

email: saranyaphat.khag@gmail.com; kdhyde3@gmail.com





http://dx.doi.org/10.11646/phytotaxa.176.1.11

Clavatispora thailandica gen. et sp. nov., a novel taxon of *Venturiales* (Dothideomycetes) from Thailand

SARANYAPHAT BOONMEE¹, JAYARAMA D. BHAT², SAJEEWA S.N. MAHARACHCHIKUMBURA¹ & KEVIN D. HYDE¹

¹ Institute of Excellence in Fungal Research, Mae Fah Luang University, Chiang Rai 57100, Thailand; School of Science, Mae Fah Luang University, Chiang Rai 57100, Thailand ² Formerly at Department of Botany, Goa University, Goa 403 206, India

Abstract

An unusual wood-inhabiting fungus was found in northern Thailand. The combination of characters of this taxon, viz. setose, ostiolate ascomata, bitunicate asci, dark brown, wedge-shaped to clavate, muriform ascospores, and a hyphomycetous asexual state producing ellipsoidal, septate, highly constricted, holoblastic conidia is not found in any hitherto described genus of Dothideomycetes. An LSU phylogeny showed this taxon to be clearly placed in the family *Sympoventuriaceae*, *Venturiales*. The name *Clavatispora thailandica* gen. and sp. nov., is introduced to accommodate this taxon, which is described and illustrated and compared with other genera in this family.

Key words: bitunicate asci, hyphomycetous asexual state, molecular phylogeny, muriform ascospores, *Sympoventuriaceae*

Introduction

Pleosporales is the largest order in the class Dothideomycetes that comprises 41 families and 181 genera (Hyde *et al.* 2013). In this order, the family *Pleosporaceae* contains several genera, e.g. *Clathrospora, Lewia, Platysporoides, Pleospora,* and *Pyrenophora* that have brown muriform ascospores (Sivanesan 1984, Barr 1987, 1990a, Zhang *et al.* 2012, Hyde *et al.* 2013). Species with muriform ascospores are also found in *Aigialaceae, Cucurbitariaceae, Halojulellaceae, Montagnulaceae, Platystomaceae, Pleomassariaceae, Shiraiaceae* and *Teichosporaceae*, thus this character has evolved on numerous occasions even outside Pleosporales (*e.g. Gloniopsis, Hysterobrevium, Hysterographium* and other genera in *Hysteriales*). Barr (1990a) placed species with muriform ascospores in 12 families within the Pleosporales, *i.e. Arthopyreniaceae, Cucurbitariaceae, Dacampiaceae, Pleomassariaceae, Pleosporaceae* and *Pyrenophoraceae*, *Pleomassariaceae, Plaeosphaeriaceae, Phaeotrichaceae, Pleomassariaceae, Pleosporaceae* and *Pyrenophoraceae*. Species in these families have differing ascomata characters and asexual states (Barr 1990a, 1990b, Zhang *et al.* 2012, Hyde *et al.* 2013).

In our ongoing study of Dothideomycetes of Thailand (Boonmee *et al.* 2011, 2012, Chomnunti *et al.* 2011, 2012a,b, Liu *et al.* 2011, 2012), we collected an intriguing species with ascospores resembling *Alternaria*-like conidia and bitunicate asci indicative of *Pleosporaceae*. The superficial, solitary, ascomata with dark, septate setae, forming on a basal subiculum, and bright-coloured ostiole, indicated that this fungus differed from other dictyosporous genera in families of the Pleosporales (Simmons 1986, Barr 1987, 1990a, 1992). The fungus also developed a *Trichocladium*-like asexual state in culture. The aim of this paper is to introduce this unusual ascomycete, which is described as *Clavatispora thailandica gen. nov., sp. nov.* It is only described and illustrated but also compared with morphologically similar genera. We sequenced our isolate to establish the phylogenetic placement of our taxon. The trees generated using LSU region place our new genus in the *Sympoventuriaceae*, *Venturiales*.

Materials and methods

Collection, examination and isolation of fungi

Dead herbaceous stems were randomly collected from Doi Tung forest, Chiang Rai, Thailand. Microscopic studies were carried out and the fungus illustrated following the procedures outlined in Boonmee *et al.* (2011). Type specimens are deposited at the herbarium of Mae Fah Luang University (MFLU), Chiang Rai, Thailand and cultures are deposited at Mae Fah Luang University Culture Collection (MFLUCC), BIOTEC Culture Collection (BCC) and IFRD culture collections, International Fungal Research & Development Centre, Kunming, China, the latter under MTA no. 4/2010.

Single spore isolation was carried out following the methods outlined by Chomnunti *et al.* (2011). Germinating ascospores were transferred to fresh malt extract agar (MEA, Difco Laboratories, Detroit, Michigan, USA) and incubated at 28°C for one week in darkness. When colonies appeared, subcultures were made onto fresh MEA media at room temperature. Morphology and cultural characters of the fungus were recorded based on MEA cultures after 60 days (Boonmee *et al.* 2011, 2012).

