboom, L.E. & Cabrera, B.D. (1964) Filariasis in Mountain Province, Luzon, Republic of the Philippines. *Journal of Medical Entomology*, 1, 18–28.
- Ruang-Areerate, T., Kittayapong, P., Baimai, V. & O'Neill, S.L. (2003) Molecular phylogeny of *Walbachia* endosymbionts in Southeast Asian mosquitoes (Diptera: Culicidae) based on *wsp* gene sequences. *Journal of Medical Entomology*, 4, 1–5.
- Rudnick, A., Wallace, H.G., Furumizo, R.T., Burton, J.J.S. & Neill, W.A. (1986) *Dengue Virus Ecology in Malaysia, Further Studies, 1976–1980*. Bulletin Number 23, Institute for Medical Research Malaysia, Kuala Lumpur, Malaysia, pp. 127–153.
- Rueda, L.M. (2004) Pictorial keys for the identification of mosquitoes (Diptera: Culicidae) associ-

- ated with dengue virus transmission. *Zootaxa*, 589, 1–60.
- Sakakibara, M. (1965) Outbreak of mosquitoes in the Dakuma Reservoir Dam after a great flood caused by heavy rains in Ina Valley on June, 1961. *Endemic Diseases Bulletin of Nagasaki University*, 7, 130–141.
- Sarkar, P.K., Das, N.G. & Rao, K.M. (1981) Mosquito fauna in certain places of Dibrugarh District in Assam. *Indian Journal of Medical Research*, 73, 331–334.
- Sasa, M. (1948) I. Synoptic table for the identification of Japanese mosquitoes by larvae and male hypopygium. *Japanese Medical Journal*, 1, 530–534.
- Sasa, M., Kamimura, K. & Miyagi, I. (1977) Mosquitoes. In: *Animals of Medical Importance in the Nansei Islands in Japan*. Sasa, M., Takahasi, H., Kano, R. & Tanaka, H. eds. Shinjuku Shobo Limited, Tokyo, Japan, pp. 137–175.
- Sasa, M. & Nakahashi, Y. (1952) A new species of the *Aedes* (*Finlaya*) *niveus* subgroup of mosquito from Japan. *Japanese Journal of Experimental Medicine*, 22, 257–265.
- Sasa, M., Nakahara, Y., Ushiroku, N., Hashimoto, H., Uno, A., Ogino, T., Miyachi, T., Yokomizo, F., Koyama, S., Akagi, A., Yamaguchi, K., Saito, C. & Kumazawa, H. (1947) studies on mosquitoes (7). Species of lowlands and mountains, observations in the Okayama District. *Medical Biology, Tokyo*, 11, 152–154.
- Scanlon, J.E. & Esah, S. (1965) Distribution in altitude of mosquitoes in northern Thailand. *Mosquito News*, 25, 137–144.
- Scanlon, J.E. & Peyton, E.L. (1965) Mosquito fauna of Thailand. *Southeast Asia Treaty Organization Annual Report*. Medical Study Number 47, U.S. Army-SEATO Medical Research Laboratory, APO San Francisco 96346, pp. C17–C26.
- Senior-White, R. (1923) *Catalogue of Indian Insects Part 2–Culicidae*. Superintendent Government Printing, Calcutta, India, 124 pp.
- Senior-White, R. (1927) Notes on Ceylon mosquitoes, –II. The larvae of the commoner non-anopheline mosquitoes. *Spolia Zeylanica*, 14, 61–76.
- Service, M.W. (1993) Mosquitoes (Culicidae). In: Lane, R.P. & Crosskey, R.W. eds. *Medical Insects and Arachnids*. Chapman and Hall, London, United Kingdom, pp. 120–240.
- Shriram, A.N., Ramaiah, K.D., Krishnamoorthy, K. & Sehgal, S.C. (2005) Diurnal pattern of human-biting activity and transmission of subperiodic *Wuchereria bancrofti* (Filariidea: Dipetalonematidae) by *Ochlerotatus niveus* (Diptera: Culicidae) on the Andaman and Nicobar Islands of India. *American Journal of Tropical Medicine and Hygiene*, 72, 273–277.
- Stojanovich, C.J. & Scott, H.G. (1965) *Illustrated Key to Aedes* [sic] *Mosquitoes of Vietnam*. U.S. Department of Health, Education and Welfare, Public Health Service. Atlanta, GA, 34 pp.
- Stojanovich, C.J. & Scott, H.G. (1966) *Illustrated Key to Mosquitoes of Vietnam*. U.S. Department of Health, Education and Welfare, Public Health Service. Atlanta, GA, 158 pp.
- Stojanovich, C.J. & Scott, H.G. (1995) *Mosquitoes of Asiatic Russia*. Published by authors, 112 pp.
- Stojanovich, C.J. & Scott, H.G. (1996a) *Mosquitoes of Korea*. Published by authors, 146 pp.
- Stojanovich, C.J. & Scott, H.G. (1996b) *Illustrated Key to the Adult Male Mosquitoes of Russia*. Published by authors, 56 pp.
