



## A new species of *Pyropia* (Rhodophyta, Bangiaceae), from the Pacific coast of Mexico, based on morphological and molecular evidence

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### Abstract

*Pyropia raulaguilarii* sp. nov. is described from Michoacán, tropical Mexican Pacific, on basis of comparative morphology and nrSSU, *rbcL* sequence analysis. It is distinguished from other *Pyropia* species by the foliose and lanceolate gametophyte, a monoecious thallus and the zygotosporangia in packets of 2x2x4. The phylogenetic analysis showed that the two Pacific Mexican samples, from Caletilla and Carrizalillo (Michoacán), were almost identical and formed a distinctive and well supported clade segregated from other species of *Pyropia* from Brazil, USA and Mexico. The Mexican entity is morphologically and genetically distinct from other *Pyropia* species, suggesting that this species should be assigned to a new taxon.

**Key words:** Bangiales, molecular phylogeny, nrSSU, *rbcL*, marine red algae.

### Introduction

Species of *Porphyra* C.Agardh have few characters for distinguishing species, however, these characters alone have proved to be misleading based on the discovery, using molecular sequences, of many cryptic taxa among species with very similar morphologies (e.g. Brodie & Irvine 1997, Broom *et al.* 2002, 2004, Neefus *et al.* 2002, Teasdale *et al.* 2002, Lindstrom & Fredericq 2003, Niwa *et al.* 2005, Brodie *et al.* 2007, Lindstrom 2008). Almost all of these studies have used either nrSSU or plastid *rbcL* gene. The use of the more conservative nrSSU locus has generally resulted in phylogenies with a well-supported “backbone” but little differentiation of closely related taxa. The protein-coding *rbcL* gene clearly distinguishes species, but support for ancient divergences is often less than the one obtained in analyses based on the nrSSU gene (Lindstrom & Fredericq 2003, Nelson *et al.* 2006).

“*Porphyra sensu lato*” as the cited papers include species no longer considered part of *Porphyra* (*sensu stricto*) (Sutherland *et al.* 2011). Molecular studies have revealed extensive cryptic speciation within the genus *Porphyra* (Broom *et al.* 1999, 2004, 2010, Jones *et al.* 2004) and has been used in the genetics (Kuang *et al.* 1998) and breeding of *Porphyra* species since the end of the twentieth century (Jia *et al.* 2000, Xu *et al.* 2011). Most previous reports were focused on the identification, protection, construction, and classification of different strains (Iitsuka *et al.* 2002, Kunimoto *et al.* 2003, Liu *et al.* 2004, Sun *et al.* 2005, Weng *et al.* 2005, Qiao *et al.* 2007).

Recently, Sutherland *et al.* (2011) carried out an important revision that constitutes a major change in understanding relationships and evolution in Bangiales, and combined analyses of the nuclear SSU rRNA and chloroplast *rbcL* regions recognizing five new genera and two others that were resurrected. The genus *Porphyra* is now restricted to five described species and a number of undescribed species. Other foliose taxa previously placed in *Porphyra* are now recognized to belong to the genera *Boreophyllum* S.C.Lindstr. *et al.*, *Clymene* W.A.Nelson, *Fuscifolium* S.C.Lindstr., *Lysithea* W.A.Nelson, *Miuraea* Kukuchi *et al.*, *Pyropia* J.Agardh and *Wildemania* De Toni.

The genus *Pyropia* is characterized by a) gametophytic thalli monostromatic, linear, ovate, orbicular or funnel shaped, b) margins entire or dentate, planar, undulate, or ruffled, c) color variable, d) blades sessile or with brief stipe, e) vegetative cells embedded in colorless, firm gelatinous matrix, f) cells with one or two stellate chloroplasts, each one with central pyrenoid, g) plants monoecious or dioecious, h) sexual thalli monoecious with groups of cells forming either spermatangia and zygotosporangia, these often in streaks or rectangular patches, or divided into separate male and female sectors by a vertical or horizontal line, i) in dioecious thalli, the spermatangia and zygotosporangia forming as continuous areas along the margins of the blade, and j) the conchocelis phase is a microscopic and filamentous stage (Sutherland *et al.* 2011).

During recent research on the diversity of benthic marine algae of the state of Michoacán, tropical Pacific Mexico coast, an undescribed red alga was collected. This entity is characterized by thalli with monostromatic blades, arising from a single discoid attachment, and sexual thalli monoecious with spermatangia and zygotosporangia in patches. This internal organization corresponds to the genus *Pyropia*.

## Materials & Methods

**Morphological analysis.** Thalli of *Pyropia* were observed at two localities in southern Michoacán: Caletilla and El Carrizalillo. Slides of fertile specimens were prepared for microscopic observation from material preserved in 5% formalin-seawater. Whole mount slides with cross and transverse sections were made by hand with razor blades, these sections were stained in 1% aqueous aniline blue, acidified by the addition of 1N HCl, according to Mateo-Cid *et al.* (2005) and permanently mounted on microscope slides with 70% Karo Syrup® with 1% phenol added as a preservative. Photomicrographs were taken with a Sony MPEGMOVIEVX (Tokyo, Japan) coupled to an Olympus CX31 Microscope (Manila, Filipinas). Images of specimens were taken with a digital SONY camera (MPEGMOVIEVX). Digital images were edited and assembled on plates using Adobe Photoshop 7.0 (San Jose, CA, USA).

**Molecular analysis.** Samples used for molecular analysis were dried in silica gel. Total DNA was extracted using the DNeasy Plant Mini Kit (Qiagen, Valencia, CA, USA) following the manufacturer's instructions. Sequences of the nrSSU and the *rbcL* genes from Bangiales taxa were obtained. The specimens sequenced in this study, plus other available sequences used for the phylogenetic analysis, are shown in Table 1, including their accession numbers in the National Center for Biotechnology Information (NCBI) GenBank data base.

The *rbcL* and nrSSU regions were amplified with primers suggested by Freshwater & Rueness (1994) and Saunders *et al.* (1996), respectively, by using the Taq PCR Core Kit (Quiagen). All polymerase chain reaction (PCR) products were electrophoresed on 1% agarose gel to verify product size. PCR products were purified with Qiagen QIAquick Purification Kit following the manufacturer's instructions. Sequencing was carried out using PCR primers with the BigDye Terminator Cycle Sequencing Reaction Kit (Applied Biosystems, NJ, USA) on an ABI PRISM 3100 Genetic Analyzer (Applied Biosystems). The authors performed the nucleotide BLAST (Basic Local Alignment Search Tool) of the sequences with the program implemented in the NCBI (National Center for Biotechnology Information) home page. Sequences editing was performed with the Sequence Navigator (Applied Biosystems) and aligned according to the CLUSTAL algorithm (Thompson *et al.* 1994).