Molecular procedures

Fresh mycelium was scraped from the surface of colonies grown on MEA for 60 days at 28°C. DNA extraction, PCR amplification and sequencing of LSU region using primer pair LROR/LR5 was performed under the conditions described in Boonmee *et al.* (2011). Sequencing was done in Shanghai Sangon Biological Engineering Technology & Services Co., China and new sequences generated in this study were blasted to search for related taxa in GenBank database (www.ncbi.nlm.nih.gov/blast/).

Phylogenetic analyses

Sequence data were aligned with Clustal W (Thompson *et al.* 1994) and manually adjusted using BioEdit (Hall 1999). Most of the taxa used in this study are derived from Schoch *et al.* (2009) and Zhang *et al.* (2011). The LSU dataset for showing the placement of *Clavatispora thailandica* comprises 40 taxa including *Acrospermum compressum (Acrospermales)* as an outgroup (Table 1). Poorly aligned regions on nucleotide multiple alignments were removed using Gblock 0.91b (Castersana 2000), following nonconservative settings; the maximum number of contiguous nonconserved positions was set to 10, the minimum length blocks set to 5, allowing for gap positions in half the sequences. The construction of the maximum likelihood (ML) analysis using RAxML version 7.6.3 (Stamatakis 2006, Stamatakis *et al.* 2008) as part of the "RAxML-HPC BlackBox on TG tool" was performed at the CIPRES Science Gateway V. 3.3 (http://www.phylo.org/portal2/, Miller *et al.* 2010). RAxML rapid bootstrapping and subsequent ML search used distinct model/data partitions with joint branch length optimization, executing 1,000 rapid bootstrap inferences and thereafter a thorough ML search. All free model parameters were estimated by RAxML and ML estimate of 25 per site rate categories. Likelihood of final tree were evaluated and optimized under GAMMA. GAMMA Model parameters were estimated to an accuracy of 0.1000000000 Log Likelihood units. Every 100th tree was saved. Phylogenetic trees were illustrated in Treeview (Page 1996).

Bayesian posterior probability analysis was constructed with the MrBayes 3.1.2 run on XSEDE at the CIPRES webportal (Ronquist & Huelsenbeck 2003), using the parameter setting of 2 parallel runs, 4 chains, run for 4,000,000 generations, sample frequency every 1,000 generations and all other parameters were left as default. The 50% majority rule consensus tree was created from the remaining trees in Treeview (Page 1996).

Results

Phylogenetic studies

The RAxML phylogenetic tree (Fig. 1) was constructed through analysis of the LSU gene region and comprised 40 taxa and 754 aligned characters out of 4% of ambiguous characters. The RAxML phylogram received the best scoring tree with a final ln value of -4136.348470. The isolate of *Clavatispora thailandiaca* (MFLUCC 100107) clustered with *Scolecobasidiella avellanea* (CBS 772.73) and grouped within the family *Sympoventuriaceae* with high support values (98% BS and 1.00 PP). Additionally, taxa from the genera *Ochroconis* and *Scolecobasidium* formed a sister-group to *Clavatispora*. To confirm the placement of new genus we built a further tree including

Dothideomycetes and some species of *Trichocladium* (*Sordariales*) (tree not shown). Results indicate that *Clavatispora* belongs to order *Venturiales* and is well separated from taxa in *Pleosporales*.

	TABLE 1	Taxa	included	in the	phylogenetic study.
--	---------	------	----------	--------	---------------------

Species	Culture no.	GenBank Accession number	
		LSU	
Acrospermum compressum	M 151	EU940084	
Apiosporina collinsii	CBS 118973	GU301798	
Apiosporina morbosa	dimosp	EF114694	
Caproventuria hanliniana	ATCC 96019	AF050290	
Caproventuria hystrioides	CBS 117727	EU035459	
Clavatispora thailandiaca	MFLUCC 100107	KF770458	
Coleroa robertiani	CBS 458.64	JQ036231	
Fusicladium africanum	CPC 12828	EU035423	
Fusicladium amoenum	CBS 254.95	EU035425	
Fusicladium intermedium	CBS 110746	EU035432	
Fusicladium proteae	CPC18282	JN712551	
Gibbera conferta	CBS 191.53	GU301814	
Metacoleroa dickiei	medipc	EF114695	
Metacoleroa dickiei	Kruys503 (UPS)	DQ384100	
Ochroconis calidifluminalis	IFM54738	AB385698	
Ochroconis constricta	NBRC9375	AB564619	
Ochroconis gallopava	IFM54737	AB272164	
Ochroconis humicola	NBRC32054	AB564618	
Ochroconis tshawytschae	CBS100438	GU328005	
Protoventuria barriae	CBS 300.93	JQ036232	
Protoventuria major	CBS 114594	JQ036233	
Scolecobasidiella avellanea	CBS 772.73	EF204505	
Scolecobasidium cateniphorum	P050	EU107309	
Scolecobasidium terreum	P043	EU107306	
Scolecobasidium tricladiatum	P051	EU107286	
Scolecobasidium variabile	P041	EU107310	
Scolecobasidium verruculosum	P044	EU107308	
Spilocaea pomi	CBS176.42	GU348998	
Sympoventuria capensis	CPC 12839	DQ885905	
Sympoventuria capensis	CBS 120136	DQ885906	
Tyrannosorus pinicola	CBS 124.88	DQ470974	
Venturia alpina	CBS 373.53	EU035446	
Venturia asperata	ATCC34052	EF114711	
Venturia carpophila	ICMP5402	AY849967	
Venturia chlorospora	Kruys 502 UPS	DQ384101	
Venturia inaequalis	CBS 815.69	GU301878	
Venturia inaequalis	ATCC 60070	EF114712	
Venturia populina	CBS 256.38	GU323212	
Venturia pyrina	ATCC38995	EF114714	
Veronaeopsis simplex	CBS 588.66	EU041877	

