



The genus *Magia* Distant (Hemiptera: Lophopidae): review and biogeographic history

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Abstract

Kasserota illingworthi Baker, 1925 is transferred to the genus *Magia* Distant, 1907 and placed in synonymy with *Magia subocellata* Distant, 1907. The first description of the male genitalia of this species is provided along with the first description of the fifth instar nymph. A new species for this genus is described, *Magia stuarti* Soulier-Perkins sp. n. The presence of this genus in Australia is discussed in terms of its historical biogeography.

Key words: synonymy, new species, Fulgoromorpha, Australia

Introduction

Magia Distant, 1907, belongs to a monophyletic group of seven genera within the family Lophopidae. All of these are present in New Guinea except for *Magia*, which is found in Australia only. The genera *Maana* Soulier-Perkins, 1998, *Onycta* Fennah, 1955 and *Megacarna* Baker, 1925 are endemic to New Guinea while the genera *Zophiuma* Fennah, 1955, *Kasserota* Distant, 1906, *Jugoda* Melichar, 1915 and *Acarina* Stål, 1863 are also found on some islands surrounding New Guinea (Soulier-Perkins, 2000).

In 1997, a significant number of specimens of *M. subocellata* were collected on *Archontophoenix alexandrae* (F. Muell.) H. Wendl. & Drude (Arecaceae) in a greenhouse in Kuranda, North Queensland. This allowed larvae to be associated with the adults collected and has enabled the first description of the fifth instar nymph to be published. In addition, description and illustrations of the male genitalia have not previously been published and are presented below. In 2003 some specimens of *Magia* were collected by P. Erbe at Cape Tribulation, North Queensland. Their cuticle is paler and the male genitalia are different from those of *M. subocellata*. This new species is described in this paper.

Material and methods

Dried preserved specimens were examined. For observation of the genitalia and the posterior wax plates of the fifth instar nymph, dissection and observations were done according to the following protocol. The abdomens were removed and placed in a 10% KOH bath with 1–2 drops of black chlorazol (Carayon, 1969) as a general endocuticular stain, and boiled for 5–10 minutes. Gross dissection and cleaning of the abdomens were performed in 70% ethyl alcohol. Final and precise dissections were done in distilled water with blue paragon staining and then transferred to glycerol for drawing using a camera lucida.