**Phylogenetic analysis.** Three data sets were constructed -nrSSU data set, the *rbcl* data set, and a combined data set- with the genes concatenated. Outgroup species (*Dione arcuata* W.A.Nelson, *Minerva aenigmata* W.A.Nelson and *Miuraea migitae* (N. Kikuchi *et al.*) N. Kikuchi *et al.*) were selected because of a close phylogenetic relationship with the ingroup (Sutherland *et al.* 2011).

Phylogenetic relationships were inferred with PAUP\* 4.0b10 (Swofford 2002) and MrBayes v.3.0 beta 4 (Huelsenbeck & Ronquist 2001). Maximum-parsimony trees (MP) were constructed using the heuristic search option, tree-bisection-reconnection branch swapping algorithm, unordered and unweight characters, and gaps treated as missing data. Furthermore, distance analysis was conducted under Neighbour-Joining (NJ) algorithm.

Support values for the relationships discovered in analyses were calculated by performing bootstrap analyses (Felsenstein 1985) as implemented in PAUP\*. Twenty thousand heuristic search replicates were executed using the TBR branch-swapping algorithm. Consistency (CI) and Homoplasy (HI) indexes were computed to evaluate the level of homoplasy in the most parsimonious tree.

Bayesian trees were constructed for both the single gene data sets and the concatenated data set. The model used was the general-time-reversible model of nucleotide substitution with invariant sites and gamma distributed rates for the variable sites (GTR + I + G) for all three data sets. This model was selected based on maximum likelihood ratio tests implemented by the software Model Test version 3.06 (Posada & Crandall 1998) with a significance level of 0.01. Analyses were started from random trees, and consisted of two runs, each of four chains (one hot and three cold), of 4 million generations for each data set. The stationary level was reached at generation 17000. Therefore, trees saved until generation 16500 were the 'burn in' of the chain, and inferences about the phylogeny were based on those trees sampled after generation 16500. To visualize the burn-in of -LnL or model parameters, we use the software Tracer v1.4 (Rambaut & Drummond 2007). A 50% consensus tree as implemented by PAUP\* was computed after the 'burnin'. The range of *rbcl* divergence values within and among species was calculated using uncorrected 'p' distances using PAUP\*.

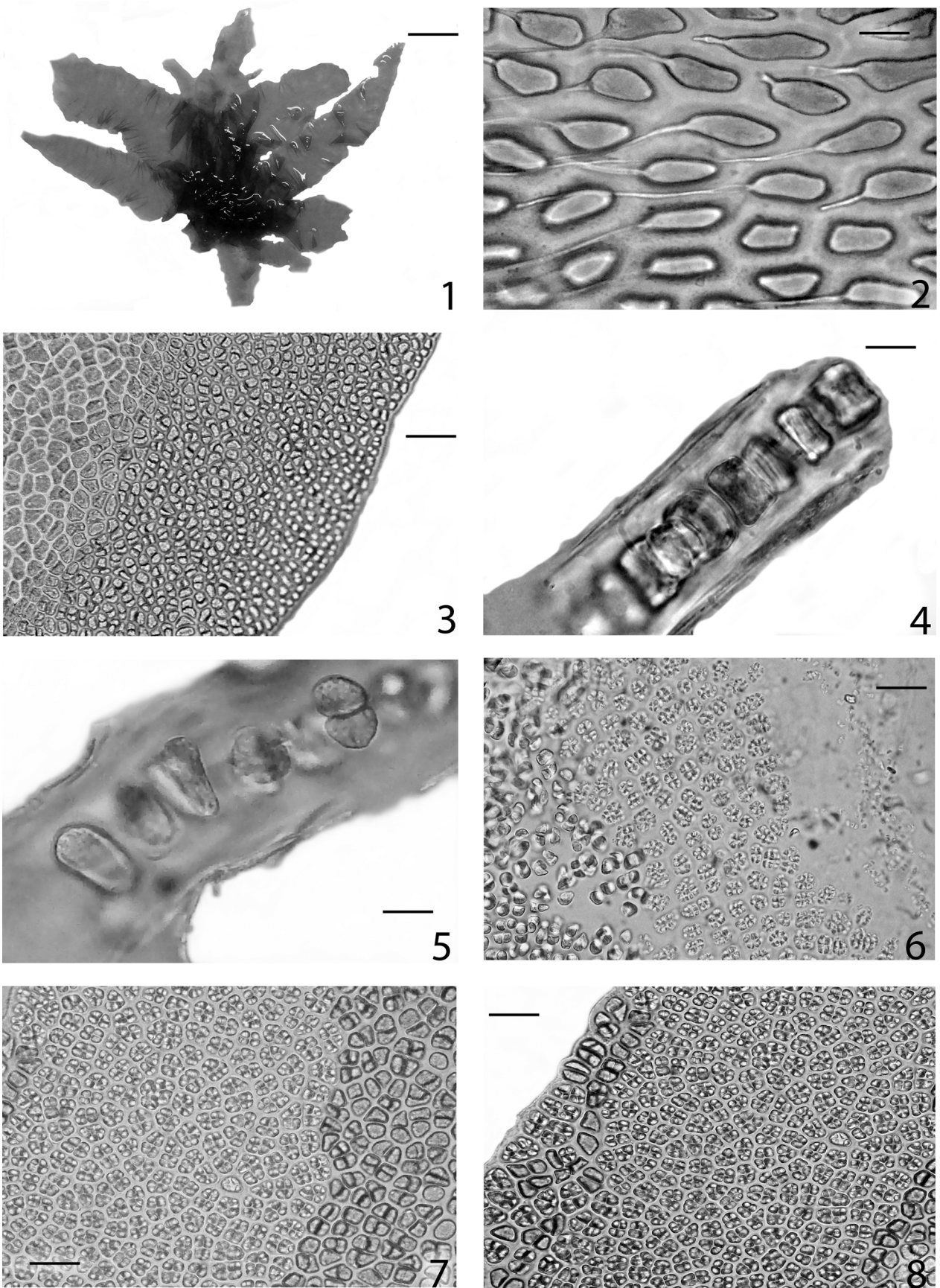
## Results

### *Pyropia raulaguilarii* Mateo-Cid, Mendoza-González & Senties, *sp. nov.* (Figs 1–8, Table 2)

*Gametophytum foliosum, monostromaticum, lanceolatum, basis umbilicalis, margo crispata. Thalli 4 ad 6 cm, adhaerentes perfecte ad papyrus. Thalli steriles usque 40–42 µm crassi. Color rubrum ad aliquando purpureus. Thalli sexuales monoecii spermatangia in viridi inaequalis intermixtam zygotosporangia in rectangulum vel irregularis inaequaliter. Sectiones fertiles (feminei) 50–55 µm crassi. Structurae reproductivae marginales, maculis extensiae intro. Spermatangia in sarcinula 2 × 2 × 4 × 4. Zygotosporangia in sarcinula 2 × 2 × 4.*

**Type:**—MEXICO. Michoacán: Carrizalillo, low deciduous forest, 18°36'00" N, 102°33'03" W, 0 m a.s.l., coll. Mendoza-González and Mateo-Cid, 04-04-2009, *MICH-09-A/01* (holotype: ENCB!, isotype: UAMIZ!, paratype: Michoacán: Caletilla, 18°03'28" N, 102°42'13" W, coll. Mendoza González and Sánchez Heredia, 17-05-2006, *MICH-06-30/01*, ENCB!).