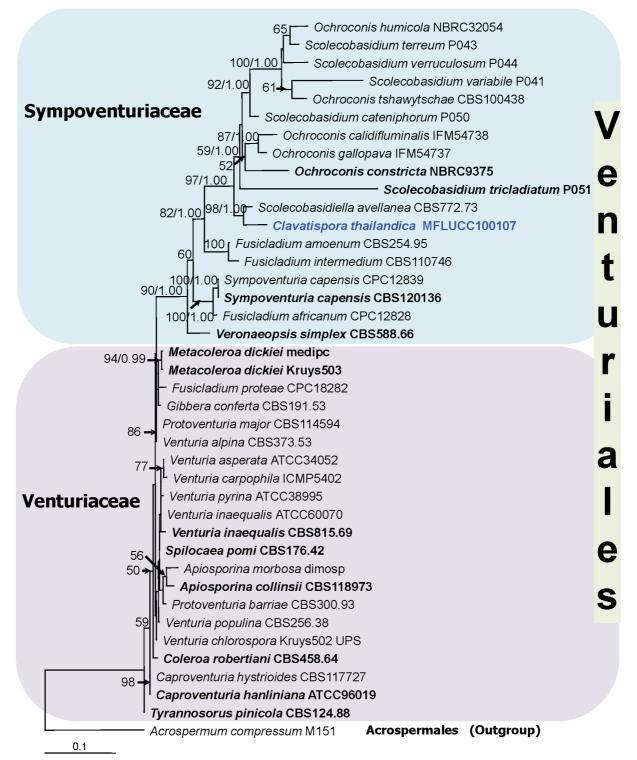


FIGURE 1. RAxML phylogenetic tree based on the LSU sequence data. Bootstrap support values for maximum likelihood (\geq 50% BS) and Bayesian posterior probabilities (\geq 0.95 PP) are shown in the tree (values below these thresholds not shown). The tree was rooted with *Acrospermum compressum (Acrospermales)* as outgroup. The new taxon is highlighted in blue and types are in bold.

Taxonomy

Clavatispora S. Boonmee & K.D. Hyde, gen. nov. MycoBank: MB 805923

Etymology:—"*clavatispora*," name recognizes the clavate ascospores. **Type species:**—*Clavatispora thailandica* S. Boonmee & K.D. Hyde Saprobic on dead stems of herbaceous plants. Sexual state: *Ascomata* superficial, solitary, scattered on substrate, developing on subiculum of brown hyphae, subglobose to globose, dark brown, covered with dark brown, thick-walled, septate setae, and strands of radiating fused hyphae at base, with a barely raised, bright, central ostiole. *Peridium* comprising 6–8 layers of dark brown cells of *textura angularis*, almost black at outside. *Hamathecium* of *ca* 2 µm wide, anastomosing, septate, branched pseudoparaphyses, hyaline, embedded in gelatinous matrix. *Asci* bitunicate, subclavate to broadly obovoid, thick-walled, with a short pedicel, apically rounded, lacking an ocular chamber. *Ascospores* overlapping 2–3-seriate, clavate, asymmetrical, yellowish to reddish brown, becoming dark brown at maturity, muriform, constricted at septa, tapering towards a subacute base, with basal cell pale brown or brownish, smooth-walled. Asexual state: directly developing on MEA. *Mycelium* superficial, pale to moderately dark brown, composed of septate, branched, thin- to thick-walled, smooth-walled, 1–3 µm wide hyphae. *Conidiophores* erect, developing on hyphae, brown or light brown, septate, smooth, sometimes branched. *Conidiogenous cells* holoblastic, subglobose. *Conidia* ellipsoidal to subglobose, pale brown to brown, guttulate, rounded at apex, sub-acute at base, trans-septate, constricted and darkened at septa, smooth-walled.

