



<http://dx.doi.org/10.11646/zootaxa.3994.3.4>

<http://zoobank.org/urn:lsid:zoobank.org:pub:C55F47BC-F08B-40E3-A802-29BF1B840D6C>

## ***Subsphaerolaimus minor* sp. n. and *Micromicron cephalatum* Cobb, 1920 (Nematoda) from the Yen River Estuary of Vietnam**

VLADIMIR G. GAGARIN<sup>1</sup> & NGUYEN VU THANH<sup>2</sup>

<sup>1</sup>*Institute of Inland Waters Biology, Russia Academy of Sciences, Borok, Yaroslavl Province, 152742, Russia*

<sup>2</sup>*Institute of Ecology and Biological Resources Vietnamese Academy of Sciences and Technologies, 1000 Hanoi, Vietnam*

### **Abstract**

Two nematode species found in Yen River Estuary of Vietnam are described and illustrated. *Subsphaerolaimus minor* sp. n. is similar to *S. lamasus* Gerlach, 1956, but differs from it in the shorter body, comparatively shorter pharynx and shorter cephalic setae. A pictorial key for determination of valid species in the genus *Subsphaerolaimus* Lorenzen, 1978 is given. *Micromicron cephalatum* Cobb, 1920 is redescribed and reillustrated based on numerous males and females. The genus *Micromicron* Cobb, 1920 is confirmed as a valid genus with type and only species, *M. cephalatum* Cobb, 1920.

**Key words:** description, free-living nematodes, taxonomy

### **Introduction**

The free-living nematode fauna of Vietnam has been investigated recently. Increased knowledge of nematode biodiversity has improved the basic biomonitoring data for water quality assessment in watershed and wetland ecosystems along the country's coastline.

### **Material and methods**

The material for this study was obtained from 8 nematode samples collected in November 2013 by members of the Institute of Ecology and Biological Resources, Vietnamese Academy of Science and Technology in Donghg Rhi, Song Yen River, Quang Ninh Province of North Vietnam. The samples were taken using an elastic tube with a mouth diameter of 40 µm and length of 40 cm and then fixed in hot 10% formalin. The nematodes were extracted by a centrifugation method using Ludox-TM 50 Solution, gradually transferred to anhydrous glycerin and finally mounted on permanent slides (de Grisse 1965). The worms were identified, measured and photographed using a Nikon Eclipse 80i light microscope equipped with Nomarski DIC accessories, Nikon DS-Fil digital camera and PC with NIS-Elements D 3.2 software for imaging and analysis.

Abbreviations:

a—body length divided by maximum body diameter;

a.d.—anal or cloacal body diameter, in µm;

am.w.—width of the amphidial fovea, in µm;

b—body length divided by pharyngeal length;

c—body length divided by tail length;

c'—tail length in anal or cloacal body diameter;

c.s.—length of cephalic setae (third ring), in µm;

diam.c.s.—body diameter at the level of lips, in µm;