Gametophyte foliose, monostromatic, lanceolate, rarely branched (Fig. 1), often deeply ruffled, margin undulating (Fig. 3). Thalli 4–6 cm high, blades 1–2 cm diameter, red to purple color. Adhering completely to paper. The plants have a well-developed umbilicate base, which is closely adherent to the substratum and attached by rhizoidal cells (Fig. 2). Sterile thalli 40–42 µm thick (Fig. 4), vegetative cells measuring 14–17 µm in diameter and 20–25 µm in length. Vegetative cells with a single stellate plastid with one pyrenoid. Reproductive plants: sexual thalli monoecious, spermatangia in green patches intermixed with zygotosporangia in rectangular or irregular patches in fertile regions of the blades. Fertile female sectors 50–55 µm thick (Fig. 5). Reproductive structures marginal, extending inward in alternating patches. Submarginal tissue paler than the adjacent outer zygotosporangial and inner sterile tissues. Zygotosporangia in packets of 2 × 2 × 4 (Figs 7, 8). Spermatangia in packets of 2 × 2 × 4 × 4 (Fig. 6).

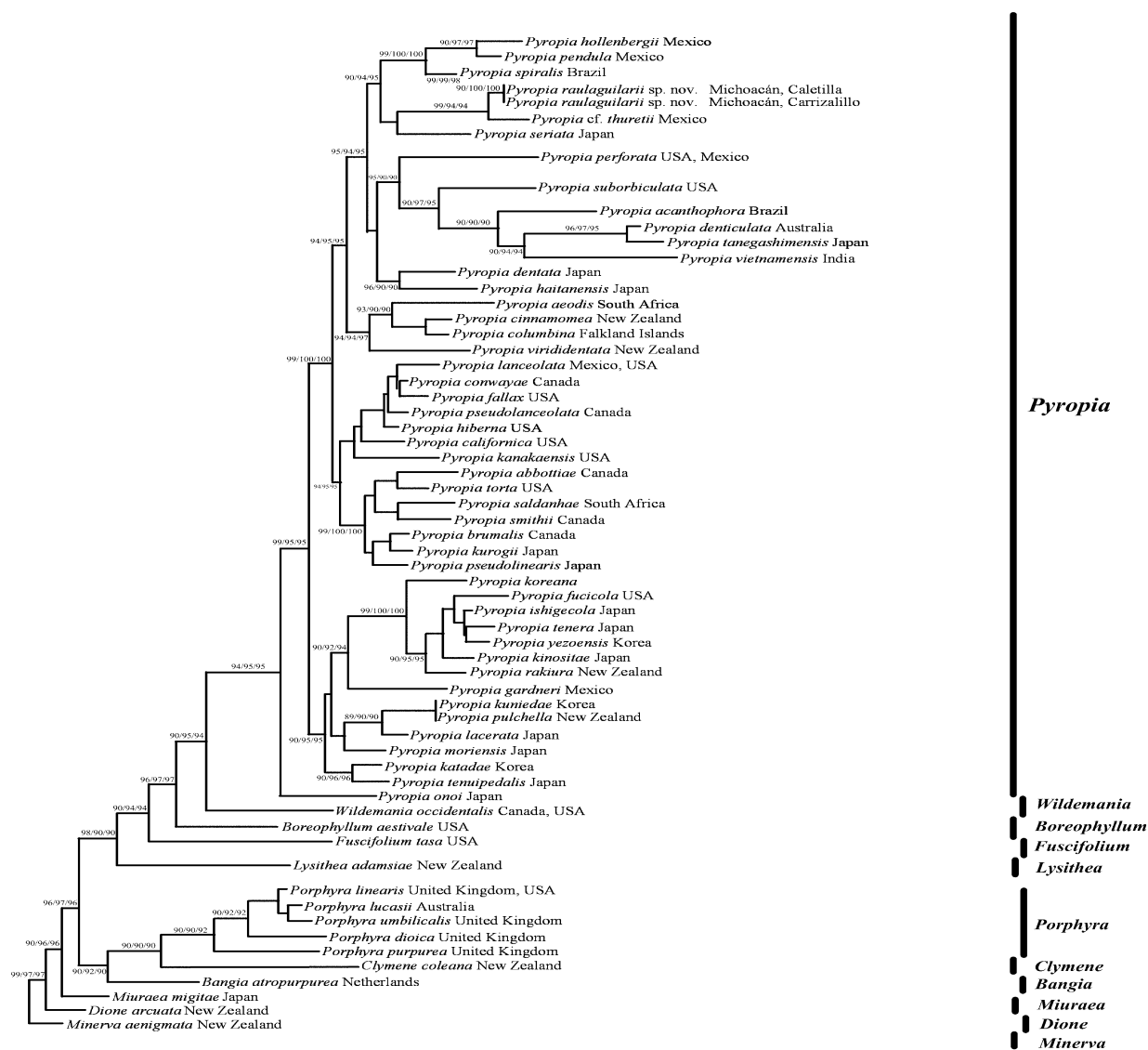


**FIGURES 1–8:** *Pyropia raulaguilarii* sp. nov. Fig. 1: Habit of the foliose gametophyte. Holotype. El Carrizalillo (ENCB 19 603). Fig. 2: Rhizoidal cells in surface view. Fig. 3: Margin of thallus in surface view. Fig. 4: Transverse section through thallus showing monostromatic blade. Fig. 5: Transverse section of female portion. Fig. 6: Mature spermatangia in surface view. Fig. 7: Surface view of zygotosporangial region of thallus. Fig. 8: Mature marginal zygotosporangias in surface view. Scale bars: Fig. 1: 12 mm; Figs 2, 6: 30  $\mu$ m; Figs 3, 7, 8: 40  $\mu$ m; Fig. 4: 15  $\mu$ m; Fig. 5: 20  $\mu$ m.

**Habitat:** —On rocks, intertidal pools.

**Etymology:** —The species epithet is named in honor of Prof. Raúl Aguilar Rosas, a prominent Mexican phycologist and beloved colleague from Facultad de Ciencias Marinas, Universidad Autónoma de Baja California, Mexico.