Type species:—*Clavatispora thailandica* S. Boonmee & K.D. Hyde, *sp. nov.* MycoBank: MB 805924 (Figs 2–3)

Holotype:—THAILAND. Chiang Rai: Mae Fah Luang, Doi Tung, elev. *ca*. 1000 m., on dead stems of unidentified herbaceous plants, 10 June 2009, *Saranyaphat Boonmee* DT04 (MFLU 100038!)—extype living culture = MFLUCC 100107 = BCC 38863 = IFRD 2170.

Sequence data:—LSU = KF770458, SSU = KF770457 and TEF1a = KF770459.

Etymology:-The specific epithet "thailandica" refers to the country from where the fungus was collected.

Saprobic on dead stems of herbaceous plants. Sexual state: Ascomata (115-)130-135 μ m high × (120–)135–150(–160) μ m diam (\overline{x} = 130 × 140 μ m, n = 5), superficial, solitary, scattered on substrate, developing on subiculum of brown hyphae, subglobose to globose, dark brown, covered with dark brown, thick-walled, septate setae, and strands of radiating fused hyphae at base, with a barely raised, bright central ostiole under light microscope. Peridium 16-21 µm wide, comprising 6-8 layers of dark brown cells of textura angularis, almost black at outside. Hamathecium of ca 2 µm wide, anastomosing, septate, branched pseudoparaphyses, hyaline, embedded in gelatinous matrix. Asci 90–110 × 24–30 μ m (\overline{x} = 100 × 28 μ m, n = 20), 8-spored, bitunicate, fissitunicate, subclavate to broadly obovoid, thick-walled, with a short pedicel, apically rounded, lacking an ocular chamber. Ascospores $32-44 \times 10-13 \ \mu m \ (\overline{x} = 37 \times 11 \ \mu m, n = 20)$, overlapping 2–3-seriate, clavate, slightly curved, asymmetrical, yellowish to reddish brown, becoming dark brown at maturity, 4-7(-8) transversely septate, with 1–3 vertical septa in some cells, constricted at septa, tapering towards a subacute base, with basal cell pale brown or brownish, smooth-walled. Asexual state: single spore isolate growing on MEA. Mycelium slightly raised, pale to moderately dark brown, composed of septate, branched, thin- to thick-walled, smooth-walled, $1-3 \mu m$ wide hyphae. Conidiophores erect, developing on hyphae, brown or light brown, septate, smooth, sometimes branched, $(5-)10-15 \ \mu m \ long.$ Conidiogenous cells holoblastic, subglobose. Conidia $(14.5-)15-18(-21) \times 5-7(-8) \ \mu m \ (\overline{x}=$ $17 \times 6 \mu m$, n = 20), ellipsoidal to subglobose, pale brown to brown, guttulate, rounded at apex, sub-acute at base, 3septate, 1–2-septate when immature, constricted and darkened at septa, smooth-walled.

Cultural characteristics:—Ascospores germinating on WA within 12 h and germ tubes produced from spore cells. Colonies on MEA slow growing, reaching 7 mm diameter in 1 week at 28°C, low convex, slightly effuse hairy, edge entire, dark brown. Aerial mycelium, radiating outwards, superficial, septate.

Notes:—The dark brown ascomata, bitunicate asci and muriform brown ascospores of *Clavatispora thailandica* are similar to characteristics of many genera of *Pleosporales* and some taxa in *Capnodiales* and *Hysteriales* (Barr 1990, Boehm *et al.* 2009, Zhang *et al.* 2012, Hyde *et al.* 2013). *Clavatispora* can be distinguished from other genera by its solitary ascomata, covered by superficial, dark setae, a bright ostiole and hyphomycetous asexual state. The hyphomycetous conidia of the asexual state, derived from single ascospores, developed directly on MEA within 60 days. The asexual conidia which formed chains, resemble species of *Trichocladium*, i.e. *T. constrictum* (Goh & Hyde 1999), but *C. thailandica* differs in having branched conidiophores, light pigmented conidia and cells with conspicuous guttules (Fig. 3E–L). *Trichocladium constrictum* has not been sequenced, however, *T. asperum* (generic type) and *T. opacum* are not phylogenetically related to *C. thailandica* (data not shown).