- Stone, A. (1957a) Corrections in the taxonomy and nomenclature of mosquitoes (Diptera, Culicidae). *Proceedings of the Entomological Society of Washington* (1956), 58, 333–344.
- Stone, A. (1957b) Notes on types of mosquitoes in the Hungarian National Museum (Diptera, Culicidae). *Annals of the Entomological Society of America*, 50, 171–174.
- Stone, A. (1961) A synoptic catalog of the mosquitoes of the world, supplement I (Diptera: Culicidae). *Proceedings of the Entomological Society of Washington*, 63, 29–52.
- Stone, A. (1963) A synoptic catalog of the mosquitoes of the world, supplement II (Diptera: Culicidae). *Proceedings of the Entomological Society of Washington*, 65, 117–140.
- Stone, A. (1967) A synoptic catalog of the mosquitoes of the world, supplement III (Diptera: Culicidae). *Proceedings of the Entomological Society of Washington*, 69, 197–224.

- Stone, A. (1970) A synoptic catalog of the mosquitoes of the world, supplement IV (Diptera: Culicidae). *Proceedings of the Entomological Society of Washington*, 72, 137–171.
- Stone, A. & Delfinado, M.D. (1973) Family Culicidae. In: *A catalog of the Diptera of the Oriental Region. Volume I. Suborder Nematocera*. Delfinado, M.D. & Hardy D.E. eds. The University Press of Hawaii, Honolulu, HI, pp. 266–343.
- Stone, A. & Knight, K.L. (1956) Type specimens of mosquitoes in the United States National Museum: II, The genus *Aedes* (Diptera, Culicidae). *Journal of the Washington Academy of Science*, 46, 213–228.
- Stone, A., Knight, K.L. & Starcke, H. (1959) *A Synoptic Catalog of the Mosquitoes of the World (Diptera, Culicidae)*. Volume VI. The Thomas Say Foundation, Entomological Society of America, Washington, DC, 358 pp
- Stone, A., Scanlon, J.E., Bailey, D.L., Delfinado, M.D. & Bram, R.A. (1966) *Preliminary Keys to the Mosquitoes of Vietnam*. First Revision, Contribution Number 127, Army Research Program in Malaria, Department of Entomology, Smithsonian Institution, U.S. National Museum, Washington, DC, 92 pp.+ pl. I–V.
- Strickman, D., Miller, M.E., Kim, H-C. & Lee, K-W. (2000) Mosquito surveillance in the demilitarized zone, Republic of Korea, during an outbreak of *Plasmodium vivax* malaria in 1996 and 1997. *Journal of the American Mosquito Control Association*, 16, 100–113.
- Tanaka, K. (1971) Mosquitoes of the Ryukyu Islands. *Japanese Journal of Sanitary Zoology*, 21, 4.
- Tanaka, K. (2002) Studies on the pupal mosquitoes of Japan (6) *Aedes (Finlaya)* (Diptera, Culicidae). *Japanese Journal of Systematic Entomology*, 8, 137–177.
- Tanaka, K. (2003) Studies on the pupal mosquitoes of Japan (8) A key to species of the genus *Aedes* (Diptera, Culicidae). *Medical Entomology and Zoology*, 54, 105–111.
- Tanaka, K., Mizusawa, K. & Saugstad, E.S. (1979) A revision of the adult and larval mosquitoes of Japan (including the Ryukyu Archipelago and the Ogasawara Islands) and Korea (Diptera: Culicidae). *Contributions of the American Entomological Institute (Ann Arbor)*, 16, 1–987.
- Tanaka, K., Saugstad, E.S. & Mizusawa, K. (1975) Mosquitoes of the Ryukyu Archipelago (Diptera: Culicidae). *Mosquito Systematics*, 7, 207–233.
- Tewari, S.C. & Hiriyani, J. (1995) Description of *Aedes (Finlaya) niveus* (Diptera: Culicidae) from Andaman and Nicobar, India. *Mosquito Systematics*, 27, 167–176.
- Theobald, F.V. (1903) *A Monograph of the Culicidae or Mosquitoes. Mainly Compiled from the Collections Received at the British Museum from Various Parts of the World in Connection with the Investigation into the Cause of Malaria Conducted by the Colonial Office and the Royal Society*. Volume III. British Museum (Natural History), London, United Kingdom, 259 pp. + pls. I–XVII.
- Theobald, F.V. (1905a) A catalogue of the Culicidae in the Hungarian National Museum with descriptions of new genera and species. *Annales Musei Nationalis Hungarici*, 3, 61–119 + pls. I–IV.
- Theobald, F.V. (1905b) *Diptera Fam. Culicidae*. Genera Insectorum. Wytsman P., Bruxelles, 50pp. + pls. 1–2.
- Theobald, F.V. (1907) *A Monograph of the Culicidae or Mosquitoes. Mainly Compiled from Collections Received at the British Museum*. Volume IV. British Museum (Natural History), London, United Kingdom, 639 pp. + pls. I–XVI.