**Molecular analyses:** —Samples of *Pyropia raulagularii* from Carrizalillo and Caletilla (Michoacán, Mexico) diverged each other 0.02–0.03%. These samples diverged 3.0 to 3.4% from the GenBank sequence called *Pyropia cf. thuretii* from Mazatlán, Mexico (Table 1). The concatenated phylogenetic data set consisted of 62 taxa (Table 1), including 59 ingroup and three outgroup taxa, with 2767 characters: 1300 from the nrSSU and 1467 from the *rbcL* gene. Bayesian, parsimony (not shown) and distance (not shown) analyses resulted in similar trees. The topology of the Bayesian tree showed eight lineage corresponding to representatives of the following Bangiales genera: *Bangia* Lyngbye, *Boreophyllum*, *Clymene*, *Fuscifolium*, *Lysithea*, *Porphyra* C.Agardh, *Pyropia* and *Wildemanina* (Fig. 9). The Pacific Mexican samples processed in the present study were located in two well-supported terminal *Pyropia* sister clades. The samples collected in Carrizalillo and Caletilla (Michoacán) formed a high supported monophyletic clade with a sister clade called *Pyropia cf. thuretii* from Mazatlán, Mexico (Table 1).



**FIGURE 9:** Bayesian result for concatenated nuclear SSU ribosomal RNA (nrSSU) and *rbcL* data set. Support values are Bayesian posterior probabilities, and parsimony and distance bootstrap values, respectively. Some internal support values are omitted for clarity.

**TABLE 1:** Specimen collection information, voucher numbers and GenBank accession numbers of sequences used in phylogenetic analysis.

Taxon	Collection data	GenBank Accession Numbers	
		nrSSU	<i>rbcL</i>
<i>Bangia atropurpurea</i> (Roth) C.Agardh	Netherlands, Ysselmeer, Enschede.	AF169341	AF169330
<i>Boreophyllum aestivale</i> (S.C. Lindstr. & Fredericq) S.C.Lindstr.	USA, Alaska, Amaknak Island, Captains Bay, 4 August 2004, UBC A86208 (nrSSU); SCL 1325 ( <i>rbcL</i> ).	GU319836	EU223033
<i>Clymene coleana</i> (W.A.Nelson) W.A.Nelson	New Zealand, North I, Leigh, WELT A22181 (nrSSU); WELT A22181 ( <i>rbcL</i> ).	AF136423	FJ263672
<i>Dione arcuata</i> <sup>b</sup>	New Zealand, South I, Kaikoura, Ohau Stream, WELT A23126 (nrSSU); WELT A23126 ( <i>rbcL</i> ).	AY465354	EU570052
<i>Fuscifolium tasa</i> (Yendo) S.C.Lindstr.	USA, Alaska, Unalaska Island, Spray Cape, 4 June 2005, UBC A86511 (nrSSU); SCL 12178 ( <i>rbcL</i> ).	GU319862	EU223226
<i>Lysithea adamsiae</i> (W.A.Nelson) W.A.Nelson	New Zealand, Antipodes Islands, Orde Lees I, WELT A023233 (nrSSU); WELT A023233 ( <i>rbcL</i> ).	HQ687566	HQ687515
<i>Minerva aenigmata</i> W.A. Nelson <sup>b</sup>	New Zealand, South Island, Westland, Cobden (nrSSU); New Zealand, North I, Puheke, WELT A25775 ( <i>rbcL</i> ).	AY184347	EU570053
<i>Miuraea migitae</i> <sup>b</sup>	Japan, Osaka, Osaka Bay.	EU521642	EU521643
<i>Porphyra dioica</i> J.Brodie & L.M.Irvine	United Kingdom, Sidmouth, WELT A024417 (nrSSU); WELT A024417 ( <i>rbcL</i> ).	HQ687579	HQ687546
<i>Porphyra linearis</i> Grev.	United Kingdom, Aberystwyth, BM000898797 (nrSSU); USA, ME, Kittery, Seapoint, NHA 61204 ( <i>rbcL</i> ).	HQ687580	AF078745
<i>Porphyra lucasii</i> Levring	Australia, WA, Trigg Beach, WELT A23051 (nrSSU); WELT-A23051 ( <i>rbcL</i> ).	AY139685	AY139687
<i>Porphyra purpurea</i> (Roth) C.Agardh	United Kingdom, Litstock, Somerset, BM000898974 (nrSSU); BM000898974 ( <i>rbcL</i> ).	HQ687567	HQ687516
<i>Porphyra umbilicalis</i> Kütz.	United Kingdom, Sidmouth, WELT A024414 (nrSSU); WELT A024414 ( <i>rbcL</i> ).	HQ687584	HQ687559
<i>Pyropia acanthophora</i> (E.C.Oliveira & Coll) M.C.Oliveira <i>et al.</i>	Brazil, São Paulo, Ubatuba.	L26197	HQ605695
<i>Pyropia abbottiae</i> (Krishnam) S.C.Lindstr.	Canada, BC, Victoria, Harling Point, 25 April 2005, UBC A85149 (nrSSU); SCL 12567 ( <i>rbcL</i> ).	GU319835	EU223024
<i>Pyropia aeodis</i> (N.J.Griffin <i>et al.</i> ) J.E. Sutherland	South Africa, Paternoster.	AY292624	GU165843
<i>Pyropia brumalis</i> (Mumford) S.C. Lindstr.	Canada, BC, Vancouver, Stanley Park, 23 February 1998, UBC A84426 (nrSSU); UBC A84426 ( <i>rbcL</i> ).	GU319837	EU223038
<i>Pyropia californica</i> J.Agardh	USA, AK, Northeast Ushagat Island, 25 August 2006, UBC A86168 (nrSSU); SCL 13101 ( <i>rbcL</i> ).	GU319849	EU223117
<i>Pyropia cf. thuretii</i>	Mexico, Sinaloa, Mazatlán, Olas Altas, WELT A024423 (nrSSU); WELT A024423 ( <i>rbcL</i> ).	HQ687587	HQ687519
<i>Pyropia cinnamomea</i> (W.A.Nelson) W.A.Nelson	New Zealand, South I, Otago, Bruce's Rock.	AH008010	EU521637

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**TABLE 1** (continued)