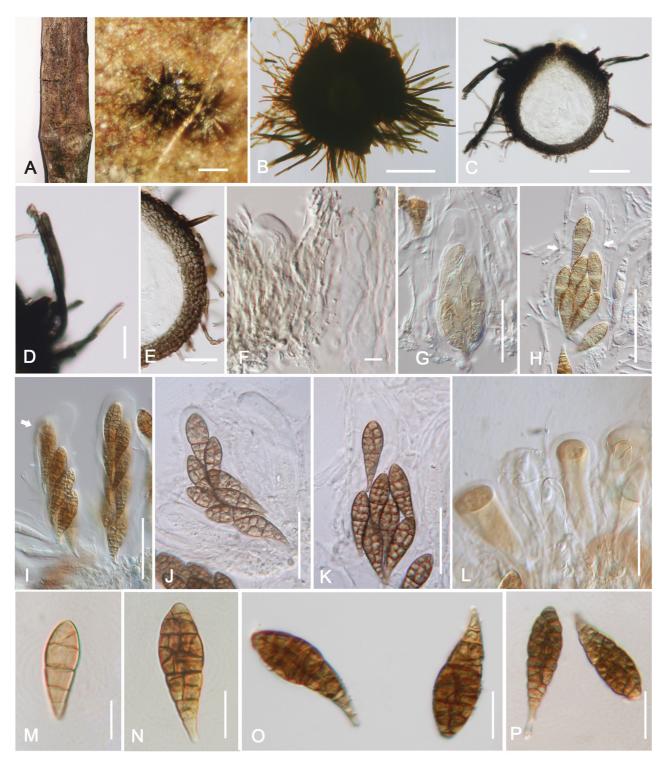


FIGURE 2. Sexual state of *Clavatispora thailandica* (MFLU100038, holotype). A. Substrate and ascomata appearance. B. Squash mount of ascoma. C. Section of ascoma. Note the fused hyphae forming hairs at the base. D. Setae. E. Peridium. F. Pseudoparaphyses. G–H. Immature bitunicate asci with young developing ascospores. I–K. Mature asci. Fissitunicate dehiscence arrowed in H and I. L. Characteristic dehiscence of asci after release of ascospores. M–P. Young and mature ascospores. Scale bars: A–C, G–L = 50 μ m, D–E = 20 μ m, F = 5 μ m, M–P = 10 μ m.

Sequence data placed *Clavatispora* in the order *Venturiales* but it differs in morphology from most other genera. The only genus in *Venturiales* with setose ascomata similar to *Clavatispora* is *Trichodothis* (Zhang *et al.* 2011). However, *Trichodothis* differs in having stromatic ascomata, oval asci and 2-celled, hyaline to pale brown, ornamented ascospores. Molecular data are needed to confirm the placement of *Trichodothis* in *Sympoventuriaceae* as it is rather atypical of the family. The presence of stromatic ascomata covered by spine-like setae, cylindrical

asci, 2-celled and hyaline, smooth ascospores are also found in *Gibbera conferta* and *Venturia compacta* (Peck 1873, Petrak 1947). Furthermore, LSU data analysis paired *G conferta* with an isolate of *Fusicladium proteae* in the *Venturiaceae* clade (Fig. 1). *Venturia compacta* lacks DNA sequence data. *Acantharia, Coleroa, Metacoleroa, Protoventuria* and *Pyrenobotrys* also have setose ascomata, but ascospores of these genera are 1-septate, hyaline or coloured, with smooth or verrucose walls (Barr 1989, Carris & Poole 1993). Phylogenetic analysis based on LSU sequence data shows that *Clavatispora* clusters with the hyphomycete genus *Scolecobasidiella* in *Sympoventuriaceae* as well as *Ochroconis, Scolecobasidium, Fusicladium* and *Veronaeopsis*. The asexual state of *Clavatispora* has 1–3-phragmoseptate, smooth conidia that are deeply constricted at septa, whereas in other genera the conidia and conidiogenous apparatus are mostly verrucose to denticulate.

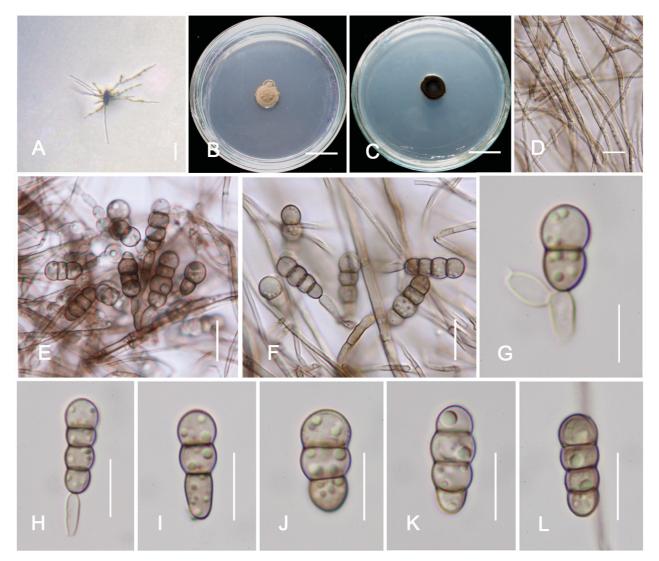


FIGURE 3. Asexual state of *Clavatispora thailandica* (MFLUCC100107, extype living culture). A. Germinating ascospore. B–C. Colonies on MEA, upper surface and lower surface. D. Vegetative hyphae in culture. E–H. Conidiophores with conidia. I–L. Conidia. Scale bars: $A = 10 \ \mu m$, B–C = 1 cm, D = 5 μm , E–L = 10 μm .