- Theobald, F.V. (1908) XXX. First report on the collection of Culicidae and Corethridae in the Indian Museum, Calcutta, with descriptions of new genera and species. *Records of the Indian Museum*, 2, 287–302.
- Theobald, F.V. (1910) *A Monograph of the Culicidae or Mosquitoes. Mainly Compiled from Collections Received at the British Museum*. Volume V. British Museum (Natural History), London, United Kingdom, 646 pp. + pls. I–VI.
- Thurman, E.B.H. (1959) *A Contribution to a Revision of the Culicidae of Northern Thailand*. Bulle-

- tin A-100. University of Maryland Agricultural Experiment Station, College Park, MD, 182 pp.
- Thurman, E.B. (1963) The mosquito fauna of Thailand (Diptera: Culicidae). *Proceedings of the IX Pacific Science Congress*, 9, 47–57.
- Toma, T. & Miyagi, I. (1986) The mosquito fauna of the Ryukyu Archipelago with identification keys, pupal descriptions and notes on biology, medical importance and distribution. *Mosquito Systematics*, 18, 1–109.
- Townsend, F.C., Chainey, J.E., Crosskey, R.W., Pont, A.C., Lane, R.P., Boorman, J.P.T. & Lowry, C.A. (1990) *A Catalogue of the Types of Bloodsucking Flies*. Occasional Papers on Systematic Entomology, Number 7. Natural History Museum, London, United Kingdom, 371 pp.
- Traub, R. & Macdonald, W.W. (1963) Notes on the ecology of some culicine mosquitoes in Malaya. *Proceedings of the IX Pacific Science Congress*, 9, 44–45.
- Tsukamoto, M. & Horio, M. (1985) Electrophoretic comparison of the lactate dehydrogenase banding pattern among Japanese mosquito larvae (Diptera: Culicidae). *Journal of Medical Entomology*, 22, 491–498.
- Tsukamoto, M., Miyagi, I., Toma, T., Sucharit, S., Tumrasvin, W., Khamboonruang, C., Choochote, W., Phanthumachinda, B. & Phanurai, P. (1987) The mosquito fauna of Thailand (Diptera: Culicidae): An annotated checklist. *Japanese Journal of Tropical Hygiene*, 15, 291–326.
- van Peenen, P.F.D., Atmosoedjono, S., Mulijono, S.E., Lien, J.C., Saroso, J.S. & Light, R.H. (1975) Mosquitoes collected in south and east Kalimantan. *Bulletin Penelitian Kesehatan Health Studies in Indonesia*, 3(2), 21–27.
- Vargas, L. (1950) Los sugeneros de *Aedes*. *Downsiomyia* n. subgen. (Diptera: Culicidae). *Revista del Instituto de Sallubridad y Enfermedades Tropicales (Mexico, D.F.)*, 9, 61–69.
- Wada, Y., Mogi, M., Oda, T., Mori, A., Suzuki, H., Hayashi, K. & Miyagi, I. (1976) Notes on mosquitoes of Amami-Oshima Island and the overwintering of Japanese encephalitis virus. *Tropical Medicine*, 17, 187–199.
- Ward, R.A. (1984) Second supplement to “A Catalog of the Mosquitoes of the World” (Diptera: Culicidae). *Mosquito Systematics*, 16, 227–270.
- Ward, R.A. (1992) Third supplement to “A Catalog of the Mosquitoes of the World” (Diptera: Culicidae). *Mosquito Systematics*, 24, 177–230.
- Wattal, B.L., Bhatia, M.L. & Kalra, N.L. (1958) Some new records of culicines of Dehra Dun (Uttar Pradesh) with a description of a new variety. *Indian Journal of Malariology*, 12, 217–230.
- Wharton, R.H. (1962) *The Biology of Mansonia [sic] Mosquitoes in Relation to the Transmission of Filariasis in Malaya*. Bulletin Number 11, Institute for Medical Research Federation of Malaya, 114 pp.
- White, G.B. (1979) The identification of mosquitoes as vectors of malaria and filariasis. Problems in the identification of parasites and their vectors. *Symposium British Society of Parasitology* (1978), 17, 103–143.
- Wu, C.F. (1940) *Catalogue of Chinese Insects, Order Diptera, Suborder Nematocera, Superfamily Culicoidea, Family Culicidae*. Volume 5. Catalogus Insectorum Sinensium, 524 pp.
- Xu, B.-H. (1984) A preliminary survey of the mosquitoes in Wuyi Mountain, Fujian, China (Diptera, Culicidae). *Wuyi Science Journal*, 4, 205–208.
- Yamada, S. (1932) *Nippon konchu zukan. Iconographia Insectorum Japonicorum*, 1st Edition, Hokuryukan, Tokyo, 2241 pp.
- Zagaria, N. & Savioli, L. (2002) Elimination of lymphatic filariasis: a public-health challenge. *Annals of Tropical Medicine & Parasitology*, 96(Supplement No. 2), S3–S13.