Taxon	Collection data	GenBank Accession Numbers	
<i>Pyropia columbina</i> (Mont.) W.A. Nelson	Falkland Islands, East Falkland, Top Island, Port Stanley, February 2001, WELT A023553 (nrSSU); WELT A023553 ( <i>rbcL</i> ).	GU046398	GU046423
<i>Pyropia conwayae</i> (S.C.Lindstr. & K.M.Cole) S.C.Lindstr.	Canada, BC, French Beach, 12 March 2007, UBC A86515 (nrSSU); SCL 13109 ( <i>rbcL</i> ).	GU319838	EU223045
<i>Pyropia dentata</i> (Kjellm.) N.Kikuchi & M.Miyata	Japan, Chiba, Shirahama, WELT A024400 (nrSSU); WELT A024400 ( <i>rbcL</i> ).	HQ687588	HQ687520
<i>Pyropia denticulata</i> (Levring) J.A.Phillips & J.E.Sutherland.	Australia, Queensland, Mooloolaba, WELT A023224 (nrSSU); WELT A023224 ( <i>rbcL</i> ).	HQ687570	HQ687521
<i>Pyropia fallax</i> (S.C.Lindstr. & K.M.Cole) S.C. Lindstr.	USA, Alaska, Avatanak Island, 12 June 2008, UBC A87380 (nrSSU); UBC A87380 ( <i>rbcL</i> ).	GU319840	GU319865
<i>Pyropia fucicola</i> (Krishnam. S.C.Lindstr.	USA, Alaska, Japonski Island, Sitka airport, SW side, 29 end of runway, 19 August 2005, UBC A87689 (nrSSU); SCL 12715 ( <i>rbcL</i> ).	GU319841	EU223088
<i>Pyropia gardneri</i>	Mexico, Baja California, Caminitos, Cabo Punta Banda, CMMEX 4355 (nrSSU); WELT A024420 ( <i>rbcL</i> ).	DQ084423	HQ687522
<i>Pyropia haitanensis</i>	Japan, Ehime, Yuge.	AB013181	AB118585
<i>Pyropia hiberna</i> (S.C.Lindstr. & K.M.Cole) S.C.Lindstr.	USA, CA, Monterey County, Pacific Grove, 31 December 2007, UBC A87474 (nrSSU); UBC A87474 ( <i>rbcL</i> ).	GU319843	GU319866
<i>Pyropia hollenbergii</i>	Mexico, Baja California Sur, Bahía Agua Verde, WELT A023231 (nrSSU).	HQ687589	AY794401
<i>Pyropia ishigeicola</i> (A.Miura) N.Kikuchi & M.Miyata	Japan, Chiba, Katsuura, Yoshio, WELT A023199 (nrSSU); WELT A023199 ( <i>rbcL</i> ).	HQ687571	HQ687524
<i>Pyropia kanakaensis</i> (Mumford) S.C.Lindstr.	USA, WA, Olympic Peninsula, between Makah Bay and Sekui, 31 May 2003, UBC A86491 (nrSSU); SCL 10932 ( <i>rbcL</i> ).	GU319844	EU223099
<i>Pyropia katadae</i> (A. Miura) M.S. Hwang <i>et al.</i>	Korea, Gyeongsangbukdo, Pohang.	HQ728191	HQ728199
<i>Pyropia koreana</i> (M.S.Hwang & I.K.Lee) M.S.Hwang <i>et al.</i>	-	HQ728190	HQ728198
<i>Pyropia kinositae</i> (Yamada & Tak.Tanaka) N.Kikuchi <i>et al.</i>	Japan, Hokkaido, Suttu, Arito.	EU521640	EU521641
<i>Pyropia kuniedae</i> (Kurogi) M.S.Hwang & H.G.Choi	Korea, Sachon, Gyeongsangnamdo, Namhae.	HQ728192	HQ728200
<i>Pyropia kurogii</i> (S.C.Lindstr.) S.C.Lindstr.	Japan, Hokkaido, Utoro, WELT A023201 (nrSSU); WELT A023201 ( <i>rbcL</i> ).	HQ687573	HQ687526
<i>Pyropia lacerata</i> (A.Miura) N.Kikuchi & M.Miyata	Japan, Chiba, Shirahama, WELT A024399 (nrSSU); WELT A024399 ( <i>rbcL</i> ).	HQ687574	HQ687527
<i>Pyropia lanceolata</i>	Mexico, Baja California, Punta Popotla (nrSSU); USA, California, Monterey County, Spanish Bay, 1 January 2008, UBC A87475 ( <i>rbcL</i> ).	AY909594	GU319867
<i>Pyropia leucosticta</i>	United Kingdom, Sidmouth, BM000898751 (nrSSU).	HQ687593	DQ191359

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**TABLE 1** (continued)