Discussion

Clavatispora thailandica, discovered on dead stems of an unidentified herbaceous plant in northern Thailand, is characterized by its setiferous black ascomata, bitunicate asci with fissitunicate dehiscence, with a shrunken ectotunica, endotunica and coloured plasmalemma layers, and clavate, dark brown, muriform ascospores. The unique ascospore features suggest this species belongs in the *Pleosporales*. However, molecular studies place *C*.

thailandica in the order *Venturiales* (Dothideomycetes), although its morphological characters are rather different from all taxa in *Venturiales* (Zhang *et al.* 2012). Members of the order *Pleosporales* are distantly related (data not shown). The phylogenetic analysis based on LSU sequence data show the placement of *Clavatispora* in family *Sympoventuriaceae*, where it clusters with *Scolecobasidiella avellanea* (Fig. 1). Most taxa in *Sympoventuriaceae* are hyphomyceteous, and, with the exception of *Sympoventuria*, the sexual state of most species is unknown (Crous *et al.* 2007, Seifert *et al.* 2011, Samerpitak *et al.* 2014). *Clavatispora* also produced a hyphomyceteous state in culture characterized by erect, coloured, septate, sometimes branched conidiophores and ellipsoidal to subglobose, light coloured, guttulate, 3-septate, deeply constricted conidia (Fig. 3). The asexual state of *Clavatispora*, which is *Trichocladium*-like, differs from all genera in the same clade *i.e. Fusicladium, Ochroconis, Scolecobasidiella, Scolecobasidium* and *Veronaeopsis* (Abbott 1927, Ellis 1971, 1976, Fothergill 1996, Schubert *et al.* 2003, Seifert *et al.* 2011, Martin-Sanchez *et al.* 2012, Zhang *et al.* 2012, Samerpitak *et al.* 2014). In the light of these unique morpho-molecular features, the taxonomic affinity of *Clavatispora thailandica* as a new genus and species is confirmed.

Acknowledgements

This work was supported by TRF/BIOTEC program Biodiversity Research and Training Grant BRT R_251181, BRT R_253012 and partially by the Mushroom Research Foundation, Chiang Rai Province. The International Fungal Research and Development Centre, The Research Institute of Resource Insects, the Chinese Academy of Forestry (Bailongsi, Kunming 650224, China) and Mae Fah Luang University (grant for study Dothideomycetes No. 56101020032) are also thanked for research facilities. Amy Rossman is thanked for her comments on the draft manuscript and thanks are also extended to SMML-USDA for the use of laboratory facilities and for providing Saranyaphat Boonmee with a scholarship to support her stay in USDA. We are grateful to Dr. Eric H.C. McKenzie and Nalin N Wijayawardene for their helpful suggestions.

References

Abbott, E.V. (1927) Scolecobasidium, a new genus of soil fungi. Mycologia 19: 29-31.

- Barr, M.E. (1987) *Prodromus to class Loculoascomycetes*. Published by the Author, Amherst, Massachusetts, University of Massachusetts, U.S.A.
- Barr, M.E. (1989) The Venturiaceae in North America: revisions and additions. Sydowia 41: 25-40.
- Barr, M.E. (1990a) Some dictyosporous genera and species of Pleosporales in North America. *Memoirs of the New York Botanical Garden* 62: 1–92.
- Barr, M.E. (1990b) Melanommatales (Loculoascomycetes). North American Flora Series II 13: 1–129.

Barr, M.E. (1992) Notes on the Lophiostomataceae (Pleosporales). Mycotaxon 45: 191-221.

- Boehm, E.W.A., Mugambi, G.K., Miller, A.N., Huhndorf, S.M., Marincowitz, S., Spatafora, J.W. & Schoch, C.L. (2009) A molecular phylogenetic reappraisal of the Hysteriaceae, Mytilinidiaceae and Gloniaceae (Pleosporomycetidae, Dothideomycetes) with keys to world species. *Studies in Mycology* 64: 49–83. http://dx.doi.org/10.3114/sim.2009.64.03
- Boonmee, S., Zhang, Y., Chomnunti, P., Chukeatirote, E., Tsui, C.K.M., Bahkali, A.H. & Hyde, K.D. (2011) Revision of lignicolous *Tubeufiaceae* based on morphological reexamination and phylogenetic analysis. *Fungal Diversity* 51: 63–102. http://dx.doi.org/10.1007/s13225-011-0147-4
- Boonmee, S., Hyde, K.D., Ko Ko, T.W., Chukeatirote, E., Chen, H., Cai, L., McKenzie, E.H.C., Jones, E.B.G., Kodsueb, R. & Hassan, B.A. (2012) Two new *Kirschsteiniothelia* species with *Dendryphiopsis* anamorphs cluster in *Kirschsteiniotheliaceae* fam. nov. *Mycologia* 104: 698–714. http://dx.doi.org/10.3852/11-089

Carris, L.M. & Poole, A.P. (1993) A new species of Protoventuria on leaves of Vaccinium macrocarpon. Mycologia 85: 93-99.

Castersana, J. (2000) Selection of conserved blocks from multiple alignments for their use in phylogenetic analysis. *Molecular Biology and Evolution* 17: 540–552.