Taxon	Collection data	GenBank Accession Numbers	
<i>Pyropia moriensis</i> (Ohmi) N.Kikuchi & M.Miyata	Japan, Hokkaido, Otaru.	EU521644	EU521645
<i>Pyropia onoi</i> (Ueda) N.Kikuchi & M.Miyata	Japan, Hokkaido, Mori, WELT A023216 (nrSSU); WELT A023216 ( <i>rbcL</i> ).	HQ687575	HQ687529
<i>Pyropia pendula</i>	Mexico, Baja California Sur, Calerita, CMMEX 4437 (nrSSU); CMMEX 4437 ( <i>rbcL</i> ).	DQ084430	HQ687530
<i>Pyropia perforata</i>	USA, California, Del Norte County, Crescent City Lighthouse, 24 May 2002 (nrSSU); Mexico, Baja California, Punta Popotla, 24 February 1999 ( <i>rbcL</i> ).	GU319856	GU046416
<i>Pyropia pseudolanceolata</i> (Krishnam) S.C.Lindstr.	Canada, BC, Victoria, Harling Point, 5 February 2005, UBC A87696 (nrSSU); SCL 12538 ( <i>rbcL</i> ).	GU319857	EU223145
<i>Pyropia pseudolinearis</i> (Ueda) N.Kikuchi <i>et al.</i>	Japan, Chiba, Choshi, WELT A024424 (nrSSU); WELT A024424 ( <i>rbcL</i> ).	HQ687590	HQ687531
<i>Pyropia pulchella</i> (Ackland <i>et al.</i> ) T.J.Farr & J.E.Sutherland.	New Zealand, North I, Waihou Bay East, WELT A024398 (nrSSU); WELT A024398 ( <i>rbcL</i> ).	HQ687591	HQ687532
<i>Pyropia rakiura</i> (W.A.Nelson) W.A.Nelson	New Zealand, South I, Kaikoura, Ocean View.	AF136425	EU521646
<i>Pyropia saldanhae</i> (Stegenga <i>et al.</i> ) J.E.Sutherland	South Africa, Kommetjie.	AY292630	GU165838
<i>Pyropia seriata</i> (Kjellm.) N.Kikuchi & M.Miyata	Japan, Kumamoto, Hondo, WELT A023206 (nrSSU); WELT A023206 ( <i>rbcL</i> ).	HQ687576	HQ687533
<i>Pyropia smithii</i> (Hollenberg & I.A.Abbott) S.C.Lindstr.	Canada, BC, Vancouver Island, Barkley Sound, Nudibranch Point, 10 August 2002, UBC A85584 (nrSSU); UBC A85585 ( <i>rbcL</i> ).	GU319861	EU223224
<i>Pyropia spiralis</i> (E.C.Oliveira & Coll) M.C.Oliveira <i>et al.</i>	Brazil, Espírito Santo, Vila Velha.	AY766360	HQ605696
<i>Pyropia suborbiculata</i>	USA, NC, Mansonboro Island, 20 May 1996.	HQ728193	AY028523
<i>Pyropia tanegashimensis</i> (Shimura) N.Kikuchi & E.Fujiyoshi	Japan, Kagoshima, Tanegashima, Iseki, WELT A023236 (nrSSU); WELT A023236 ( <i>rbcL</i> ).	HQ727887	HQ687542
<i>Pyropia tenera</i> (Kjellm.) N.Kikuchi <i>et al.</i>	Japan, Kumamoto, Kawaura, CMNH-BA-1354 (nrSSU); CMNH-BA-1354 ( <i>rbcL</i> ).	HQ687577	HQ687543
<i>Pyropia tenuipedalis</i> (A.Miura) N.Kikuchi & M.Miyata	Japan, Chiba, Urayasu.	EU521648	EU521649
<i>Pyropia torta</i> (V.Krishnam) S.C.Lindstr.	USA, WA, San Juan Island, Vista Mar, 22 January 2002, SCL 13289 (nrSSU); SCL 13289 ( <i>rbcL</i> ).	GU319863	EU223236
<i>Pyropia vietnamensis</i> (Takido <i>et al.</i> ) J.E.Sutherland & Monotilla	India, Kerala, Thangeseri, WELT A024426 (nrSSU); WELT A024426 ( <i>rbcL</i> ).	HQ687578	HQ687544
<i>Pyropia virididentata</i> (W.A.Nelson) W.A.Nelson	New Zealand, Wellington, North I, Lyall Bay.	AF136421	EU521650
<i>Pyropia yezoensis</i>	Korea, Gyeongsangnamdo, Tongyoung, Galmok.	HQ728189	HQ728197
<i>Pyropia raulaguilarii</i> sp. nov. <sup>a</sup>	Mexico, Michoacán, Caletilla, 6 March 2008, ENCB 19524.	JQ684704	JQ684700
<i>Pyropia raulaguilarii</i> sp. nov. <sup>a</sup>	Mexico, Michoacán, Carrizalillo, 4 April 2009, ENCB 19523.	JQ684705	JQ684701

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**TABLE 1** (continued)

Taxon	Collection data	GenBank Accession Numbers
<i>Wildemanian occidentalis</i> (Setch. & Hus) S.C.Lindstr.	Canada, Vancouver Island British Columbia, Cape Palmerston, 17 May 2003, UBC A86488 (nrSSU); USA, Washington, Clallam County, Olympic Peninsula, between Makah Bay and Sekiu, 31 May 2003, SCL 10935 ( <i>rbcL</i> ).	GU319851 EU223119

<sup>a</sup>Sequences obtained in this work. <sup>b</sup>Outgroups.

**TABLE 2:** Comparative vegetative and reproductive characteristics of *Pyropia* species.

Character	<i>P. hollenbergii</i> <sup>1</sup>	<i>P. pendula</i> <sup>1</sup>	<i>P. thuretii</i> <sup>2</sup>	<i>P. raulaguilarii</i>
Height (gametophyte thallus)	15–22 cm	25 cm	4–17	4–6 cm
Shape of the blade	Linear-lanceolate	Linear-ligulate	lanceolate	lanceolate
Width of the blade	1–7.5 cm	0.1–2	2–8 cm	1–2 cm
Thalli thickness	45–60 µm	40–54 µm	20–40 µm	40–45 µm
Vegetative cells size Diameter/length	8–14/12–30 µm	6–14/8–25 µm	4–10/8–20 µm	14–17/20–22 µm
Sexuality	Dioecius	Dioecius	Monoecius	Monoecius
Packets of spermatangia	64	128	64	64
Packets of Zygotosporangia	8	8	8	16

<sup>1</sup>Aguilar-Rosas *et al.* (2007). <sup>2</sup>Aguilar-Rosas & Aguilar-Rosas (2003).

## Discussion

In the Pacific coast of Mexico there are seven species of *Pyropia* (Aguilar-Rosas *et al.*, 2007, as *Porphyra*): *Py. gardneri* (G.M.Sm. & Hollenb.) S.C.Lindstr., *Py. hollenbergii* (E.Y.Dawson) J.E.Sutherland *et al.*, *Py. lanceolata* (Setch. & Hus) S.C.Lindstr., *Py. pendula* (E.Y.Dawson) J.E.Sutherland *et al.*, *Py. perforata* (J. Agardh) S.C.Lindstr., *Py. suborbiculata* (Kjellm.) J.E.Sutherland *et al.*, and *Py. thuretii* (Setch. & E.Y.Dawson) J.E.Sutherland *et al.* Among them, *Py. hollenbergii* and *Py. pendula* are considered endemic to the Gulf of California, while the remaining five are widely distributed from British Columbia, Canada, to Peru. According to Aguilar-Rosas *et al.* (2007), the thalli of *Py. hollenbergii*, *Py. pendula* and *Py. lanceolata* have a certain morphological and reproductive similarity. However, characters such as the packet number of spermatangia and zygotosporangia, and the length of the thallus, allows to separate these species. Additionally, *Py. thuretii* is widely distributed to British Columbia, Canada to Pacific Northwest coast of Mexico.

Regarding *Py. lanceolata*, it is distributed from the U.S.A. to temperate waters of the Pacific coast of Mexico, it has lanceolate thallus and being over 50 cm length. *Py. perforata* grows in dense groups, up to 1 m length, and wide blades lobed or divided and often located from Alaska to Baja California.

*Pyropia raulaguilarii* is most closely related with *Py. hollenbergii* and *Py. pendula*, morphological and reproductive differences between these species are shown on Table 2. The most distinctive features of *Pyropia raulaguilarii* are the length of the blade, the surface view in the spermatangial portions and the division formulas of the zygotosporangia (Table 2). In *Py. thuretii* and *Py. hollenbergii* both species exhibit differences in blade thickness and formulas of zygotosporangia division; whereas in *Py. raulaguilarii* there are 16 in *Py. thuretii* there are 32. On the other hand, Aguilar-Rosas & Aguilar-Rosas (2003) and Aguilar-Rosas *et al.* (2007) examined several species of *Porphyra* (*Pyropia*) from the Pacific coast of Mexico. Their examination revealed that there are other *Pyropia* species that may be confused with *Py. raulaguilarii* mentioned here. Table 2 shows the vegetative and reproductive characters of various species of *Pyropia* and in it observed the

main differences between species. For example, the gametophyte thallus of *Py. raulaguilarii* reaches a size of 4.0–6.0 cm, while comparatively *Py. hollenbergii* and *Py. pendula* reaches up to 25 cm high and both are dioecious. Furthermore, *Py. raulaguilarii*, is distributed in the Tropical Pacific of Mexico.

The interspecific divergence obtained for the *Pyropia* Pacific Mexican taxa was comparable to the amount of variation observed in other Bangiales taxa. Broom *et al.* (2010) recorded *rbcL* interspecific divergence values between 1.15% and 9.88%, and recorded nrSSU interspecific divergence values between 1.84% and 8.39% for *Porphyra* species recorded in the Falkland Islands. Lindstrom (2008) reported *rbcL* interspecific values between 1% and 4.7% for *Porphyra* species from Pacific region. Brodie *et al.* (2007) obtained a pairwise sequence divergence among *Py. rosenfurtii*, *Py. olivii* (Orfanidis *et al.*) J.Brodie & Neefus and *Py. leucosticta* (Thur.) Neefus & J.Brodie in their *rbcL* alignment that ranged from 1% to 11%. Xu *et al.* (2011) mentioned that *Py. yezoensis* (Ueda) M.S.Hwang & H.G.Choi diverged from *Py. haitanensis* (T.J.Chang & B.F.Zheng) N.Kikuchi & M.Miyata by 1% of *rbcL* genetic divergence value. Several studies applying molecular techniques to identify species in *Porphyra* (Stiller & Waaland 1993, Oliveira *et al.* 1995, Broom *et al.* 1999, Kunimoto *et al.* 1999) have revealed considerable interspecies diversity of the nrSSU and *rbcL* regions within this genus and suggest that provides useful characters for species identification.

The molecular phylogeny showed that samples from Carrizalillo and Caletilla (Michoacán) populations are almost identical, confirming that these samples constitute the same taxonomic entity. The phylogenetic analysis also showed that the mentioned Mexican Pacific samples represent a distinctive and well-supported clade segregated from the rest of the *Pyropia* representatives.

The high level of genetic variation observed between the Pacific Mexican samples (from Michoacán and Baja California Sur) and other *Pyropia* species support the recognition of these two taxonomic entities as new taxa within the genus *Pyropia*. Neefus *et al.* (2002) considered that Bangiales is understudied and that the number of species could be greater than those recognized at the present.

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## References

- Aguilar-Rosas, L.E. & Aguilar-Rosas, R. (2003). El género *Porphyra* (Bangiaceae, Rhodophyta) en la costa Pacífico de México. *Hidrobiológica* 13: 159–164.
- Aguilar-Rosas, R., Aguilar-Rosas, L.E., Sánchez-Rodríguez, I., Broom, J.E. & Nelson, W.A. (2007). Morfología y distribución de *Porphyra hollenbergii* (Bangiaceae, Rhodophyta) en la costa del Pacífico de México. *Revista Mexicana de Biodiversidad* 78: 351–357.
- Brodie, J. & Irvine, L.M. (1997). A comparison of *Porphyra dioica* sp. nov. and *P. purpurea* (Roth) C. Ag. (Rhodophyta: Bangiophycidae) in Europe. *Cryptogamie Algologie* 18: 283–297.
- Brodie, J., Bartsch, I., Neefus, C., Orfanidis, S., Bray, T. & Mathieson, A.C. (2007). New insights into the cryptic diversity of the North Atlantic-Mediterranean '*Porphyra leucosticta*' complex: *P. olivii* sp. nov. and *P. rosenfurtii* (Bangiales, Rhodophyta). *European Journal of Phycology* 42: 3–28. DOI: 10.1080/09670260601043946
- Broom, J.E.S., Farr, T.J. & Nelson, W.A. (2004). Phylogeny of the *Bangia* flora of New Zealand suggests a southern origin for *Porphyra* and *Bangia* (Bangiales, Rhodophyta). *Molecular Phylogenetics and Evolution* 31: 1197–2007.
- Broom, J.E., Jones, W.A., Hill, D.F., Knight, G.A. & Nelson, W.A. (1999). Species recognition in New Zealand *Porphyra* using 18S rDNA sequencing. *Journal of Applied Phycology* 11 (2): 421–428. DOI: 10.1023/A: 1008162825908
- Broom, J.E.S., Nelson, E., Farr, W.A., Phillips, T.J. & Clayton, M. (2010). Relationships of the *Porphyra* (Bangiales, Rhodophyta) flora of the Falkland Islands: a molecular survey using *rbcL* and nrSSU sequence data. *Australian*