- Chomnunti, P., Schoch, C.L., Aguirre-Hudson, B., Ko Ko, T.W., Hongsanan, S., Jones, E.B.G., Kodsueb, R., Phookamsak, R., Chukeatirote, E., Bahkali, A.H. & Hyde, K.D. (2011) *Capnodiaceae*. *Fungal Diversity* 51: 103–134. http://dx.doi.org/10.1007/s13225-011-0145-6
- Chomnunti, P., Bhat, D.J., Jones, E.B.G., Chukeatirote, E., Bahkali, A.H. & Hyde K.D. (2012a) *Trichomeriaceae*, a new sooty mould family of Chaetothyriales. *Fungal Diversity* 56: 63–76.

http://dx.doi.org/10.1007/s13225-012-0197-2

- Chomnunti, P., Ko Ko, T.W., Chukeatirote, E., Cai, L., Jones, E.B.G., Kodsueb, R., Chen, H., Hassan, B.A. & Hyde, K.D. (2012b) Phylogeny of *Chaetothyriaceae* in northern Thailand including three new species. *Mycologia* 104: 382–395. http://dx.doi.org/10.3852/11-066
- Crous, P.W., Mohammed, C., Glen, M., Verkley, G.J.M. & Groenewald, J.Z. (2007) Eucalyptus microfungi known from culture.
 3. *Eucasphaeria* and *Sympoventuria* genera nova, and new species of *Furcaspora*, *Harknessia*, *Heteroconium* and *Phacidiella*. *Fungal Diversity* 25: 19–36.
- Ellis, M.B. (1971) Dematiaceous hyphomycetes. Commonwealth Mycological Institute, Kew, pp 43–46.
- Ellis, M.B. (1976) More dematiaceous hyphomycetes. Commonwealth Mycological Institute, Kew, pp 190-195.
- Fothergill, A.W. (1996) Identification of dematiaceous fungi and their role in human disease. *Clinical Infectious Diseases* 22: 179–184.
- Goh, T.K. & Hyde, K.D. (1999) A synopsis of Trichocladium species, based on the literature. Fungal Diversity 2: 101–118.
- Hall, T.A. (1999) BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symposium Series* 41: 95–98.
- Hyde, K.D., Jones, E.B.G., Liu, J.K., Ariyawansa, H.A., Boehm, E., Boonmee, S., Braun, U., Chomnunt, P., Crous, P.W., Dai, D.Q., Diederich, P., Dissanayake, A., Doilom, M., Doveri, F., Hongsanan, S., Jayawardena, R., Lawrey, J.D., Li, Y,M., Liu, Y.X., Lücking, R., Monkai, J., Muggia, L., Nelsen, M.P., Pang, K.L., Phookamsak, R., Senanayake, I., Shearer, C.A., Suetrong, S., Tanaka, K, Thambugala, K.M., Wijayawardene, N.N., Wikee, S., Wu, H.X., Zhang, Y., Hudson, B.A., Alias, S.A., Aptroot, A., Bahkali, A.H., Bezerra, J.L., Bhat, J.D., Camporesi, E., Chukeatirote, E., Gueidan, C., Hawksworth, D.L., Hirayama, K., De Hoog, S., Kang, J.C., Knudsen, K., Li, W.J., Li, X., Liu, Z.Y., Mapook, A., McKenzie, E.H.C., Miller, A.N., Mortimer, P.E., Phillips, A.J.L., Raja, H.A., Scheuer, C., Schumm, F., Taylor, J.E., Tian, Q., Tibpromma, S., Wanasinghe, D.N., Wang, Y., Xu, J., Yan, J., Yacharoen, S. & Zhang, M. (2013) Families of Dothideomycetes. *Fungal Diversity* 63: 1–313.

http://dx.doi.org/10.1007/s13225-013-0263-4

- Kirk, P.M., Cannon, P.F., Minter, D.W. & Stalpers, J.A. (2008). Dictionary of the fungi. 10th edn. CABI Bioscience, UK.
- Liu, J.K., Phookamsak, R., Jones, E.B.G., Zhang, Y., Ko Ko T.W., Hu, H.L., Boonmee, S., Doilom, M., Chukeatirote, E., Bahkali, A.H., Wang, Y. & Hyde, K.D. (2011) Astrosphaeriella is polyphyletic, with species in *Fissuroma* gen. nov., and *Neoastrosphaeriella* gen. nov. *Fungal Diversity* 51: 135–154. http://dx.doi.org/10.1007/s13225-011-0142-9
- Liu, J.K., Phookamsak, R., Doilom, M., Wikee, S., Li, Y.M., Ariyawansa, H.A., Boonmee, S., Chomnunti, P., Dai, D.Q., Bhat, D.J., Romero, A.I., Xhuang, W.Y., Monkai, J., Jones, E.B.G., Chukeatirote, E., Ko Ko, T.W., Zhoa, Y.C., Wang, Y, & Hyde, K.D. (2012) Towards a natural classification of Botryosphaeriales. *Fungal Diversity* 57: 149–210. http://dx.doi.org/10.1007/s13225-012-0207-4
- Martin-Sanchez, P.M., Nováková, A., Bastian, F., Alabouvette, C. & Saiz-Jimenez, C. (2012) Two new species of the genus *Ochroconis, O. lascauxensis* and *O. anomala* isolated from black stains in Lascaux Cave, France. *Fungal Biology* 116: 574–589.