- Broom, J.E., Nelson, W.A., Yarish, C., Jones, W.A., Aguilar-Rosas, R. & Aguilar-Rosas, L.E. (2002). A reassessment of the taxonomic status of *Porphyra suborbiculata*, *Porphyra carolinensis* and *Porphyra lilliputiana* (Bangiales, Rhodophyta) based on molecular and morphological data. *European Journal of Phycology* 37: 227–235. DOI: 10.1017/S0967026202003566
- Felsenstein, J. (1985). Confidence limits on phylogenies: an approach using the bootstrap. *Evolution* 39: 783–791.
- Freshwater, D.W. & Rueness, J. (1994). Phylogenetic relationships of some European *Gelidium* (Gelidiales, Rhodophyta) species, based on rbcL nucleotide sequence analysis. *Phycologia* 33: 187–194.
- Huelsenbeck, J.P. & Ronquist, F.R. (2001). MRBAYES: Bayesian inference of phylogeny. *Bioinformatics* 17: 754–755. DOI:10.1093/bioinformatics/17.8.754
- Jia, J.H., Wang, P., Jin, D.M., Qu, X.P., Wang, Q., Li, C.Y., Weng, M.L. & Wang, B. (2000). The amplification of RAPD markers in diversity detection and variety identification of *Porphyra*. *Acta Botanica Sinica* 42: 403–407.
- Jones, W.A., Griffin, N.J., Jones, D.T., Nelson, W.A., Farr, T.J. & Broom, J.E. (2004). Phylogenetic diversity in South African *Porphyra* (Bangiales, Rhodophyta) determined by nuclear SSU sequence analysis. *European Journal of Phycology* 39: 197–211. DOI: 10.1080/0967026042000202145
- Kuang, M., Wang, S.J., Li, Y., Shen, D.L. & Tseng, C.K. (1998). RAPD study on some common species of *Porphyra* in China. *Chinese Journal Oceanology and Limnology* 16 : 140–146.
- Kunimoto, M., Kito, H., Mizukami, Y., Murase, N. & Levine I. (2003). Molecular features of a defined genetic marker for the determination of the *Porphyra tenera* lineage. *Journal of Applied Phycology* 15: 337–343. DOI: 10.1023/A:1025182508440
- Kunimoto, M., Kito, H., Yamamoto, Y., Cheney, D.P., Kaminishi, Y. & Mizukami, Y. (1999). Discrimination of *Porphyra* species based on small subunit ribosomal RNA gene sequences. *Journal of Applied Phycology* 11: 203–209. DOI: 10.1023/A:1008019409008
- Lindstrom, S.C. (2008). Cryptic diversity, biogeography and genetic variation in northeast Pacific species of *Porphyra* sensu lato (Rhodophyta, Bangiales). *Journal of Applied Phycology* 20: 951–962. DOI: 10.1007/s10811-008-9313-9
- Lindstrom, S.C. & Fredericq, S. (2003). rbcL gene sequences reveal relationships among north-east Pacific species of *Porphyra* (Bangiales, Rhodophyta) and a new species, *P. aestivalis*. *Phycological Research* 51: 211–224. DOI: 10.1046/j.1440-1835.2003.00312.x
- Iitsuka, O., Nakamura, K., Ozaki, A., Okamoto, N. & Saga, N. (2002). Genetic information of three pure lines of *Porphyra yezoensis* (Bangiales, Rhodophyta) obtained by AFLP analysis. *Fisheries Sciences* 68: 1113–1117.
- Liu, H.Q., Yu, W.G., Dai, J.X., Gong, Q.H., Shi, X.C. & Yang, K.F. (2004). Construction of *Porphyra yezoensis* pure line from protoplasts and its 18S rDNA sequences determination. *Journal of Ocean University of China* 3: 60–64.
- Mateo-Cid, L.E., Mendoza-González, A.C., Gavio, B. & Fredericq, S. (2005). *Grateloupia huertana* sp. nov. (Halymeniaceae, Rhodophyta): a peculiar new prostrate species from tropical Pacific Mexico. *Phycologia* 44: 4–16.
- Neefus, C.D., Mathieson, A.C., Klein, A.S., Teasdale, B., Bray, T. & Yarish, C. (2002). *Porphyra birdiae* sp. nov. (Bangiales, Rhodophyta): a new species from the northwest Atlantic. *Algae* 17: 203–216.
- Nelson, W.A., Farr, T.J. & Broom, J.E.S. (2006). Phylogenetic relationships and generic concepts in the red order Bangiales: challenges ahead. *Phycologia* 45: 249–259.
- Niwa, K., Kikuchi, N. & Aruga, Y. (2005). Morphological and molecular analysis of the endangered species *Porphyra tenera* (Bangiales, Rhodophyta). *Journal of Phycology* 41: 294–304. DOI: 10.1111/j.1529-8817.2005.04039.x
- Oliveira, M.C., Kurniawan, J., Bird, J.C., Rice, E.L., Murphy, C.A., Sungh, R.K., Gutell & Ragan, M.A. (1995). A preliminary investigation of the order Bangiales (Bangiophycidae, Rhodophyta) based on sequences of nuclear small-subunit ribosomal RNA genes. *Phycological Research* 43: 71–79. DOI: 10.1111/j.1440-1835.1995.tb00007.x
- Posada, D. & Crandall, K.A. (1998). Model test: testing the model of DNA substitution. *Bioinformatics* 14: 817–818. DOI:10.1093/bioinformatics/14.9.817
- Qiao, L.X., Weng, M.L., Kong, F.N., Dai, J.X. & Wang, B. (2007). The application of RSAP marker technique in diversity detection and germplasm identification of *Porphyra*. *Journal of Ocean University of China* 37: 951–956.
- Rambaut, A. & Drummond, A.J. (2007). *Tracer v1.4*. Available from <http://beast.bio.ed.ac.uk/Tracer> (accessed 25 April 2012).
- Saunders, G.W., Strachan, I.M., West, J.A. & Kraft, G.T. (1996). Nuclear small-subunit ribosomal RNA gene sequences from representatives Ceramiaceae (Ceramiiales, Rhodophyta). *European Journal of Phycology* 31: 23–29. DOI: 10.1080/09670269600651151
- Sun, J.W., Jin, D.M., Zhou, C.J., Yang, Q.K., Weng, M.L., Duan, D.L. Xu, P., Ma, J.H. & Wang, B. (2005). Identification of *Porphyra* lines (Rhodophyta) by AFLP DNA fingerprinting and molecular markers. *Plant Molecular Biology Reporter* 23: 251–262.
- Sutherland, J., Lindstrom, S., Nelson, W., Brodie, J., Lynch, M., Hwang, M., Choi, H., Miyata, M., Kikuchi, N., Oliveira, M.C., Farr, T. Neefus, C., Mols-Mortensen, A., Milstein, J. & Miller, K. (2011). A new look at an ancient order: generic revision of the Bangiales. *Journal of Phycology* 47: 1131–1151. DOI: 10.1111/j.1529-8817.2011.01052.x
- Stiller, J.W. & Waaland, J.R. (1993). Molecular analysis reveals cryptic diversity in *Porphyra* (Rhodophyta). *Journal of*

- Phycology* 29: 506–517. DOI: 10.1111/j.1529-8817.1993.tb00152.x
- Swofford, D.L. (2002). *PAUP\*: Phylogenetic Analysis Using Parsimony (\*and Other Methods), Version 4*. Sinauer Associates, Sunderland.
- Teasdale, B., West, A., Taylor, H. & Klein, A. (2002). A simple restriction fragment length polymorphism (RFLP) assay to discriminate common *Porphyra* (Bangiophyceae, Rhodophyta) taxa from the Northwest Atlantic. *Journal of Applied Phycology* 14: 293–298. DOI: 10.1023/A:1021180315743
- Thompson, J.D., Higgins, D.G. & Gibson, T.J. (1994). CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, positions-specific gap penalties and weight matrix choice. *Nucleic Acids Research* 22: 4673–4680.
- Weng, M.L., Liu, B., Jin, D.M., Yang, Q.K., Zhao, G., Ma, J.H., Xu, P., Duan, D.L. & Wang, B. (2005). Identification of 27 *Porphyra* lines (Rhodophyta) by DNA fingerprinting and molecular markers. *Journal of Applied Phycology* 17 : 91–97. DOI: 10.1007/s10811-005-4845-8
- Xu, P., Yang, I., Zhu, J., Xu, H. & Lu, Q. (2011). Analysis of hybridization strains of *Porphyra* based on *rbcL* gene sequences. *Journal of Applied Phycology* 23: 235–241. DOI: 10.1007/s10811-010-9537-3