http://dx.doi.org/10.1016/j.funbio.2012.02.006

- Miller, M.A., Pfeiffer, W. & Schwartz, T. (2010) Creating the CIPRES Science Gateway for inference of large phylogenetic trees. In: Proceedings of the Gateway Computing Environments Workshop (GCE). New Orleans, LA. CA, San Diego Supercomput. Center, pp. 1–8.
- Page, R.D.M. (1996) TreeView: an application to display phylogenetic trees on personal computers. *Computer Applications in the Biosciences* 12: 357–358.
- Peck, C.H. (1873) Report of the botanist (1871). *Annual Report of the New York State Museum of Natural History* 25: 57–123. Petrak, F. (1947) Über *Gibbera* Fr. und verwandte Gattungen. *Sydowia* 1: 169–201.
- Ronquist, F. & Huelsenbeck, J.P. (2003) MRBAYES 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics*
- 19: 1572–1574. Samerpitak, K., Van der Linde, E., Choi, H.J., Gerrits van den Ende, A.H.G., Machouart, M., Gueidan, C. & De Hoog, G.S.
 - (2014) Taxonomy of *Ochroconis*, genus including opportunistic pathogens on humans and animals. *Fungal Diversity* 65: 89–126.

http://dx.doi.org/10.1007/s13225-013-0253-6

Schoch, C.L., Crous, P.W., Groenewald, J.Z., Boehm, E.W., Burgess, T.I., de Gruyter, J., de Hoog, G.S., Dixon, L.J., Grube, M., Gueidan, C., Harada, Y., Hatakeyama, S., Hirayama, K., Hosoya, T., Huhndorf, S.M., Hyde, K.D., Jones, E.B., Kohlmeyer, J., Kruys, A., Li, Y.M., Lücking, R., Lumbsch, H.T., Marvanová, L., Mbatchou, J.S., McVay, A.H., Miller, A.N., Mugambi, G.K., Muggia, L., Nelsen, M.P., Nelson, P., Owensby, C.A., Phillips, A.J., Phongpaichit, S., Pointing, S.B., Pujade-Renaud, V., Raja, H.A., Plata, E.R., Robbertse, B., Ruibal, C., Sakayaroj, J., Sano, T., Selbmann, L., Shearer, C.A., Shirouzu, T., Slippers, B., Suetrong, S., Tanaka, K., Volkmann-Kohlmeyer, B., Wingfield, M.J., Wood, A.R., Woudenberg, J.H., Yonezawa, H., Zhang, Y. & Spatafora, J.W. (2009) A class-wide phylogenetic assessment of Dothideomycetes. *Studies in Mycology* 64: 1–15.

http://dx.doi.org/10.3114/sim.2009.64.01

Schubert, K., Ritschel, A. & Braun, U. (2003) A monograph of *Fusicladium* s.lat. (Hyphomycetes). *Schlechtendalia* 9: 1–132. Seifert, K.A., Morgan-Jones, G., Gams, W. & Kendrick, B. (2011) *The genera of hyphomycetes*. CBS Fungal Biodiversity

Centre, Utrecht, Netherlands.

Simmons, E.G. (1986) Alternaria themes and variations (22–26). Mycotaxon 25: 287–308.

- Sivanesan, A. (1984) The bitunicate ascomycetes and their anamorphs. J. Cramer, Vaduz, Lichtenstein.
- Stamatakis, A. (2006) RAxML-VI-HPC: Maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. *Bioinformatics* 22: 2688–2690.
- Stamatakis, A., Hoover, P. & Rougemont, J. (2008) A rapid bootstrap algorithm for the RAxML web servers. *Systematic Biology* 57: 758–771.
- Thompson, J.D., Higgins, D.G. & Gibson, T.J. (1994) ClustalW: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position specific gap penalties and weith matrix choice. *Nucleic Acids Research* 22: 4673–4680.
- Zhang, Y., Crous, P.W., Schoch, C.L., Bahkali, A.H., Guo, L.D. & Hyde, K.D. (2011) A molecular, morphological and ecological re-appraisal of Venturiales-a new order of Dothideomycetes. *Fungal Diversity* 51: 249–277. http://dx.doi.org/10.1007/s13225-011-0141-x
- Zhang, Y., Crous, P.W., Schoch, C.L. & Hyde, K.D. (2012) Pleosporales. *Fungal Diversity* 52: 1–225. http://dx.doi.org/10.1007/s13225-011-0117-x